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Review Paper

Formulation and Evaluation of Ethosomal Hydrogel Containing Vitamin A, Vitamin E and Ginger Extract for Enhanced Dermal Delivery: A Comprehensive Review

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ABSTRACT

Topical drug delivery has emerged as an attractive alternative to oral and parenteral administration owing to its ability to provide localized therapeutic action while minimizing systemic adverse effects. However, the highly organized structure of the stratum corneum significantly restricts the penetration of many therapeutic agents, particularly hydrophilic and high molecular weight compounds. Conventional topical formulations such as creams, ointments, and gels often exhibit limited skin permeation and poor bioavailability, resulting in suboptimal therapeutic outcomes. Nanovesicular carrier systems, especially ethosomes, have gained considerable attention because of their superior ability to enhance transdermal and dermal drug delivery through improved skin penetration. Ethosomes are soft, malleable phospholipid vesicles containing high concentrations of ethanol, which fluidizes both vesicular lipids and the stratum corneum lipid bilayer, thereby facilitating deeper penetration into the skin. Incorporation of ethosomes into hydrogel matrices further improves formulation stability, spreadability, residence time, patient compliance, and controlled drug release. The combination of bioactive compounds such as Vitamin A, Vitamin E, and ginger extract within an ethosomal hydrogel offers a multifunctional therapeutic strategy owing to their antioxidant, anti-inflammatory, antimicrobial, collagen-stimulating, and wound-healing properties. Vitamin A regulates epidermal differentiation and collagen synthesis, making it highly effective in skin rejuvenation and treatment of photoaging. Vitamin E protects cellular membranes against oxidative damage through its potent free radical scavenging activity while improving skin hydration and barrier function. Ginger extract contains several bioactive phytoconstituents including 6-gingerol, 6-shogaol, and zingerone that exhibit remarkable anti-inflammatory, antioxidant, antimicrobial, and wound-healing activities. The synergistic combination of these bioactives within an ethosomal hydrogel has the potential to enhance dermal penetration, improve therapeutic efficacy, and

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provide sustained drug release. This review comprehensively discusses the anatomical structure of the skin, challenges associated with dermal drug delivery, principles of ethosomal drug delivery systems, hydrogel-based formulations, pharmacological importance of Vitamin A, Vitamin E, and ginger extract, formulation strategies, evaluation parameters, recent research advances, and future prospects. The review also identifies current research gaps and highlights the potential of multifunctional ethosomal hydrogels as promising nanocarrier systems for advanced topical drug delivery.

INTRODUCTION

The skin is the largest organ of the human body, accounting for approximately 15–16% of total body weight and covering an average surface area of nearly 2 m² in adults. Besides serving as a physical barrier against environmental insults, pathogens, ultraviolet radiation, and chemical agents, the skin also plays a critical role in thermoregulation, immune surveillance, sensory perception, and maintenance of fluid homeostasis [1,2]. Owing to its accessibility and extensive vascular network, the skin has become an attractive route for localized as well as systemic drug delivery.

Topical drug delivery offers several advantages over conventional oral and injectable routes, including avoidance of hepatic first-pass metabolism, improved patient compliance, reduced gastrointestinal irritation, prolonged drug residence at the target site, and lower systemic toxicity [3]. Despite these advantages, effective dermal drug delivery remains challenging due to the barrier properties of the stratum corneum, which restricts the penetration of most therapeutic molecules. The "brick-and-mortar" organization of corneocytes embedded within a lipid matrix acts as the primary obstacle to drug permeation [4].

Several strategies have been investigated to overcome the skin barrier, including chemical penetration enhancers, microneedles, iontophoresis, sonophoresis, lipid nanoparticles, liposomes, transfersomes, niosomes, and

ethosomes [5]. Among these advanced nanocarrier systems, ethosomes have demonstrated remarkable potential because of their unique composition consisting of phospholipids, ethanol, and water. High ethanol concentration increases membrane fluidity and disrupts the highly ordered lipid arrangement of the stratum corneum, thereby facilitating enhanced penetration of therapeutic molecules into deeper skin layers [6].

Hydrogels are three-dimensional hydrophilic polymeric networks capable of absorbing large amounts of water while maintaining structural integrity. Owing to their excellent biocompatibility, non-greasy nature, ease of application, and controlled drug release properties, hydrogels have become one of the most widely used vehicles for topical drug delivery [7]. The incorporation of ethosomal vesicles into hydrogel systems combines the penetration-enhancing capability of ethosomes with the sustained-release characteristics and patient-friendly properties of hydrogels, resulting in improved therapeutic performance.

Among various dermatological bioactive compounds, Vitamins A and E have received considerable attention because of their essential roles in maintaining skin integrity and preventing oxidative damage. Vitamin A stimulates epidermal renewal, regulates keratinocyte differentiation, enhances collagen production, and reduces photoaging [8]. Vitamin E (α -tocopherol) is one of the most effective lipid-soluble antioxidants, protecting skin cells against oxidative stress induced by ultraviolet radiation and environmental pollutants while improving skin hydration and elasticity [9].

