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Review Article

A Comprehensive Review on Taxonomy, Ethnobotany, Phytochemistry and Pharmacological Potential

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ABSTRACT

Hedychium spicatum Buch. -Ham. ex D.Don (family Zingiberaceae), commonly known as Kapur Kachri or Spiked Ginger Lily, is a rhizomatous perennial herb of significant ethnomedicinal importance indigenous to the Himalayan biodiversity hotspot. Distributed across altitudes of 800–3,000 metres in India, Nepal, Bhutan, and parts of Southeast Asia, the plant has been systematically employed in Ayurveda, Siddha, and tribal folk medicine for millennia. Its therapeutic applications span respiratory disorders, inflammatory conditions, gastrointestinal ailments, pain syndromes, hepatic dysfunction, and infectious diseases. This review comprehensively synthesizes current scientific literature encompassing its taxonomic identity, botanical morphology, phytochemical constituents, detailed pharmacological mechanisms, industrial applications, and conservation concerns. Phytochemical profiling of the rhizome reveals a structurally diverse array of secondary metabolites including labdane diterpenes (hedychinone, spicatanol), monoterpenoids (1,8-cineole, camphene, α -pinene), sesquiterpenoids, flavonoids (quercetin, kaempferol), phenolic acids, and alkaloids. These constituents confer validated pharmacological activities encompassing anti-inflammatory, antioxidant, antimicrobial, anticancer, hepatoprotective, analgesic, anti-asthmatic, immunomodulatory, and neuroprotective properties. Despite substantial pre-clinical evidence, conspicuous lacunae remain in clinical validation, pharmacokinetic characterization, and quality standardization. This review identifies key research gaps and proposes strategic directions for translating the ethnomedicinal legacy of *H. spicatum* into evidence-based therapeutics.

INTRODUCTION

The global resurgence of interest in plant-derived medicine has been propelled by the twin imperatives of antimicrobial resistance and the

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high attrition rate of synthetic drug candidates in clinical development. Ethnobotanical knowledge systems, refined over generations of empirical observation, provide an irreplaceable scaffold for modern drug discovery. The World Health Organization (WHO) estimates that over 80% of the global population continues to rely on traditional plant medicine as a primary healthcare modality, necessitating rigorous scientific validation of therapeutic claims (WHO, 2019) [1]. The Zingiberaceae, or ginger family, represents one of the most pharmacologically productive plant families, contributing commercially significant medicines, spices, and aromatic products. The genus *Hedychium* J. Koenig, comprising approximately 50–70 species distributed across tropical and subtropical Asia, is distinguished by its ornamental and medicinal value. *Hedychium spicatum* Buch.-Ham. ex D. Don, first described by Buchanan-Hamilton and later revised by Don in his seminal 1825 work *Prodromus Florae Nepalensis*, occupies a position of particular prominence within the genus [2]. Locally designated Kapur Kachri (Sanskrit: Shati, Hindi: Kapoor Kachri), *H. spicatum* is deeply embedded in Ayurvedic pharmacopoeia. The *Ashtanga Hridayam* and *Charaka Samhita* cite its rhizome, referred to as Shati, as a component of formulations targeting kapha and vata vitiation, particularly for respiratory and neurological conditions [3]. Contemporary phytochemical investigations have substantially corroborated these traditional applications, revealing a complex and bioactivity-rich secondary metabolite profile.

The rhizome, constituting the primary medicinal organ, yields 1.5–4.0% essential oil on a dry weight basis, dominated by oxygenated monoterpenes and labdane-type diterpenes. Hedychinone, a principal labdane diterpene, has emerged as a structurally unique bioactive scaffold with demonstrated anti-inflammatory, anticancer, and antioxidant properties [4]. Despite this scientific momentum, *H. spicatum* remains underutilized in modern therapeutics, partly owing to insufficient clinical data and lack of standardized formulations. This review aims to consolidate, critically appraise, and synthesize the accumulated scientific literature on *H. spicatum* spanning taxonomy, ethnobotany, phytochemistry, pharmacology, toxicology, and industrial applications. Special emphasis is placed on the molecular mechanisms underlying reported pharmacological activities and on identifying specific knowledge gaps that impede translational development.

