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Review Article

Garcinol In Preclinical Cancer Models: A Systematic Review of Mechanisms, Safety, And Nanodelivery Strategies

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ABSTRACT

Background: Garcinol, a polyisoprenylated benzophenone predominantly derived from *Garcinia indica*, has gained significant attention due to its wide-ranging pharmacological activities. Traditionally used in Ayurvedic medicine, garcinol exhibits potent antioxidant, anti-inflammatory, antimicrobial, and anticancer properties. Its multitarget potential makes it an emerging candidate for preclinical drug development. **Methods:** A systematic survey of the literature was performed across PubMed, Scopus, Web of Science, and Google Scholar up to August 2025. Studies were selected based on predefined inclusion criteria encompassing in vitro and in vivo preclinical evaluations of garcinol. Data were extracted regarding disease model, dose, route of administration, pharmacological outcomes, toxicity, and underlying molecular mechanisms. **Results:** Garcinol demonstrated consistent anti-cancer effects across multiple malignancies including breast, colon, prostate, pancreatic, and lung cancers. Mechanistically, it inhibits NF- κ B, STAT3, and histone acetyltransferases, while promoting apoptosis, cell cycle arrest, and mesenchymal-to-epithelial transition in tumor cells. Its antioxidant and anti-inflammatory activities are mediated through suppression of reactive oxygen species, COX-2, and pro-inflammatory cytokines. Beyond oncology, garcinol shows promise in neuroprotection, cardiovascular modulation, antimicrobial defense, and anti-ulcer activity. Preclinical pharmacokinetic studies reveal moderate oral bioavailability, high tissue distribution, and minimal hepatic first-pass metabolism, though limited aqueous solubility remains a barrier. Toxicity profiles in rodents indicate high tolerability with no significant adverse effects. Emerging nanotechnology-based formulations including PLGA, gold nanoconjugates, and protein nanocarriers substantially improve solubility, stability,

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and therapeutic efficacy. Conclusion: Preclinical evidence positions garcinol as a multitargeted natural compound with broad therapeutic potential and favorable safety. However, translation to clinical application requires addressing formulation challenges, establishing human pharmacokinetics, and conducting well-designed clinical trials.

INTRODUCTION

Polyisoprenylated benzophenones are biologically dynamic natural products principally exposed in plants of the *Clusiaceae* family. These small molecules have numerous biological activities, including anti-inflammation and cytotoxicity towards cancer cells.[1] The anti-inflammatory mechanisms of polyisoprenylated benzophenones suggest that they target the TLR/IRAK-1 pathway to inhibit downstream NF- κ B and Akt/mTOR pathways, inhibit lipoxygenase enzyme activity and suppress nitric oxide production in LPS-stimulated RAW 264.7 macrophages. [2-4]

Benzophenones are structurally characterized as diphenylmethanone, but one or more hydrogen atoms from the phenolic groups have been substituted with hydroxyl, alkyl, alkyloxy groups, or halogen.[5] These compounds are a basis of significant interest in drug development and have been utilized as treatments for cancer, inflammation, antibacterial, and antiparasitic compounds. The demand for original and more effective treatments has led to research into the newly coined polyisoprenylated benzophenones.[6]

Functionally, these benzophenones are important intermediates in the xanthone biosynthesis pathway, a class of bioactive compounds that have mainly been reported in the *Clusiaceae* family of plants.[7] Their relative scarcity, aside from this botanical group, illustrates their unique biochemical function. Also, studies show that benzophenones can bind to tubulin inhibiting

microtubule disassembly during cell division, parallel to paclitaxel but not as potent; and a number of studies have shown that benzophenones activate caspase-3 activity and induce apoptosis. Proposed mechanisms include:

- Inhibition of microtubule disassembly
- Suppression of histone acetyltransferase activity
- Inhibition of protease function
- Modulation of kinase activity
- Prooxidant activity leading to DNA damage.[8]