Natural herbal extracts have also gained increasing interest owing to their excellent safety profile and multiple pharmacological activities. Ginger (*Zingiber officinale* Roscoe), an important medicinal plant belonging to the family *Zingiberaceae*, contains numerous bioactive



constituents such as 6-gingerol, 6-shogaol, paradols, and zingerone. These phytochemicals exhibit potent antioxidant, anti-inflammatory, antimicrobial, analgesic, and wound-healing activities, making ginger extract a promising candidate for topical therapeutic applications [10]. The combination of Vitamin A, Vitamin E, and ginger extract within an ethosomal hydrogel provides a multifunctional nanocarrier capable of addressing multiple pathological pathways simultaneously. Vitamin A promotes skin regeneration, Vitamin E protects against oxidative stress, whereas ginger extract suppresses inflammatory mediators and microbial growth. Encapsulation of these bioactive compounds into ethosomal vesicles further enhances their stability, skin penetration, and therapeutic efficacy while minimizing degradation and irritation.

Recent advances in nanotechnology have further strengthened the application of ethosomal hydrogels for treatment of inflammatory skin diseases, wound healing, anti-aging therapy, psoriasis, acne vulgaris, fungal infections, and cosmetic dermatology. Nevertheless, challenges related to formulation stability, large-scale manufacturing, regulatory approval, and long-term safety still require extensive investigation before successful clinical translation.

Therefore, this review summarizes the current understanding of ethosomal hydrogel systems containing Vitamin A, Vitamin E, and ginger extract, with particular emphasis on formulation strategies, characterization techniques, dermal delivery mechanisms, recent research progress, and future perspectives for advanced topical drug delivery.

2. Skin Anatomy and Barrier Function

The skin is a complex multilayered organ composed of three principal layers: **epidermis**, **dermis**, and **hypodermis (subcutaneous tissue)**. Each layer possesses unique structural and

physiological characteristics that collectively contribute to protection, thermoregulation, immune defense, and maintenance of internal homeostasis [11].

The **epidermis** is the outermost layer and consists primarily of keratinocytes arranged into five distinct strata: stratum basale, stratum spinosum, stratum granulosum, stratum lucidum (present only in thick skin), and stratum corneum. Among these, the **stratum corneum** is the principal barrier to percutaneous drug absorption. It is composed of dead, flattened corneocytes embedded within an extracellular lipid matrix containing ceramides, cholesterol, and free fatty acids, often described as the "brick-and-mortar" model [12].

The **dermis**, located beneath the epidermis, is composed of collagen fibers, elastin, fibroblasts, blood vessels, lymphatics, nerves, and skin appendages such as hair follicles and sweat glands. This layer provides mechanical strength, elasticity, nutrient supply, and immune support. Drug molecules reaching the dermis may enter systemic circulation through the extensive capillary network [13].

The **hypodermis** consists mainly of adipose tissue and connective tissue that serve as thermal insulation, mechanical cushioning, and energy storage. It also acts as a depot for lipophilic drugs and contributes to prolonged drug residence in certain formulations [14].

Drug permeation across the skin occurs primarily through three pathways:

1. **Intercellular pathway**
2. **Transcellular pathway**
3. **Transappendageal pathway (hair follicles and sweat glands)**

Among these, the intercellular pathway is the predominant route for passive diffusion of lipophilic molecules. Ethosomal vesicles enhance drug penetration by increasing lipid fluidity within



the stratum corneum and facilitating deeper transport through these pathways [15]

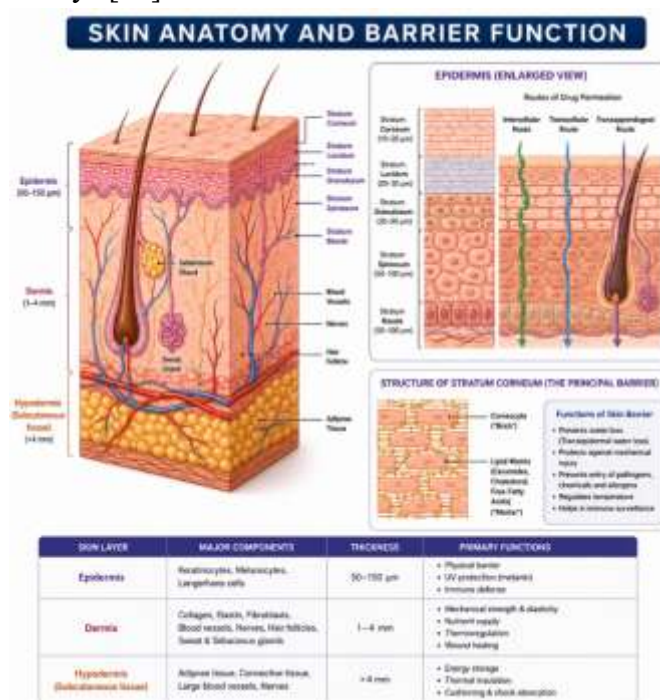


Figure 1 Skin Anatomy and Barrier Function

Table 1. Major Layers of Human Skin and Their Functions

Skin Layer	Major Components	Primary Functions
Epidermis	Keratinocytes, melanocytes, Langerhans cells	Physical barrier, UV protection, immune defense
Stratum Corneum	Corneocytes, ceramides, cholesterol	Principal barrier to drug permeation
Dermis	Collagen, elastin, fibroblasts, blood vessels	Mechanical support, nutrient supply, wound healing
Hypodermis (Subcutaneous tissue)	Adipose tissue, connective tissue, large blood vessels, nerves	Thermal insulation, energy storage, cushioning