2. Taxonomy and Botanical Description

2.1 Systematic Classification

Hedychium spicatum belongs to the monocot order Zingiberales and exhibits the characteristic zygomorphic floral architecture of the Zingiberaceae. The systematic placement follows the APG IV (Angiosperm Phylogeny Group) classification system [5].

Table 1. Systematic classification of *Hedychium spicatum* (APG IV system)

Taxonomic Rank	Classification
Kingdom	Plantae
Clade	Tracheophytes – Angiosperms – Monocots – Commelinids
Order	Zingiberales
Family	Zingiberaceae Martinov
Subfamily	Zingiberoideae
Tribe	Zingibereae

Genus	Hedychium J. Koenig
Species	H. spicatum Buch. -Ham. ex D.Don (1825)
Synonyms	H. acuminatum Roscoe; H. roxburghii Blume
Common Names	Kapur Kachri (Hindi); Shati (Sanskrit); Spiked Ginger Lily (English); Sugandhabala (Bengali)

2.2 Morphological Features

Hedychium spicatum is a robust, aromatic, perennial rhizomatous herb that exhibits characteristic features of the Zingiberoideae subfamily. Detailed morphological description facilitates field identification and authentication of herbal materials [6].

- Rhizome: Horizontal, tuberous, irregularly branched; 2–4 cm in diameter; pale yellow to cream internally; strongly camphoraceous aroma attributable to 1,8-cineole and camphene content; covered in fibrous leaf base remnants externally.
- Stem: Pseudostem formed by tightly rolled leaf sheaths; erect, terete, 1.0–1.5 m in height; green, glabrous.
- Leaves: Alternate, distichous; blade lanceolate to oblong-lanceolate, 30–50 cm × 5–8 cm; apex acuminate; base attenuate; upper surface glabrous, lower surface sparsely pubescent; petiole absent or very short; ligule 1.5–2.5 cm, membranous.
- Inflorescence: Dense terminal spike, 15–30 cm long, 5–8 cm wide; bracts oblong-elliptic, 3.5–5 cm, green to reddish-brown, each subtending 2–3 flowers.
- Flowers: Zygomorphic, bisexual, fragrant; calyx tubular, 3–5 cm; corolla tube slender, 5–7 cm; lobes 3, linear; labellum broadly obovate, 3–4 cm wide, white to pale orange with yellow central stripe; staminode 2, petaloid; stamen 1, exserted.
- Fruit: Capsule, globose to oblong, 2–3 cm; orange-red when ripe; dehiscent to reveal 4–8 seeds covered in bright red arils.
- Seeds: Angled, 6–10 mm; aril fleshy, vermilion-red; facilitates bird-mediated seed dispersal.

2.3 Micromorphological and Anatomical Features

Pharmacognostic characterization of the rhizome includes diagnostic microscopic features essential for quality control and adulteration detection. The transverse section of the rhizome reveals a well-differentiated anatomy comprising a multi-layered cork, cortex of parenchymatous cells containing starch grains (2–18 μm diameter), vascular bundles scattered throughout the ground tissue (atactostele), oil cells distributed in the cortex and pith, and calcium oxalate crystals of rosette type [7]. These features distinguish *H. spicatum* from adulterants such as *H. coronarium* and *H. flavescens*.

3. Geographical Distribution, Ecology, and Conservation Status

3.1 Distribution Range

Hedychium spicatum exhibits a predominantly Himalayan and Indo-Malesian distribution pattern, reflecting its evolutionary origins in tropical montane Asia. The species demonstrates remarkable altitudinal adaptability, colonizing ecological niches across a wide elevational gradient [8].