1.1 Garcinol: Discovery, Structural Features, and Multifaceted Bioactivity

1.1.1 Discovery and Traditional Use

Garcinol, a polyisoprenylated benzophenone, is a plant compound that originates from the fruit rinds of *Garcinia indica* (kokum) and other related species. The solvent (fruit rinds) has been used for a long time by traditional cultures in tropical areas (especially ancient and modern uses in Ayurvedic medicine) to treat a variety of ailments, including dermatitis, diarrhea, and digestive problems.[9][10] The compound gained scientific consideration due to its potential biological properties, which have only recently been clarified in detail.[9][11]

1.1.2 Chemical Structure and Analysis

Garcinol has the molecular formula $C_{38}H_{50}O_6$ and is classified as a polyisoprenylated benzophenone. Its molecular weight, as measured by QTOF LC-ESI-MS/MS analysis, is roughly 603.37 g/mol.[12] The compound's structure includes a benzophenone core with isoprenyl side chains, which contribute to its hydrophobicity and biological activity.[13][14] Its pleiotropic actions



and interaction with cellular targets depend on the presence of these isoprenyl groups.[9][15]

1.1.3 Bioactivity and Mechanisms

Garcinol has a multitude of bioactivities reported in the literature including, but not limited to, antioxidant, anti-inflammatory, anti-cancer, and antimicrobial. Importantly, its antioxidant activity has been documented and garcinol has been shown to clear free radicals and prevent oxidative stress to living tissues. [9][10] The racemic compound's anti-cancer properties are thought to modulate many key regulatory pathways including but not limited to the inhibition of nuclear factor- κ B (NF- κ B), signal transducer and activator of transcription 3 (STAT3), and histone acetyltransferases (HATs).[15][16] Garcinol negatively regulates apoptosis, cellular cycle, and

cellular replication in many cell lines including breast, colon, and prostate cancer cells.[12][16] In addition to the functional mechanisms of its anti-cancer properties, garcinol has anti-inflammatory properties by inhibiting the activity of 5-lipoxygenase (5-LOX) and cyclooxygenase-2 (COX-2) which are key enzymes often quoted for their role in inflammatory response.[13][15] Garcinol is stated having anti-microbial properties demonstrated towards pathogenic bacteria and parasites, further suggesting its functional role as a multi-target therapeutic agent.[13]

1.2 Mechanisms of Action

The mechanisms underlying garcinol's bioactivity are diverse and involve multiple cellular pathways. Key mechanisms include:

Pharmacological Activity	Mechanism of action	Key Molecular Targets	Scientific Evidence
Antioxidant Activity [9][11]	Scavenges free radicals-Protects against oxidative stress	Superoxide anion, Hydroxyl radicals, DPPH radicals	Garcinol exhibits significant antioxidant activity, comparable to DL- α tocopherol, by neutralizing various free radicals.
Anti-Cancer Effects [12][16]	Induces apoptosis-Arrests cell cycle progression, Modulates oncogenic signaling pathways	NF- κ B, STAT3, PI3K/Akt pathways	Studies demonstrate that Garcinol induces apoptosis and cell cycle arrest in cancer cells by downregulating COX-2 and NF- κ B.
Anti-Inflammatory Effects [13][15]	Inhibits pro-inflammatory enzymes and Suppresses cytokine production	COX-2, Interleukin-6 (IL-6), TNF- α	Garcinol reduces the production of pro-inflammatory cytokines and mediators, including IL-6 and COX-2, in activated macrophages.
Epigenetic Regulation [17]	Inhibits histone acetyltransferases (HATs), Modulates gene expression	p300, PCAF, miRNAs	Garcinol acts as a natural HAT inhibitor, leading to chromatin remodelling and regulation of gene expression.