Table 2. Advantages and Limitations of Topical Drug Delivery

S.No.	Advantages	Limitations
1.	Avoids first-pass metabolism	Limited penetration through stratum corneum
2.	Localized drug delivery	Low bioavailability of hydrophilic drugs
3.	Reduced systemic toxicity	Skin irritation from some excipients
4.	Improved patient compliance	Variable permeability among individuals
5.	Sustained therapeutic effect	Enzymatic degradation of certain drugs

3. Ethosomes: A Novel Vesicular Drug Delivery System

Ethosomes are advanced phospholipid-based nanovesicular carriers specifically designed to

enhance the dermal and transdermal delivery of therapeutic agents. Introduced by Touitou and co-workers in 2000, ethosomes consist primarily of phospholipids, a relatively high concentration of

ethanol (20–45%), water, and optional additives such as glycols or cholesterol [16]. Unlike conventional liposomes, the high ethanol content imparts remarkable flexibility and deformability to the vesicular membrane, allowing ethosomes to penetrate the compact lipid architecture of the stratum corneum more effectively.

The unique combination of ethanol and phospholipids enables ethosomes to transport both hydrophilic and lipophilic drugs into deeper skin layers without causing irreversible damage to the skin barrier. Ethosomes have demonstrated significant success in delivering corticosteroids, antifungal agents, antibiotics, antioxidants, anti-inflammatory drugs, herbal extracts, peptides, and cosmetic ingredients [17].

The increasing interest in ethosomal drug delivery is attributed to their high encapsulation efficiency, improved drug stability, controlled release behavior, and enhanced patient compliance. Recent developments have focused on integrating ethosomes into semisolid dosage forms such as hydrogels, creams, and emulgels to further improve topical retention and therapeutic efficacy [18].

3.1 Composition of Ethosomes

Ethosomes are composed of four major components, each contributing a distinct function to the vesicular system.

Phospholipids

Phospholipids constitute the structural framework of ethosomal vesicles. Commonly used phospholipids include: Soy phosphatidylcholine, Egg phosphatidylcholine, Hydrogenated phosphatidylcholine & Phosphatidylserine.

These amphiphilic molecules self-assemble into bilayer vesicles capable of encapsulating both hydrophilic and lipophilic therapeutic agents.

Ethanol

Ethanol is the most important component of ethosomes and is generally present at concentrations ranging from 20–45%. Ethanol performs multiple functions:

- Increases membrane flexibility
- Fluidizes phospholipid bilayers
- Disrupts intercellular lipid organization within the stratum corneum
- Enhances drug solubility
- Improves drug permeation

Water

Purified water forms the continuous phase of the vesicular dispersion and hydrates phospholipid molecules.

Polyols

Propylene glycol or Transcutol® may be incorporated to improve vesicle stability, enhance skin hydration, and further increase drug permeation

Table 3. Major Components of Ethosomes and Their Functions

Component	Typical Concentration	Function
Phospholipid	1–5%	Vesicle formation
Ethanol	20–45%	Penetration enhancement
Water	q.s.	Hydration medium
Propylene glycol	5–20%	Co-solvent and humectant
Cholesterol (optional)	0.5–2%	Membrane stabilization

3.2 Preparation Methods of Ethosomes

Several preparation methods have been reported depending upon drug characteristics and formulation requirements.



3.2.1 Cold Method

The cold method is the most widely employed technique for laboratory-scale preparation.

Procedure

1. Phospholipids are dissolved in ethanol under continuous stirring.
2. Propylene glycol is added to the ethanolic phase.
3. The aqueous phase is heated separately to approximately 30°C.
4. Water is slowly added to the ethanolic mixture under vigorous stirring.
5. The resulting dispersion is sonicated or extruded to reduce vesicle size.

3.2.2 Hot Method

- In this method, phospholipids are dispersed in warm water while ethanol is heated separately. Both phases are mixed at controlled temperature to obtain ethosomal vesicles.
- Although relatively less common, the hot method may improve the solubilization of certain lipid components

3.2.3 Thin Film Hydration Method

- A thin phospholipid film is first prepared by solvent evaporation under reduced pressure. Hydration of the lipid film with hydroethanolic solution results in ethosomal vesicles.
- This method generally provides uniform vesicle formation but requires additional processing time

Table 4. Comparison of Ethosome Preparation Methods

Method	Advantages	Limitations
Cold Method	Simple, reproducible	Requires sonication
Hot Method	Suitable for some lipids	Not ideal for thermolabile drugs
Thin Film Hydration	Uniform vesicles	Time-consuming
Reverse Phase Evaporation	High encapsulation	Organic solvent exposure

3.3 Mechanism of Skin Penetration

The superior skin penetration ability of ethosomes is primarily attributed to the synergistic action of ethanol and phospholipid vesicles.

Step 1: Ethanol Interaction

Ethanol penetrates the stratum corneum and disrupts the highly ordered lipid arrangement by increasing lipid fluidity.

Step 2: Vesicle Softening

The presence of ethanol makes phospholipid bilayers highly flexible and deformable.

Step 3: Enhanced Vesicle Penetration

Soft vesicles squeeze through the disrupted lipid domains and migrate into deeper epidermal layers.

Step 4: Drug Release

Once inside viable skin layers, the vesicles gradually release encapsulated drug molecules, producing prolonged therapeutic action.

This mechanism results in significantly greater drug deposition compared with conventional liposomes.