Table 2. Geographical distribution and altitudinal range of *Hedychium spicatum*



Country / Region	Specific States / Provinces	Altitude Range (m)
India	Uttarakhand, Himachal Pradesh, Sikkim, Arunachal Pradesh, Assam, Meghalaya	1,000–3,000
Nepal	Mid-hills to sub-alpine belt; Annapurna, Langtang corridors	800–2,800
Bhutan	Eastern Himalayan foothills, Punakha Valley	600–2,500
China	Yunnan, Sichuan, Guizhou, Guangxi provinces	1,200–2,800
Myanmar	Northern Kachin and Shan highland zones	800–2,200
Sri Lanka	Central highlands; Knuckles Range	600–1,800

3.2 Ecological Preferences

The species thrives in moist, well-drained, humus-rich soils in semi-shaded forest understories, forest margins, stream banks, and rocky escarpments. It tolerates subtropical to temperate climatic regimes with mean annual rainfall of 1,500–3,500 mm and temperatures ranging from 5°C to 28°C. The plant is commonly associated with *Quercus-Rhododendron* and mixed broad-leaved forest communities in the Western Himalayas, and with *Shorea-Dipterocarpus* formations at lower elevations^[9].

3.3 Conservation Status and Threats

Hedychium spicatum is classified as a species of concern under Schedule VI of the Wildlife Protection Act (1972, India), which restricts commercial collection from wild populations. The International Union for Conservation of Nature (IUCN) has flagged the species as potentially vulnerable in several Indian states due to unsustainable rhizome harvesting for the essential oil trade, habitat fragmentation from agricultural encroachment, and climate-induced altitudinal

migration^[10]. Ex situ conservation through botanical gardens and in vitro propagation via tissue culture represent priority conservation strategies.

4. Ethnobotanical and Traditional Uses

4.1 Position in Classical Ayurvedic Literature

In Ayurvedic pharmacopoeia, *H. spicatum* is classified under the Sanskrit name Shati (also Gandhasuparnaka and Mishreya-shati). The classical texts characterize it as tikta (bitter), katu (pungent), and laghu (light) in guna, with ushna virya (heating potency). It pacifies vitiated kapha and vata doshas. The Ashtanga Hridayam (Sutrasthana 6) prescribes Shati as a component of Dashamoola formulations for respiratory disorders. The Charaka Samhita includes it in rasayana (rejuvenative) preparations and the Sharangadhara Samhita details its use in Lavangadi Vati for nausea^[3, 11].

4.2 Documented Ethnomedicinal Applications

Table 3. Documented ethnomedicinal uses of *Hedychium spicatum* across traditional systems

Therapeutic Category	Indication / Use	Plant Part / Preparation
Respiratory	Asthma, bronchitis, chronic cough, hiccups	Rhizome powder or decoction
Gastrointestinal	Diarrhea, dyspepsia, flatulence, vomiting, nausea	Rhizome powder with honey
Musculoskeletal	Rheumatism, arthritis, joint pain, myalgia	Poultice; oil massage
Neurological	Headache, epilepsy, neuralgia, anxiety	Essential oil inhalation
Dermatological	Wound healing, skin infections, dermatitis	Paste of fresh rhizome
Haematological	Blood disorders, anaemia	Decoction of rhizome
Oral / ENT	Bad breath, oral ulcers, sinusitis	Chewing dried rhizome
Reproductive	Aphrodisiac, dysmenorrhoea	Rhizome extract in ghee
Fever	Antipyretic, anti-malarial (folk)	Aqueous decoction
Psychoactive / Ritual	Incense for meditation; funerary rites	Dried rhizome; essential oil

4.3 Regional and Tribal Uses

Among the Bhotiya communities of Uttarakhand, the dried rhizome paste is applied to the forehead as a febrifuge and to swollen joints as an anti-arthritic poultice. The Adi and Nyishi tribes of Arunachal Pradesh use the aqueous rhizome extract as an antidiarrheal and to manage postpartum complications [12]. In Nepal, Tamang and Gurung healers incorporate the rhizome in polyherbal formulations for treating altitude sickness and as an adaptogen for high-altitude trekkers. Tibetan traditional medicine (Sowa-Rigpa) uses the plant (known as Ka-ko-la) in respiratory compound medicines analogous to its Ayurvedic application [13]. In Chinese traditional medicine, *H. spicatum* rhizome (known as Jiang Huang or Ba Jiao Jiang) is used in treating digestive complaints and as a warming stomachic. Sri Lankan Sinhala traditional medicine (Deshiya Chikitsa) employs the plant as a carminative and for treating rheumatic fever, reflecting convergent

ethnomedicinal applications across geographically distant cultures [14].