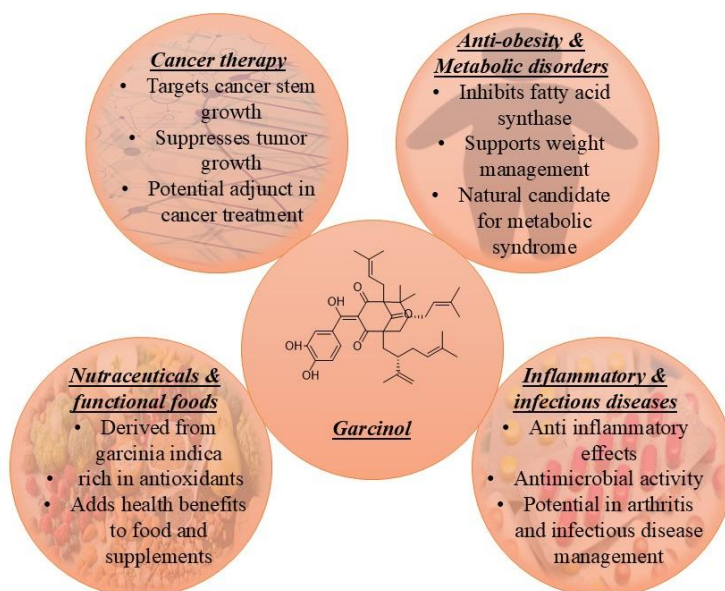


Figure 1. Potential applications in medicine and industry [10][13][14][15][18][19][20]

1.3 Pharmacokinetics and Delivery Systems

Researchers have newly studied the pharmacokinetics and biodistribution of garcinol in order to enhance its therapeutic use. Garcinol has moderate bioavailability and metabolic stability, as a large part escapes hepatic first-pass metabolism [21]. To further enhance its therapeutic efficacy, researchers have attempted different delivery systems, including a new delivery system based on bovine serum albumin (BSA)-based nanocarriers which enhances garcinol's solubility and prolonged release kinetics [22]

1.4 Natural Products in Preclinical Research

- Plant-based natural products continue to serve as a primary source of bioactive compounds in drug discovery, and with their inherent complexity of chemical structures and biological activity, are integral in the development of new therapeutics. The integration of traditional knowledge of plant-based natural products with modern analytical and computational tools have vastly improved

the identification and characterization of these compounds leading to their application in drug discovery and preclinical screening. Use of phytochemicals such as alkaloids, terpenoids, phenolics and cardiac glycosides have been explored widely, for their potential therapeutic properties against a variety of diseases, such as cancer and diabetes, as well as cardiovascular effects.[23][24] Modern techniques such as molecular docking, quantitative structure activity relationship modeling (QSAR), and machine-learning have improved our ability to predict how plant-based natural products interact with biological targets, which would facilitate the drug discovery process.[25][26] In addition, metabolomic and high-throughput assessments are offering new ways to isolation and characterization of bioactive compounds that enabled the exploration to pharmacokinetics and pharmacodynamics.[27][28] Moreover, polypharmacology, where plant extracts exert effects on multiple targets, is emerging to develop drug in a more holistic manner.[29] Furthermore, bioinformatics and

computational tools decreased the time and cost when compared to traditional drug development, making the entire process more seamless in discovering therapeutic compounds or candidates.[26] The therapeutic potential of plant-derived compounds is also credited to reduced side effects associated with synthetic drugs, which makes them promising candidates for development.[23][30] As the pharmaceutical industry continues to explore these natural resources, existing and emerging interdisciplinary approaches and enhanced technologies will maximize the potential of plant-based natural products in drug discovery.[31][32]

Objectives: To systematically summarize and critically evaluate all reported preclinical studies involving Garcinol, covering multiple therapeutic categories, mechanisms, models, and translational implications.

2. Methodology

2.1 Literature Search Strategy

A comprehensive literature search was performed to identify all relevant studies investigating the preclinical pharmacology of garcinol. The following electronic databases were systematically searched: PubMed, Scopus, Web of Science, and Google Scholar, covering all available records up to 2024. The search strategy incorporated a combination of keywords and Boolean operators, including: “Garcinol,” “preclinical,” “in vivo,” “animal models,” “mechanism,” and “pharmacology.” To enhance coverage, reference lists of retrieved articles and relevant reviews were also manually screened.