3.4 Factors Affecting Ethosomal Characteristics

Several formulation variables influence the physicochemical properties and performance of ethosomes.

Ethanol Concentration

Increasing ethanol concentration decreases vesicle size and increases flexibility. However,



concentrations above approximately 45% may destabilize vesicles.

Phospholipid Concentration

Higher phospholipid concentration generally increases vesicle size and entrapment efficiency.

Cholesterol Content

Cholesterol enhances membrane rigidity but excessive amounts may reduce drug release

Sonication Time

Longer sonication reduces particle size but excessive sonication may result in drug leakage.

Storage Temperature

Refrigerated storage (4–8°C) generally improves vesicle stability

3.5 Advantages of Ethosomes

Ethosomes possess several advantages over conventional topical formulations:

- Excellent skin penetration
- High drug entrapment efficiency
- Controlled drug release
- Enhanced bioavailability
- Improved stability
- Non-invasive administration
- Reduced dosing frequency
- Better patient compliance
- Suitable for hydrophilic and lipophilic drugs
- Lower systemic toxicity

3.6 Limitations of Ethosomes

Despite numerous advantages, ethosomes also exhibit certain limitations:

- Phospholipid oxidation during storage
- Possible ethanol evaporation
- Relatively high production cost
- Limited long-term stability
- Scale-up challenges
- Requirement for specialized characterization techniques

Table 5. Comparison of Liposomal Nanocarriers

Property	Liposomes	Transfersomes	Ethosomes	Transethosomes
Ethanol	No	No	High	High
Edge Activator	No	Yes	No	Yes
Vesicle Flexibility	Moderate	High	Very High	Extremely High
Skin Penetration	Moderate	High	Excellent	Excellent
Drug Entrapment	Moderate	High	High	High
Dermal Delivery	Good	Better	Excellent	Excellent

4. Hydrogel-Based Drug Delivery Systems

Hydrogels are three-dimensional crosslinked polymeric networks capable of absorbing large quantities of water while maintaining their structural integrity. Owing to their hydrophilic nature, excellent biocompatibility, softness, and controlled drug release behavior, hydrogels have become one of the most widely investigated semisolid dosage forms for topical drug delivery [19].

Hydrogels provide a moist environment at the site of application, improving skin hydration and

facilitating enhanced drug permeation. Their non-greasy texture, cooling sensation, ease of washing, and patient acceptability make them superior to conventional ointments and creams for many dermatological applications.

When ethosomal dispersions are incorporated into hydrogel matrices, the resulting **ethosomal hydrogel** combines the penetration-enhancing capability of ethosomes with the prolonged residence time and controlled release properties of hydrogels. Such hybrid systems improve formulation stability, reduce vesicle aggregation,



enhance skin retention, and increase therapeutic efficacy [20].

4.1 Ideal Characteristics of Hydrogels

An ideal hydrogel intended for topical application should possess the following characteristics:

- Biocompatible and non-toxic
- Non-irritant and non-sensitizing
- Appropriate viscosity
- Good spreadability
- Easy extrudability
- High drug-loading capacity
- Controlled drug release
- Good mechanical strength
- Stable during storage
- Easily washable with water

4.2 Polymers Used in Ethosomal Hydrogels

Common polymers include: Carbopol 934, Carbopol 940, Carbopol 971, Hydroxypropyl methylcellulose (HPMC), Sodium carboxymethyl cellulose (NaCMC), Poloxamer 407, Chitosan, Xanthan gum, Sodium alginate and Polyvinyl alcohol (PVA).

Among these, **Carbopol 934 and Carbopol 940** are the most extensively used owing to their excellent gelling ability, transparency, bioadhesion, and compatibility with ethosomal dispersions.

5. Vitamin A in Dermal Drug Delivery

Vitamin A is a fat-soluble vitamin that plays an indispensable role in maintaining skin integrity, cellular differentiation, epithelial growth, immune regulation, and collagen synthesis. It belongs to the retinoid family, which includes retinol, retinaldehyde, retinoic acid, and retinyl esters. Among these, retinol and retinoic acid are the most extensively utilized in dermatological and cosmetic formulations due to their proven efficacy in improving skin texture, reducing wrinkles, stimulating collagen production, and treating various hyperproliferative skin disorders [21].

Topically applied Vitamin A promotes epidermal renewal by regulating keratinocyte proliferation and differentiation through interaction with nuclear retinoic acid receptors (RARs) and retinoid X receptors (RXRs). Activation of these receptors modulates the expression of genes responsible for collagen synthesis, epidermal turnover, and extracellular matrix remodeling [22].

Vitamin A also inhibits matrix metalloproteinases (MMPs), enzymes responsible for collagen degradation during aging and photoaging. Consequently, topical Vitamin A formulations improve skin elasticity, reduce fine wrinkles, accelerate wound healing, and restore epidermal barrier function [23].

Despite its remarkable therapeutic potential, Vitamin A exhibits several pharmaceutical limitations including poor aqueous solubility, sensitivity to light and oxygen, rapid degradation, and skin irritation at higher concentrations. Nano-vesicular carriers such as ethosomes provide protection against oxidative degradation while significantly improving dermal penetration and sustained release [24].

6. Vitamin E in Dermal Drug Delivery

Vitamin E represents a family of lipid-soluble antioxidants consisting of tocopherols and tocotrienols, among which α -tocopherol exhibits the highest biological activity. Vitamin E is naturally present within the epidermis and sebaceous glands where it protects cellular membranes against oxidative damage induced by ultraviolet radiation and environmental pollutants [25].