5. Phytochemical Constituents

5.1 Overview of Secondary Metabolite Profile

The phytochemical richness of *H. spicatum* is concentrated predominantly in the rhizome, though leaves, flowers, and seeds also contain pharmacologically relevant constituents. Systematic phytochemical investigations employing GC-MS, HPLC-DAD, LC-MS/MS, and NMR spectroscopy have characterized over 80 individual compounds across multiple chemical classes [15, 16].

5.2 Essential Oil Composition

The essential oil, obtained by hydrodistillation or steam distillation of the dried rhizome at a yield of 1.5–4.0% (w/w), represents the most extensively characterized phytochemical fraction. Its



composition demonstrates notable chemotypic altitude, harvest season, and drying conditions [17, variation correlated with geographical origin, 18].

Table 4. Major phytochemical constituents of *Hedychium spicatum* essential oil and their reported biological activities. NV = non-volatile fraction.

Compound	Chemical Class	Typical Content (%)	Key Reported Activity
1,8-Cineole (Eucalyptol)	Oxygenated monoterpene	15–35	Anti-inflammatory, antimicrobial, bronchodilatory
Camphene	Bicyclic monoterpene	8–20	Antioxidant, antimicrobial, flavour
Myrcene	Acyclic monoterpene	3–12	Analgesic, anti-inflammatory, sedative
α -Pinene	Bicyclic monoterpene	2–8	Antimicrobial, anti-inflammatory, bronchodilatory
β -Phellandrene	Cyclic monoterpene	2–7	Antifungal, cytotoxic
Linalool	Monoterpenoid alcohol	1–5	Anxiolytic, anti-nociceptive, anticonvulsant
Terpinen-4-ol	Monoterpenoid alcohol	1–4	Antimicrobial, anti-inflammatory
β -Caryophyllene	Sesquiterpene	2–6	Anti-inflammatory (CB2 agonist), antifungal
Hedychinone	Labdane diterpene	Major NV constituent	Anticancer, anti-inflammatory
Spicatanol	Labdane diterpene	Minor constituent	Cytotoxic, antimicrobial

5.3 Non-Volatile Phytoconstituents

5.3.1 Terpenoids

The non-volatile terpenoid fraction is dominated by labdane-type diterpenes, a structurally rare class that contributes substantially to the plant's pharmacological profile. Hedychinone (14-oxo-labda-8(17),12E-dien-16-oic acid), first isolated by Reddy et al. (1996), represents the most pharmacologically investigated labdane from this species [19]. Related diterpenes including 14-hydroxylabdatriene, spicatanene, and 8 β -hydroxylabda-12,14-diene have been subsequently characterized. Sesquiterpene lactones and monoterpene glycosides constitute additional minor terpenoid components.

5.3.2 Flavonoids and Polyphenols

HPLC-DAD analysis of rhizome methanol extracts has identified quercetin, kaempferol, rutin, and isorhamnetin as principal flavonoids. Flavonoid glycosides including quercetin-3-O-glucoside and kaempferol-3-O-rutinoside have been detected in the leaf fraction. Total phenolic content ranges from 28–68 mg gallic acid equivalents per gram dry weight depending on solvent polarity and extraction conditions [20, 21]. Phenolic acids including gallic acid, caffeic acid, ferulic acid, and p-coumaric acid contribute to the antioxidant capacity.

5.3.3 Alkaloids and Other Constituents

Minor alkaloid fractions have been detected in rhizome ethanolic extracts, though individual alkaloids remain to be fully characterized. Sterols including β -sitosterol, stigmasterol, and daucosterol have been isolated. The rhizome contains significant quantities of starch (25–35% dry weight), contributing to its nutritional and pharmaceutical excipient potential. Mucilaginous polysaccharides, tannins, saponins, and resins round out the phytochemical inventory. Seeds and arils contain fixed oils rich in linoleic and oleic acid [15].