2.2 Inclusion and Exclusion Criteria

Studies were eligible for inclusion if they met the following criteria:

Articles reporting either in vitro or in vivo preclinical studies.

- Subjects/models: Animal models or established cancer/biological cell lines.
- Endpoints reported: Pharmacological activity, mechanism of action, pharmacokinetics, or safety evaluations of garcinol. are-reviewed articles.

Exclusion criteria included:

- Clinical or human studies.
- Editorials, commentaries, or conference abstracts.

Studies without sufficient mechanistic or pharmacological data.

- Non-English publications.

2.3 Data Extraction and Management

Data from eligible studies were independently extracted by [insert researcher initials or “two reviewers” if being written like a systematic review] using a pre-designed data extraction form. Extracted information included:

- Disease model / biological system (animal species, cell type, or in vivo model)
- Dosage and formulation (dose level, delivery system, formulation strategy)
- Administration route and duration of treatment
- Primary outcomes (therapeutic effects, efficacy, toxicity, survival, tumor weight, inflammatory markers, etc.)
- Mechanistic insights (molecular pathways, targets, and signaling cascades affected by garcinol)



- Combination strategies, where applicable (e.g., co-administration with chemotherapeutics).

2.4 Study Selection Process

Titles and abstracts were screened for relevance, followed by full-text evaluation of potentially eligible studies. Any discrepancies during selection or data extraction were resolved through discussion and consensus among reviewers.

3. Pharmacological and Chemical Overview of Garcinol

3.1 Source and chemical structure

Garcinol is a polyisoprenylated benzophenone most commonly isolated from the dried rind of *Garcinia indica*, the tropical fruit of the *Garcinia* tree distributed in Southeast Asia and Central Africa.[14][33][34] The garcinol structure is encompassed of a phenolic catechol group and three beta-diketone groups, which are mainly responsible for its marked antioxidant and anti-inflammatory activities. Its exclusive structure allows garcinol to display a variety of biological activities, including anticancer and antithrombotic activity. Garcinol is an integrin $\alpha\text{IIb}\beta 3$ inhibitor that is hepatoprotective against non-steroidal anti-inflammatory drugs and aspirin, and shows inhibition of platelet aggregation and thrombus formation without prolongation of bleeding time, indicating its development as a new generation of antithrombotic agent.[33] Additionally, in non-small cell lung cancer, garcinol target cancer stem cells through induction of cellular senescence, apoptosis, and cell cycle arrest.[14] In breast cancer research, garcinol is an acetyltransferase inhibitor that inhibits estrogen-induced proliferation, and promotes apoptosis in MCF-7 cells via effects on acetylation and modulation of the NF- κ B pathway.[34] Overall, the variety of

bioactivities potentially indicates garcinol's place as a therapeutic agent in many areas of medicine.

3.2 Key physicochemical properties

Garcinol is a polyisoprenylated benzophenone extracted from *Garcinia indica*, that shows good pharmacological properties with both anticancer and anti-inflammatory effects. Furthermore, it is characterized by many physicochemical properties, including its extreme insolubility in water, which complicates translational application of Garcinol in the clinical setting. However, new formulations, such as PLGA nanoparticles, have been developed to minimize the effects of this insolubility and increase bioavailability and activity.[35][36] Garcinol's biological activity is mainly associated with its effects as an antioxidant, which has been widely studied, and used in traditional medicine for use in many conditions for thousands of years.[9] Several anti-cancer properties of Garcinol in various cancers, such as colon, breast, prostate, head and neck cancers, and hepatocellular carcinomas have been shown. The mechanism of action of Garcinol inhibits cellular activity by altering transcription factors NF- κ B and JAK/STAT3 regulation in tumor cells, thereby inhibiting the growth of populations of malignant cells. However, even though Garcinol has shown promise as a clinically relevant anti-cancer agent, its translational submission remains in pre-clinical trials as it is limited by an inability to definitely evaluate pharmacological parameters; [16] Furthermore, it is also a histone acetyl transferase inhibitor, which can have effects on epigenetic regulation and shows great promise against breast and pancreatic cancer.[37] The ongoing research into garcinol's precise metabolic mechanism underscores its potential significance in the field of natural therapeutic agents.[16]