Oxidative stress generated by reactive oxygen species (ROS) accelerates lipid peroxidation, collagen degradation, inflammation, and premature skin aging. Vitamin E interrupts these oxidative chain reactions by donating hydrogen atoms to lipid radicals, thereby preventing



membrane damage and preserving cellular integrity [26].

Topical Vitamin E also enhances epidermal hydration by stabilizing intercellular lipids within the stratum corneum. Moreover, it exhibits anti-inflammatory activity through inhibition of prostaglandin synthesis and suppression of inflammatory cytokines

Nanoencapsulation of Vitamin E within ethosomal vesicles protects it against oxidation and enhances penetration into deeper skin layers, providing prolonged antioxidant protection.

7. Ginger Extract in Dermal Drug Delivery

Ginger (*Zingiber officinale* Roscoe) is one of the oldest medicinal plants belonging to the family *Zingiberaceae*. Its rhizome contains numerous pharmacologically active phytoconstituents including: 6-Gingerol, 8-Gingerol, 10-Gingerol, 6-Shogaol, Zingerone, Paradols, Essential oils, Flavonoids and Terpenoids

These compounds exhibit broad-spectrum biological activities including antioxidant, anti-inflammatory, antimicrobial, analgesic, anticancer, and wound-healing effects [28].

Among these constituents, **6-gingerol** is considered the principal bioactive compound responsible for most pharmacological actions

8. Synergistic Therapeutic Effects of Vitamin A, Vitamin E and Ginger Extract

- The simultaneous incorporation of Vitamin A, Vitamin E, and ginger extract into an ethosomal hydrogel provides a multifunctional therapeutic platform with complementary mechanisms of action. Each component contributes distinct pharmacological benefits while enhancing the efficacy of the others.
- Vitamin A promotes epidermal regeneration and collagen synthesis, Vitamin E protects against oxidative stress and stabilizes cellular membranes, whereas ginger extract suppresses inflammation and microbial growth. Their combined use may therefore improve wound healing, reduce oxidative damage, enhance skin hydration, and restore epidermal barrier function more effectively than individual agents [29].
- Furthermore, Vitamin E protects Vitamin A from oxidative degradation during storage and after topical application, thereby improving formulation stability. Ginger-derived antioxidants further reinforce this protective effect by scavenging reactive oxygen species generated during inflammatory skin conditions.

Table 6. Comparative Pharmacological Activities of Vitamin A, Vitamin E and Ginger Extract

Parameter	Vitamin A	Vitamin E	Ginger Extract
Antioxidant activity	Moderate	Excellent	Excellent
Anti-inflammatory activity	Moderate	High	Very High
Collagen stimulation	Excellent	Moderate	Moderate
Wound healing	Excellent	High	High
Antimicrobial activity	Limited	Mild	Excellent
Anti-aging effect	Excellent	Excellent	Moderate
Skin hydration	Moderate	Excellent	Moderate
Photo-protection	Moderate	Excellent	Moderate
Stability	Low	Moderate	Moderate

Table 7. Major Bioactive Components and Their Pharmacological Activities

Bioactive Compound	Source	Major Pharmacological Activities
Retinol	Vitamin A	Cell differentiation, collagen synthesis
Retinoic acid	Vitamin A	Anti-aging, acne treatment
α -Tocopherol	Vitamin E	Antioxidant, membrane stabilization
γ -Tocopherol	Vitamin E	Anti-inflammatory
6-Gingerol	Ginger	Anti-inflammatory, antioxidant
6-Shogaol	Ginger	Antioxidant, antimicrobial
Zingerone	Ginger	Wound healing, anti-inflammatory
Paradol	Ginger	Anti-inflammatory

9. Rationale for Developing an Ethosomal Hydrogel Containing Vitamin A, Vitamin E and Ginger Extract

Despite the established therapeutic benefits of these bioactive agents, their clinical application is often limited by poor aqueous solubility, instability, susceptibility to oxidation, and inadequate penetration across the stratum corneum. Incorporating these compounds into an ethosomal hydrogel addresses these limitations by combining the penetration-enhancing properties of ethosomes with the sustained-release and patient-friendly characteristics of hydrogels. Such a formulation has the potential to improve drug stability, increase dermal retention, reduce dosing frequency, and enhance patient compliance. This integrated strategy may therefore offer a promising approach for the topical management of inflammatory skin disorders, wound healing, photoaging, and other dermatological conditions.

10. Formulation Development of Ethosomal Hydrogel

The development of an ethosomal hydrogel containing Vitamin A, Vitamin E, and ginger extract requires careful optimization of formulation variables to ensure high encapsulation efficiency, desirable vesicle size, stability, controlled drug release, and enhanced skin permeation. A systematic approach involving pre-

formulation studies, vesicle preparation, hydrogel incorporation, and physicochemical evaluation is essential for obtaining a stable and effective topical formulation.

10.1 Pre-formulation Studies

Pre-formulation studies provide essential information regarding the physicochemical properties of the active ingredients and excipients. Parameters commonly evaluated include:

- Organoleptic characteristics
- Solubility profile
- Melting point
- Partition coefficient (Log P)
- pKa determination
- UV-visible spectroscopic analysis
- Fourier Transform Infrared Spectroscopy (FTIR) for compatibility studies
- Differential Scanning Calorimetry (DSC)
- X-ray Diffraction (XRD)

These studies help identify potential incompatibilities and guide the selection of suitable formulation components.