5.3.4 Chemotypic Variation

Significant chemotypic variation in essential oil composition has been documented between populations from different geographical origins. High-altitude populations (>2,000 m) from Uttarakhand tend to be richer in 1,8-cineole and α -pinene, while lower-altitude Nepal populations show higher camphene content. Seasonal variation also significantly impacts oil yield and composition, with pre-flowering harvests typically yielding higher terpenoid content [18, 22].

6. Pharmacological Activities

6.1 Anti-inflammatory Activity

The anti-inflammatory potential of *H. spicatum* is the most extensively investigated pharmacological property, supported by multiple in vitro and in vivo study paradigms. Ethanolic and aqueous rhizome extracts significantly attenuate carrageenan-induced paw edema in Wistar rats at doses of 200–400 mg/kg, with efficacy comparable to diclofenac sodium at 10 mg/kg [23]. Mechanistic studies indicate inhibition of COX-2 enzymatic activity, suppression of NF- κ B nuclear translocation, and reduction of pro-inflammatory cytokine (IL-1 β , IL-6, TNF- α) biosynthesis. Hedychinone specifically inhibits the COX-

2/PGE₂ pathway with an IC₅₀ of approximately 18 μ M in lipopolysaccharide-stimulated RAW264.7 macrophages [24]. The sesquiterpene β -caryophyllene contributes an additional anti-inflammatory dimension via selective cannabinoid receptor 2 (CB2) agonism, modulating macrophage activation without CNS side effects.

6.2 Antioxidant Activity

Free radical scavenging capacity has been demonstrated across multiple assay platforms. The methanolic rhizome extract yields DPPH IC₅₀ values of 48–78 μ g/mL, ABTS \bullet + inhibition IC₅₀ of 38–65 μ g/mL, and ferric reducing antioxidant power (FRAP) values of 120–185 mg ascorbic acid equivalent/g dry weight [20, 21]. Flavonoid and polyphenolic fractions account for the majority of radical-scavenging activity. The essential oil demonstrates moderate antioxidant activity (DPPH IC₅₀: 180–320 μ g/mL), with camphene and terpinen-4-ol as principal contributors. Cellular antioxidant assays confirm cytoprotective effects in H₂O₂-challenged HepG2 cells, with significant upregulation of Nrf2-dependent antioxidant enzymes (SOD, CAT, GPx) [25].

6.3 Antimicrobial Activity

The essential oil demonstrates broad-spectrum bacteriostatic and bactericidal activity. The minimum inhibitory concentration (MIC) ranges documented against common pathogens are: *Staphylococcus aureus* (MIC 0.3–1.2 mg/mL), *Bacillus subtilis* (0.2–0.9 mg/mL), *Escherichia coli* (0.6–2.4 mg/mL), *Pseudomonas aeruginosa* (1.2–3.0 mg/mL), and *Salmonella typhi* (0.4–1.5 mg/mL) [26]. Antifungal activity against *Candida albicans* (MIC 0.4–0.8 mg/mL), *Aspergillus flavus*, and *Aspergillus niger* has been reported. The mechanism of antibacterial action involves disruption of bacterial membrane integrity, inhibition of cell wall biosynthesis, and



impairment of membrane-bound ATPase activity. 1,8-Cineole and α -pinene synergistically contribute to this membrane-disrupting mechanism. Rhizome extracts also show activity against drug-resistant MRSA strains, suggesting potential adjunct utility in infectious disease management [27].

6.4 Analgesic Activity

Significant analgesic effects have been documented in both acute and chronic pain models. The acetic acid-induced writhing test (visceral pain) and hot plate test (supraspinal analgesia) demonstrate dose-dependent pain inhibition at 100–400 mg/kg in Swiss albino mice. Naloxone pre-treatment partially reverses the analgesic effect, implicating opioid receptor involvement. Peripheral mechanisms include inhibition of prostaglandin synthesis and bradykinin-mediated pain sensitization. The monoterpene myrcene, known to act synergistically with opioid receptors, likely contributes a central analgesic component [23].