3.3 Preclinical ADME Profile



3.3.1 Stability

Garcinol also showed a medium level of metabolic stability in human liver microsomes (HLM), with an observed half-life of about 39 minutes and intrinsic clearance (CL_{int}) of 33.94 $\mu\text{L}/\text{min}/\text{mg}$ microsomal protein. It can be characterized as medium clearance drug and is close to the high clearance category. Because of this, we can conclude that garcinol will require better stability to reach steady blood concentrations over an extensive time period. Stability can be improved via chemical modifications (deuteration, halogens/isosteric groups).[21]

3.3.2 Solubility

Garcinol is hydrophobic and nearly insoluble in aqueous solutions but freely soluble in organic solvents, including methanol and acetonitrile. In addition, we identified acetonitrile with 0.1% formic acid as the ideal solvent for chromatographic analysis due to its sensitivity and resolution.[21]

3.3.3 Metabolism

Although garcinol is exposed to limited hepatic first-pass metabolism, out of the total dose 94% was released into systemic circulation. The metabolism of garcinol occurs via HLM and follows first order kinetics and we assessed the metabolic stability using a logarithmic linear plot and first order rate equation approach arriving at similar conclusions. The lipophilicity of the drug ($\log P = 6.85$) contributes to the high volume of distribution and suggests partitioning into the lipid bilayers and intracellular organelles. Overall garcinol is likely to have promising pharmacokinetic properties for future development as an anticancer agent.[21]

3.4 Safety evaluations

The toxicity profile of Garcinol from *Garcinia indica* was assessed through acute, sub-acute, sub-chronic, and reproductive/developmental toxicity studies in experimental rodents. Notable results are:

- **Acute Toxicity:** In female Wistar rats, a very high single dose (2000 mg/kg) was administered by oral gavage without significant clinical signs, mortality, or toxicity observed; gave it GHS category 5/unclassified considerations on safety to human health.[38]
- **28-Day Sub-Acute Toxicity:** No clinical signs of toxicity or mortality with no significant changes in body weight, body temperature, food consumption, and haematological, biochemical, or histopathological parameters were observed. Recovery groups showed no adverse effects. [38]
- **90-Day Sub-Chronic Toxicity:** There were no abnormal clinical signs, mortality, or significant changes in body weight, organ weight or histopathology; very minor changes in some haematology and biochemical parameters were detected, which were well within physiological limits. The No Observed Adverse Effect Level (NOAEL) was determined to be 100 mg/kg/day.[38]
- **Reproductive/Developmental Toxicity:** There were no adverse effects on reproductive parameters including pregnancy consequences, pup survival, organ histopathology. The body weights of dams and pups were comparable to controls and no abnormalities were stated. Based on the studies, Garcinol revealed a low toxicity profile in rodents and there were no observed adverse effects in the experimental conditions applied.[38]

3.5 Delivery systems used in preclinical studies



In preclinical studies, various delivery systems for garcinol, a potential anticancer agent, have been developed mainly with nanoparticle formulations. One study adopted an approach that utilized hyaluronic acid (HA) and poly(lactic-co-glycolic acid) (PLGA) to develop HA-coated garcinol-loaded nanoparticles (HA-GA-NPs). The HA-GA-NPs not only enhanced the antiproliferative effects of garcinol in breast cancer stem cells under hypoxic conditions, but were able to improve cellular uptake through receptor-mediated endocytosis.[39] One study adopted an iRGD peptide-conjugated.

biodegradable nanoparticles formulation, which significantly improved the cytotoxic effects of garcinol against colorectal carcinoma cells, as evidenced by a 2.3-fold change in IC₅₀ values of garcinol when compared with free garcinol.[40] Next, PLGA nanoparticles that were loaded with garcinol and emulsified with vitamin E TPGS exhibited significant in vitro cytotoxicity in multiple cancer cell lines, while also exhibiting targeted tumor uptake in vivo.[36] There is a great potential for using nanoparticles to enhance the

anticancer efficacy for garcinol. These studies can demonstrate that targeting nanocarriers may play a large role in improving the efficacy of drug delivery in cancer therapies.