10.2 Preparation of Drug-Loaded Ethosomes

The cold method is generally preferred for preparing ethosomes containing heat-sensitive vitamins and herbal extracts.

Typical formulation composition

Ingredient	Function
Soy phosphatidylcholine	Vesicle former
Ethanol	Penetration enhancer



Propylene glycol	Co-solvent
Vitamin A	Skin regeneration
Vitamin E	Antioxidant
Ginger extract	Anti-inflammatory
Purified water	Vehicle

After preparation, the ethosomal suspension is incorporated into a pre-neutralized Carbopol hydrogel under gentle stirring to obtain a homogeneous ethosomal hydrogel.

11. Evaluation of Ethosomal Hydrogel

Comprehensive characterization is essential to ensure formulation quality, stability, and therapeutic performance

11.1 Physical Appearance

The hydrogel should be evaluated for:

- Color
- Homogeneity
- Clarity
- Presence of phase separation
- Grittiness
- Consistency

A smooth, homogeneous, translucent gel without phase separation is generally considered acceptable.

11.2 pH Determination

Skin-compatible pH is essential to minimize irritation.

- Instrument: Digital pH meter
- Ideal pH range: 5.0–6.5

Maintaining physiological pH helps preserve skin barrier integrity and enhances patient acceptability

11.3 Vesicle Size and Polydispersity Index (PDI)

Particle size significantly influences skin penetration.

Measurement:

- Dynamic Light Scattering (DLS)

Ideal values:

- Particle size: 100–300 nm

- PDI: <0.30

Smaller and uniformly distributed vesicles exhibit superior dermal penetration.

11.4 Zeta Potential

Zeta potential indicates colloidal stability.

Ideal value:

- Greater than ± 30 mV

High absolute zeta potential prevents vesicle aggregation during storage.

11.5 Entrapment Efficiency

Entrapment efficiency determines the proportion of active compounds encapsulated within the ethosomes.

Entrapment Efficiency (%)

$$= \frac{\text{Total Drug} - \text{Free Drug}}{\text{Total Drug}} \times 100$$

Higher entrapment efficiency generally correlates with improved sustained release and therapeutic efficacy.

11.6 Drug Content

Drug content is determined using UV-visible spectrophotometry or High-Performance Liquid Chromatography (HPLC) to ensure uniform distribution of active ingredients within the formulation

11.7 Viscosity

Viscosity affects spreadability, patient compliance, and residence time.

Instrument:

- Brookfield Viscometer

An optimized viscosity ensures easy application without runoff.



11.8 Spreadability

Spreadability determines the ease of application over the skin surface.

$$S = \frac{M \times L}{T}$$

Where:

- S = Spreadability
- M = Weight tied to upper slide
- L = Length moved
- T = Time taken

Good spreadability enhances patient compliance and uniform drug distribution.

11.9 Extrudability

Extrudability evaluates the ease with which gel can be expelled from a collapsible tube.

Excellent extrudability indicates convenient patient use.

11.10 In Vitro Drug Release

Drug release studies are generally performed using:

- Franz Diffusion Cell

Membrane:

- Cellophane membrane
- Dialysis membrane

Release media:

- Phosphate buffer pH 7.4

Drug release kinetics may be analyzed using:

- Zero-order model
- First-order model
- Higuchi model
- Korsmeyer–Peppas model

11.11 Ex Vivo Skin Permeation Study

Animal skin (rat, porcine, or goat skin) mounted on a Franz diffusion cell is commonly used to assess:

- Drug permeation
- Drug deposition
- Flux
- Permeability coefficient

These studies simulate topical drug delivery under physiological conditions.

11.12 Stability Studies

Stability studies should follow ICH Q1A(R2) guidelines to evaluate formulation robustness under different storage conditions.

Parameters monitored include: Appearance, pH, Particle size, Entrapment efficiency, Drug content & Viscosity.

Table 8. Evaluation Parameters of Ethosomal Hydrogel

Parameter	Instrument/Method	Acceptance Criteria
Appearance	Visual inspection	Smooth, homogeneous
Ph	Digital pH meter	5.0–6.5
Particle size	Dynamic Light Scattering	100–300 nm
Polydispersity Index	DLS	<0.30
Zeta potential	Zeta analyzer	> ±30 mV
Drug content	UV/HPLC	95–105%
Entrapment efficiency	Centrifugation	>80%
Viscosity	Brookfield Viscometer	Optimized consistency
Spreadability	Glass slide method	Good
Drug release	Franz Diffusion Cell	Sustained release
Stability	ICH guidelines	No significant changes



12. Recent Advances in Ethosomal Hydrogel Research

Recent investigations have highlighted several innovations in ethosomal technology for dermal drug delivery:

- Development of **transethosomes**, incorporating both ethanol and edge activators to further enhance skin permeation.
- Use of **Quality by Design (QbD)** and **Design of Experiments (DoE)** for systematic optimization of formulation variables.
- Integration of **natural bioactive compounds**, including curcumin, resveratrol, quercetin, aloe vera, and ginger extract, into ethosomal systems for improved antioxidant and anti-inflammatory effects.
- Exploration of **stimuli-responsive hydrogels** (e.g., pH-, temperature-, or ROS-responsive systems) for controlled drug release.
- Application of advanced analytical tools such as **confocal laser scanning microscopy** and **Raman spectroscopy** to visualize vesicle penetration and drug distribution within skin layers.