6.5 Anticancer Activity

In vitro cytotoxicity of *H. spicatum* extracts and isolated constituents has been evaluated against a panel of human cancer cell lines. IC₅₀ values reported include: HeLa (cervical carcinoma) 28–55 μ g/mL, MCF-7 (breast adenocarcinoma) 32–62 μ g/mL, A549 (non-small cell lung carcinoma) 41–75 μ g/mL, and HCT116 (colorectal carcinoma) 38–70 μ g/mL [28]. Hedychinone induces intrinsic apoptosis in MCF-7 cells via upregulation of Bax, downregulation of Bcl-2, cytochrome-c release, and caspase-3/7 activation. Flow cytometric analysis confirms G2/M phase cell cycle arrest at IC₅₀ concentrations. Molecular docking studies indicate hedychinone binding affinity for both EGFR kinase (binding energy –9.2 kcal/mol) and tubulin polymerization sites. Anti-angiogenic

effects evidenced by VEGF inhibition in EAhy926 endothelial cells further support anticancer potential [29].

6.6 Hepatoprotective Activity

Administration of *H. spicatum* rhizome extract at 200–400 mg/kg significantly attenuates paracetamol (3 g/kg)- and CCl₄ (1.5 mL/kg)-induced hepatotoxicity in Wistar rats. Biochemical parameters normalized include serum ALT, AST, ALP, total bilirubin, and albumin. Total protein restoration is also observed. Histopathological evaluation confirms a marked reduction in hepatocellular necrosis, inflammatory cell infiltration, and hepatic ballooning, with efficacy approaching that of the reference hepatoprotectant silymarin (100 mg/kg) [30]. The mechanism involves antioxidant-mediated hepatocyte protection, suppression of hepatic NF- κ B activation, and enhancement of glutathione biosynthesis.

6.7 Anti-asthmatic and Bronchodilatory Activity

In isolated guinea pig tracheal chain preparations, *H. spicatum* essential oil and its constituents (1,8-cineole, α -pinene) produce significant relaxation of carbachol- and histamine-induced bronchospasm. The bronchodilatory mechanism involves both β_2 -adrenoceptor agonism and phosphodiesterase inhibition leading to intracellular cAMP accumulation. 1,8-Cineole additionally inhibits the release of histamine from mast cells, reducing the allergen-stimulated acute phase of asthmatic response. These findings provide robust pharmacological validation for the traditional use of *H. spicatum* in Ayurvedic respiratory formulations [3, 17].

6.8 Immunomodulatory Activity

Immunomodulatory properties of rhizome polysaccharide fractions and flavonoid-enriched extracts have been assessed in cyclophosphamide-induced immunosuppressed mice. Parameters evaluated include hemagglutination antibody titre, delayed-type hypersensitivity (DTH) response, and neutrophil adhesion. Significant stimulation of both humoral and cell-mediated immunity is observed, suggesting potential as an adjuvant in immunocompromised states. The polysaccharide fraction activates macrophage phagocytic activity via TLR4 signalling [31].

6.9 Neuroprotective and CNS Activity

Emerging evidence from rodent models suggests neuroprotective activity of *H. spicatum* extracts against scopolamine-induced cognitive deficits in the Morris Water Maze paradigm. Inhibition of acetylcholinesterase (AChE) activity in cortical homogenates provides a mechanistic basis for the observed cognitive enhancement, relevant to Alzheimer's disease drug discovery. The essential oil component linalool demonstrates GABA-A receptor modulation, contributing sedative and anxiolytic effects validated in elevated plus maze models [32].

6.10 Additional Pharmacological Activities

- Diuretic: Aqueous extracts increase urine output in Wistar rats, validating folk use in renal calculi.
- Anti-ulcerogenic: Cytoprotective effects on gastric mucosa demonstrated in pylorus-ligated rat model.
- Antidiabetic: Moderate α -glucosidase inhibitory activity (IC₅₀: 180–320 μ g/mL) in vitro.
- Insecticidal: Essential oil vapour repels *Aedes aegypti* mosquitoes with >85% repellency at 5% concentration.
- Anti-inflammatory (topical): Gel formulations show anti-inflammatory efficacy in croton oil ear-oedema model in mice [33].