4.Preclinical Studies by Therapeutic Area

Oncology is an important area of therapeutic research which comprises the study, diagnosis, treatment and prevention of cancer. Cancer is one of the leading causes of death around the world, and therefore a very active area of research and drug development. Garcinol, which is a polyisoprenylated benzophenone derivative, has become of interest in medicinal chemistry and cancer research because of its reported anti-oxidative, anti-inflammatory and anti-cancer activities [16][41]. Several in vitro and in vivo studies highlight the potential of Garcinol against a variety of cancers including breast, colon, pancreatic, leukemia and glioma [42]

4.1 Summary of *In Vitro* Studies Investigating Garcinol in Cancer

Cancer Type	Cell line used	Key Findings	Molecular Mechanisms	Combination Therapy	Reference
Breast Cancer	MCF-7 human breast cancer cell line	- Inhibited E2-induced proliferation (time- and dose-dependent) - Induced G0/G1 cell cycle arrest - Increased apoptosis in E2-treated cells.	↓ ac-H3 and ac-p65 levels - Inhibited NF-κB/p65 nuclear translocation ↓ CyclinD1, Bcl-2, Bcl-xl expression.	Not tested	[34]



Breast Cancer	Triple-negative breast cancer cell lines: MDA-MB-231 and BT-549	- Garcinol induced mesenchymal-to-epithelial transition (MET) in aggressive breast cancer cells. - It inhibited cell invasion. Garcinol induced apoptosis in breast cancer cells.	↓ NF-κB activity ↑ miR-200 and let-7 family ↓ nuclear β-catenin, ↑ p-β-catenin ↑ E-cadherin, ↓ vimentin, ZEB1 and 2	Not tested	[43]
Breast Cancer	Breast cancer stem cells (bCSCs) of MDA-MB-231 cells; hypoxic conditions	HA-coated garcinol-loaded nanoparticles (HA-GA-NPs) showed superior antiproliferative effects, enhanced cellular uptake, and induced apoptosis more effectively than free garcinol or uncoated NPs.	Significant downregulation of hypoxia-inducible factors (HIF-1α, HIF-2α) and notch pathway genes (DLL1, Jagged1); activation of caspase-3/7-dependent apoptosis	Not tested	[39]
Breast Cancer	MDA-MB-231 breast cancer cells (ER-); MCF-10A normal breast epithelial cells	Garcinol inhibited nicotine-induced proliferation of MDA-MB-231 cells in a dose-dependent manner; no cytotoxicity to normal MCF-10A cells	- Down-regulation of α9-nAChR and cyclin D3 expression; - G0/G1 cell cycle arrest via upregulation of p53 and p21; - Inhibition of AP-1 (c-Jun)-mediated cyclin D3 transcription.	Not tested	[44]
Breast Cancer	MDA-MB-231, MCF-7 (breast cancer)	Garcinol inhibited proliferation and induced apoptosis	- Downregulation of NF-κB, upregulation of caspase-3; - Suppression of cyclin D3; α9-nAChR downregulation (nicotine model)	Not tested	[42]