These advances have broadened the potential of ethosomal hydrogels for treating inflammatory skin diseases, wound healing, fungal infections, acne, psoriasis, and photoaging,

13. CHALLENGES AND FUTURE PERSPECTIVES

Although ethosomal hydrogels show significant promise, several challenges remain:

- Long-term physical and chemical stability
- Oxidation of phospholipids and vitamins
- Large-scale manufacturing and process reproducibility
- Regulatory approval and quality standardization
- Limited clinical trials evaluating long-term safety and efficacy

- Cost-effective industrial production

Future research should focus on optimizing formulation stability, incorporating advanced nanocarrier technologies, conducting well-designed clinical studies, and developing scalable manufacturing processes. The application of artificial intelligence and machine learning for formulation optimization may further accelerate the translation of ethosomal hydrogels from laboratory research to clinical practice.

CONCLUSION

Ethosomal hydrogels represent a promising platform for enhanced dermal drug delivery by combining the penetration-enhancing properties of ethosomes with the sustained-release and patient-friendly characteristics of hydrogels. Incorporation of Vitamin A, Vitamin E, and ginger extract into an ethosomal hydrogel provides a multifunctional therapeutic approach that targets multiple aspects of skin health, including oxidative stress, inflammation, impaired collagen synthesis, and microbial contamination.

Vitamin A promotes epidermal renewal and collagen production, Vitamin E provides potent antioxidant protection and improves barrier function, while ginger extract contributes anti-inflammatory, antimicrobial, and wound-healing activities. Encapsulation within ethosomal vesicles enhances the stability and skin permeation of these bioactive agents, resulting in improved local bioavailability and therapeutic performance. Current evidence supports the potential of ethosomal hydrogels as advanced topical delivery systems for applications in wound healing, photoaging, inflammatory skin disorders, acne, and cosmetic dermatology. However, further optimization, comprehensive stability studies, and well-controlled clinical trials are necessary to establish their long-term safety, efficacy, and commercial feasibility. With continued advances in nanotechnology and formulation science,



ethosomal hydrogels are expected to play an increasingly important role in the development of next-generation dermal drug delivery systems.

REFERENCES

1. Barry BW. *Dermatological Formulations: Percutaneous Absorption*. New York: Marcel Dekker; 1983.
2. Prausnitz MR, Langer R. Transdermal drug delivery. **Nat Biotechnol**. 2008;26(11):1261–1268.
3. Benson HAE. Transdermal drug delivery: penetration enhancement techniques. **Curr Drug Deliv**. 2005;2(1):23–33.
4. Williams AC, Barry BW. Penetration enhancers. **Adv Drug Deliv Rev**. 2012;64(Suppl):128–137.
5. Elias PM. Skin barrier function. **J Invest Dermatol**. 2005;125(2):183–200.
6. Proksch E, Brandner JM, Jensen JM. The skin: an indispensable barrier. **Exp Dermatol**. 2008;17(12):1063–1072.
7. Bouwstra JA, Honeywell-Nguyen PL. Skin structure and mode of action of vesicles. **Adv Drug Deliv Rev**. 2002;54(Suppl 1):S41–S55.
8. Hadgraft J. Skin, the final frontier. **Int J Pharm**. 2001;224(1-2):1–18.
9. Touitou E, Dayan N, Bergelson L, Godin B, Eliaz M. Ethosomes—novel vesicular carriers for enhanced delivery: characterization and skin penetration properties. **J Control Release**. 2000;65(3):403–418.
10. Verma DD, Fahr A. Synergistic penetration enhancement effect of ethanol and phospholipids on the topical delivery of cyclosporin A. **Eur J Pharm Biopharm**. 2004;58(1):89–99.
11. Paiva-Santos AC, Silva ALR, Guerra C, et al. Ethosomes as nanocarriers for dermal and transdermal drug delivery. **Int J Pharm**. 2021;601:120571.
12. Ascenso A, Raposo S, Batista C, et al. Nanotechnology-based topical formulations: Current advances and future perspectives. **Drug Discov Today**. 2021;26(11):2681–2692.
13. Peppas NA, Bures P, Leobandung W, Ichikawa H. Hydrogels in pharmaceutical formulations. **Eur J Pharm Biopharm**. 2000;50(1):27–46.
14. Ahmed EM. Hydrogel: Preparation, characterization and applications. **J Adv Res**. 2015;6(2):105–121.
15. Caló E, Khutoryanskiy VV. Biomedical applications of hydrogels. **Eur Polym J**. 2015;65:252–267.
16. Mukherjee S, Date A, Patravale V, Korting HC, Roeder A, Weindl G. Retinoids in the treatment of skin aging. **Clin Interv Aging**. 2006;1(4):327–348.
17. Kang S, Fisher GJ, Voorhees JJ. Molecular mechanisms of retinoid action in human skin. **J Investig Dermatol Symp Proc**. 1998;3(1):53–57.
18. Griffiths CEM. The role of retinoids in the prevention and repair of aged skin. **Clin Exp Dermatol**. 2001;26(7):613–618.
19. Thiele JJ, Ekanayake-Mudiyanselage S. Vitamin E in human skin. **Mol Aspects Med**. 2007;28(5-6):646–667.
20. Briganti S, Picardo M. Antioxidant activity, lipid peroxidation and skin diseases. **J Eur Acad Dermatol Venereol**. 2003;17(6):663–669.
21. Darr D, Combs S, Dunston S, Manning T, Pinnell S. Topical vitamin C protects porcine skin from ultraviolet radiation-induced damage. **Br J Dermatol**. 1992;127(3):247–253.
22. Puglia C, Bonina F. Lipid nanoparticles as novel delivery systems for topical retinoids. **Expert Opin Drug Deliv**. 2012;9(4):429–441.