7. Industrial and Commercial Applications

The multifaceted utility of *H. spicatum* underpins its commercial importance across several industry sectors. The high essential oil content and chemically diverse non-volatile fraction offer exploitable resources for product development.

Table 5. Industrial applications of *Hedychium spicatum* and associated key phytochemicals

Industry Sector	Application	Key Compound(s)
Pharmaceuticals	Herbal tablets, capsules, syrups, standardized extracts	Hedychinone, 1,8-cineole
Cosmetics & Perfumery	High-end perfumes, aromatic oils, hair care, skin lighteners	Essential oil blend, linalool
Aromatherapy	Diffuser oils, massage blends, steam inhalation	1,8-Cineole, camphene
Food & Flavouring	Spice blends, confectionery, beverages (GRAS status)	Camphene, β -phellandrene
Ayurvedic Formulations	Churna, kwatha, ghrita, tailam, vati	Multi-component synergy
Agrochemicals	Biopesticide, mosquito repellent formulations	1,8-Cineole, α -pinene

Veterinary Medicine	Anti-parasitic preparations in livestock	Essential oil
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The global market for natural fragrances and aromatherapy products, valued at USD 7.8 billion in 2023 and projected to reach USD 12.4 billion by 2030, presents a significant commercial opportunity for *H. spicatum* essential oil. The rhizome's GRAS (Generally Recognized as Safe) status in the United States and its historical use in food systems position it favourably for nutraceutical development^[34].

8. Toxicology and Safety Profile

8.1 Acute Toxicity

Acute oral toxicity studies conducted in Swiss albino mice and Wistar rats following OECD Test Guideline 423 indicate that the LD₅₀ of aqueous and ethanolic rhizome extracts exceeds 2,000 mg/kg body weight, classifying them in Category 5 (practically non-toxic) according to the Globally Harmonized System (GHS) of Classification^[35]. No mortalities or signs of acute toxicity were observed at doses up to 3,000 mg/kg in rodent studies.

8.2 Sub-acute and Sub-chronic Toxicity

Twenty-eight-day sub-acute toxicity studies at doses of 100, 300, and 500 mg/kg in Wistar rats reveal no clinically significant alterations in body weight gain, feed consumption, organ-to-body weight ratios, haematological parameters (CBC, differential leucocyte count), serum biochemistry (ALT, AST, ALP, creatinine, blood urea nitrogen, total protein, albumin, glucose), or gross and histopathological organ morphology. The NOAEL (No Observed Adverse Effect Level) is established at 500 mg/kg/day for sub-acute administration^[35].

8.3 Essential Oil Safety

The neat essential oil demonstrates mild mucous membrane irritancy and should not be applied undiluted to skin. Dermal sensitization risk is low at concentrations below 2% in carrier oils. 1,8-Cineole at high doses (>2 g/kg in rodents) induces CNS stimulation and convulsions; however, therapeutic doses used in aromatherapy pose negligible risk. Contraindicated in infants under 2 years and pregnant women in the first trimester based on precautionary principles^[36].

8.4 Genotoxicity and Mutagenicity

The Ames test (*Salmonella typhimurium* strains TA98, TA100) shows no mutagenic activity up to 5,000 µg/plate for standard rhizome extracts. In vitro chromosomal aberration and micronucleus assays in Chinese hamster ovary cells are negative at pharmacologically relevant concentrations. Formal in vivo genotoxicity data remain limited and constitute a significant gap in the safety documentation^[37].

9. Research Gaps and Challenges

Despite the breadth of pre-clinical evidence, significant scientific lacunae impede the translational development of *H. spicatum* as a mainstream therapeutic agent:

- Absence of Phase I/II randomized controlled clinical trials for any of the documented pharmacological activities, rendering all clinical claims extrapolatory from animal models.
- No validated pharmacokinetic data (absorption, distribution, metabolism, excretion — ADME) for hedychinone or major essential oil constituents in human subjects.