Breast Cancer	4T1 murine breast cancer cells	Garcinol sensitized 4T1 cells to Taxol, enhancing antitumor and anti-metastasis effects	• Inhibition of caspase-3/iPLA2 signaling; • Suppression of NF-κB/Twist1 pathway; • Downregulation of VEGF, MMP-2, MMP-9, TIMP-1; • Upregulation of E-cadherin.	• Combination with Taxol showed synergistic effects (CI < 1.0); • Enhanced G2/M arrest, • Reduced metastasis markers	[45]
Oral Cancer	Oral squamous cell carcinoma (OSCC) cell lines: SCC-4, SCC-9, SCC-25	• Garcinol inhibited cell proliferation and clonogenic survival in a dose-dependent manner • Induced apoptosis; caused S-phase cell cycle arrest; minimal toxicity on PBMCs	• Downregulation of NF-κB and COX-2; • Reduced VEGF expression; • Inhibition of NF-κB nuclear translocation.	Not tested	[44]
Oral Cancer	Oral squamous cell carcinoma (OSCC) cell line: SCC-25	8-Allyl garcinol significantly reduced cell viability and clonogenicity in SCC-25 cells in a dose-dependent manner.	• Induced apoptosis via modulation of mitochondrial membrane potential and increased ROS generation; • Downregulation of anti-apoptotic Bcl-2, upregulation of pro-apoptotic Bax and cleaved caspase-3.	Not tested	[46]

Lung Cancer	A549, H1299, H460, H1650, H358, HCC827, H441.	• Downregulation of Wnt/β-catenin/STAT3 pathway through: • Impaired phosphorylation of LRP6. • Decreased expression of Axin2, β-catenin, Dvl2, and cyclin D1.	• Inhibits cancer cell growth and proliferation. • Shows anti-proliferative responses. • Inhibited cell proliferation/suppressed cell viability.	Not tested	[16]
Colon Cancer	HT-29 human colon cancer cells	Garcinol exhibits anti-proliferative, anti-angiogenic, anti-migratory, and pro-apoptotic effects on HT-29 cells.	• Garcinol suppresses mPGES-1/PGE2/HIF-1α signaling pathways, leading to reduced expression of VEGF, CXCR4, MMP-2, and MMP-9, and increased caspase 3	Not tested	[47]
Pancreatic cancer	BxPC-3, PANC-1, PaCa	Garcinol inhibits proliferation and induces apoptosis effectively.	• Modulation of NF-κB, VEGF, PARP, MMPs, interleukins, and caspases expression.	• Garcinol sensitized PaCa cells to gemcitabine treatment. • Cumulative effect induced apoptosis and inhibited cell proliferation. • Synergistic growth inhibition was observed with combination	[48],[49]

				of curcumin treatment inhibited BxPC-3 and Panc-1 cells in dose-dependent manner.	
Prostate cancer	LNCaP, C4-2B, PC3	Garcinol inhibits cell growth and induces apoptosis in a dose-dependent manner.	Down-regulation of NF- κ B signaling and its target genes (Bcl-2, Bcl-xL); caspase-3 activation.	Not tested	[49]
Leukaemia	HL-60, NB4, U937, K562	Garcinol exhibits growth inhibitory effects, induces apoptosis, and disrupts mitochondrial membrane potential.	Activation of caspase-3, caspase-9, and Bax; degradation of PARP; downregulation of Bcl-2; disruption of mitochondrial transmembrane potential.	Not tested	[42]
Glioma	C6 (rat glioblastoma)	Garcinol inhibited cell viability, induced apoptosis, and was non-toxic to normal cells	Inhibition of NF-κB signalling, downregulation of Bcl-2, Bcl-XL, Survivin, Cyclin D1, activation of caspase-3/9, increased ROS.	Not tested	[50]

5 Summary of *In Vivo* Studies Investigating Garcinol in Cancer

Cancer Type	Model Used	In Vivo System	Key Findings	Molecular Mechanisms	Combination Therapy	Reference
Breast Cancer (TNBC)	MDA-MB-231	Xenograft in SCID mice	↓ Tumor growth ↓ Proliferation (Ki-67) and vascular density (CD31) - EMT reversal (↑ E-cadherin, ↓ vimentin)	↓ NF-κB and nuclear β-catenin ↑ miR-200, let-7 family ↓ Cyclin D1, ↑ GSK-3β	Not tested	[43]