23. Mashhadi NS, Ghiasvand R, Askari G, Hariri M, Darvishi L, Mofid MR. Anti-oxidative and anti-inflammatory effects of ginger. **Int J Prev Med.** 2013;4(Suppl 1):S36–S42.
24. Semwal RB, Semwal DK, Combrinck S, Viljoen A. Gingerols and shogaols: Important phytochemicals. **Phytochemistry.** 2015;117:554–568.
25. Ali BH, Blunden G, Tanira MO, Nemmar A. Some phytochemical, pharmacological and toxicological properties of ginger. **Food Chem Toxicol.** 2008;46(2):409–420.
26. Grzanna R, Lindmark L, Frondoza CG. Ginger—An herbal medicinal product with broad anti-inflammatory actions. **J Med Food.** 2005;8(2):125–132.
27. Kundu P, Das M, Tripathy K. Herbal nanocarriers in topical drug delivery: Current perspectives. **Drug Deliv Transl Res.** 2022;12(8):1872–1893.
28. Souto EB, Cano A, Martins-Gomes C, et al. Nanoparticles for topical drug delivery. **Pharmaceutics.** 2022;14(5):958.
29. Patravale VB, Mandawgade SD. Novel cosmetic delivery systems. **Int J Cosmet Sci.** 2008;30(1):19–33.
30. Cevc G. Lipid vesicles and other colloids as drug carriers on the skin. **Adv Drug Deliv Rev.** 2004;56(5):675–711.
31. Honeywell-Nguyen PL, Bouwstra JA. Vesicles as a tool for transdermal drug delivery. **Drug Discov Today Technol.** 2005;2(1):67–74.
32. Gupta PN, Mishra V, Rawat A, Dubey P, Mahor S, Jain S, et al. Non-invasive vaccine delivery using lipid vesicles. **J Control Release.** 2005;100(2):255–266.
33. Jain S, Umamaheshwari RB, Bhadra D, Jain NK. Ethosomes: A novel vesicular carrier. **Drug Deliv.** 2003;10(4):291–295.
34. Song CK, Balakrishnan P, Shim CK, Chung SJ, Chong S, Kim DD. A novel vesicular carrier for enhanced skin delivery. **Int J Pharm.** 2012;428(1-2):44–52.
35. Dragicevic N, Maibach HI. **Percutaneous Penetration Enhancers Chemical Methods in Penetration Enhancement.** Berlin: Springer; 2015.
36. Baki G, Alexander KS. **Introduction to Cosmetic Formulation and Technology.** Hoboken: Wiley; 2015.
37. Aulton ME, Taylor KMG. **Aulton's Pharmaceutics: The Design and Manufacture of Medicines.** 6th ed. London: Elsevier; 2022.
38. Sinko PJ. **Martin's Physical Pharmacy and Pharmaceutical Sciences.** 7th ed. Philadelphia: Wolters Kluwer; 2017.
39. Allen LV. **Ansel's Pharmaceutical Dosage Forms and Drug Delivery Systems.** 11th ed. Philadelphia: Wolters Kluwer; 2020.
40. Rowe RC, Sheskey PJ, Quinn ME. **Handbook of Pharmaceutical Excipients.** 9th ed. London: Pharmaceutical Press; 2020.
41. ICH Harmonised Guideline. Q1A(R2): Stability Testing of New Drug Substances and Products. International Council for Harmonisation; 2003.
42. ICH Harmonised Guideline. Q8(R2): Pharmaceutical Development. International Council for Harmonisation; 2009.
43. European Pharmacopoeia Commission. **European Pharmacopoeia.** 11th ed. Strasbourg: Council of Europe; 2023.
44. United States Pharmacopeial Convention. **United States Pharmacopeia 47–National Formulary 42.** Rockville (MD): USP; 2024.
45. Indian Pharmacopoeia Commission. **Indian Pharmacopoeia.** 9th ed. Ghaziabad: IPC; 2022.
46. Souto EB, Zielinska A, Ferreira NR, et al. Lipid-based nanocarriers for topical administration. **Pharmaceutics.** 2020;12(12):1193.



47. Mura S, Nicolas J, Couvreur P. Stimuli-responsive nanocarriers for drug delivery. **Nat Mater.** 2013;12(11):991–1003.
48. Gupta R, Rai B. Nanotechnology in cosmeceuticals: Current trends and future prospects. **J Cosmet Dermatol.** 2023;22(4):1187–1199.
49. Torchilin VP. Multifunctional nanocarriers. **Nat Rev Drug Discov.** 2014;13(11):813–827.
50. Sharma G, Sharma AR, Lee SS, Chakraborty C. Advances in nanocarrier-based topical drug delivery systems. **Pharmaceutics.** 2024;16(2):214.

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