- Lack of globally standardized quality specifications, pharmacopoeial monographs, or WHO herbal monograph for *H. spicatum* raw material and herbal preparations.
 - Insufficient characterization of drug–herb interaction potential, particularly with CYP450 enzyme substrates and P-glycoprotein-mediated transporters.
 - Limited mechanistic data at the molecular level for neuroprotective, immunomodulatory, and antidiabetic activities; most studies rely on crude extract fractions.
 - Inadequate investigation of epigenetic mechanisms and gut microbiome-mediated biotransformation of *H. spicatum* phytoconstituents.
 - Absence of controlled studies on geographical, seasonal, and post-harvest processing-related chemotypic variation and its impact on biological activity.
 - Scarcity of population-level ethnobotanical surveys to document the full scope of traditional knowledge before irreversible cultural erosion.
 - No validated in vitro dissolution and bioavailability models for standardized oral formulations.
 - Insufficient data on reproductive and developmental toxicity, carcinogenicity, and long-term chronic toxicity.
- provides unprecedented tools to accelerate the development of *H. spicatum*-derived therapeutics:
- Targeted isolation and synthetic optimization of hedychinone analogues as novel anti-inflammatory or anticancer lead compounds, leveraging structure-activity relationship (SAR) studies.
 - Network pharmacology and systems biology approaches to map the polypharmacological target landscape of *H. spicatum* constituents against disease-relevant protein interaction networks.
 - Nanotechnology-enabled delivery systems (polymeric nanoparticles, solid lipid nanoparticles, nanoemulsions, liposomal encapsulation) to improve bioavailability and targeted delivery of hydrophobic labdane diterpenes.
 - Development of standardized fixed-dose combination herbal formulations with defined biomarker content (e.g., minimum 0.3% hedychinone, 1.5% 1,8-cineole) and validated analytical methods (HPLC, TLC densitometry).
 - Clinical validation through multi-centre randomized controlled trials prioritizing the anti-inflammatory and respiratory indications with highest pre-clinical evidence base.
 - Micropropagation via tissue culture (organogenesis, somatic embryogenesis) and callus cultures for sustainable and genetically uniform biomass production independent of wild harvesting.
 - Hairy root culture technology using *Agrobacterium rhizogenes* transformation for scalable production of labdane diterpenes under controlled conditions.

10. Future Prospects

The convergence of modern analytical chemistry, network pharmacology, and translational medicine



- Metabolomics-guided quality control employing ¹H-NMR fingerprinting and HPLC-QTOF-MS profiling to establish comprehensive chemotypic databases linking origin to bioactivity.
- Integration of *H. spicatum* cultivation into Himalayan agroforestry systems as a high-value intercrop to incentivize conservation by providing sustainable rural livelihoods.

CONCLUSION

Hedychium spicatum stands as a pharmacognostically significant medicinal plant whose therapeutic legacy, rooted in millennia of traditional practice, is increasingly validated by rigorous scientific inquiry. The integration of ethnobotanical knowledge with modern phytochemical and pharmacological methodologies has established a compelling evidence base supporting its utility in inflammatory, respiratory, hepatic, antimicrobial, and oncological disease contexts. The structurally distinctive labdane diterpene hedychinone, the dominant oxygenated monoterpene 1,8-cineole, and the diverse flavonoid-polyphenol matrix collectively underpin a multi-target pharmacological profile uniquely suited to the poly-pathological nature of chronic diseases. The anti-inflammatory and antioxidant activities, in particular, are supported by mechanistic evidence of sufficient depth to justify clinical investigation. However, the translational gap between pre-clinical promise and clinical application remains substantial. Bridging this gap necessitates concerted investment in ADME characterization, formal clinical trials, pharmacopoeial standardization, and sustainable cultivation infrastructure. Conservation of wild populations, imperilled by overexploitation and habitat loss, must proceed in parallel with research intensification. With appropriately directed

scientific and regulatory investment, *H. spicatum* has the potential to yield a new generation of evidence-based botanical medicines addressing unmet clinical needs, particularly in respiratory, anti-inflammatory, and oncological therapeutics, while simultaneously safeguarding the Himalayan biodiversity that sustains its existence.

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