Breast Cancer (TNBC)	MDA-MB-231	SCID mice (female, ICR)	• ↓ Tumor growth significantly ($p < 0.05$) • No observed systemic toxicity • ↓ Invasion (validated in vitro as well)	• ↓ Total and phosphorylated STAT-3 (Tyr705 and Ser727) • ↓ IL-6-induced STAT-3 activity • ↓ STAT-3 target genes: VEGF, MMP-9, uPA • ↓ Angiogenesis and metastatic potential due to target downregulation	Not tested	[43]
Breast Cancer (TNBC murine breast cancer cells)	4T1	Orthotopic model in BALB/c mice	• ↓ Tumor volume and weight • ↓ Lung and spleen metastasis • ↓ Proliferation (Ki-67), angiogenesis (CD31), MMP-2/-9, VEGF • ↑ E-cadherin • No systemic toxicity observed	• ↓ NF-κB/Twist1 signaling • ↓ caspase-3/iPLA2 • ↓ PGE2 • ↓ EMT markers (Twist1, Snail, ZEB1) • ↑ IκBα, ↓ p-NF-κB • ↓ Cyclin A2, Cdc25A, Cdc2, Bcl-2	Taxol (5 mg/kg i.p. + 1 mg/kg garcinol oral) — synergistic effect (CI < 1)	[45]

Lung Cancer	Human NSCLC cell lines A549 and H441	Immunodeficient NOD/SCID mice	Garcinol significantly inhibits tumor growth and stem cell-like properties of NSCLC in vivo, reducing cancer stem cell populations and tumorigenicity without causing systemic toxicity.	- Wnt/β-catenin and STAT3 pathway suppression; $\downarrow$ CSC markers and tumorigenicity	STAT3-siRNA + garcinol: Greater reduction in H441 cell viability than either treatment alone; indicates potential synergy for targeting STAT3 with garcinol.	[52]
Tongue Squamous Cell Carcinoma	4-NQO-induced carcinogenesis	Fischer 344 rats	- Dietary garcinol significantly reduced the incidence and multiplicity of tongue tumors (papillomas and SCCs), as well as preneoplastic lesions. - No toxicity was observed.	Garcinol suppressed Cell proliferation markers (BrdU-labeling index, Cyclin D1 expression) COX-2 expression in preneoplastic and neoplastic lesions.	Not tested	[53]
Prostate Cancer	Human prostate cancer cell line PC-3	Nude mice xenograft model	Garcinol treatment led to over 80% tumor reduction in volume; apoptosis was significantly induced in tumor tissues.	- Induced apoptosis via increased Bax/Bcl-2 ratio, caspase-3/-9 activation, PARP and DFF-45 cleavage. - Inhibited autophagy via activation of PI3K/Akt/mTOR and GSK-3β pathways.	Not tested	[54]

				Suppressed anti-apoptotic proteins (Bcl-2, Bcl-xL)		
Colorectal Cancer	Human colorectal cancer cell lines (HCT116, SW480)	Nude mice xenograft model	Garcinol significantly inhibited tumor growth and tumor weight in vivo in a dose-dependent manner without noticeable toxicity.	Garcinol inhibited the Wnt/β-catenin pathway by: - Reducing β-catenin levels - Decreasing downstream targets (c-Myc, Cyclin D1, Survivin) - Enhancing GSK-3β expression (a β-catenin suppressor)	Not tested	[40]

6. Safety, Pharmacokinetics, and Delivery Challenges

6.1 Safety Profile and Toxicity Studies

6.1.1 Comprehensive Toxicity Assessment in Rodents

The safety profile of garcinol has been thoroughly assessed through repeated toxicity studies according to OECD guidelines.[55] Acute toxicity studies in female Wistar rats showed virtually no toxicity, with no observed abnormal clinical signs, abnormal behavior, or mortality in Wistar rats given 2000 mg/kg doses during the 14-day observations.[55]

The 28-day repeated dose toxicity study tested garcinol for repeated dosing in Wistar rats at 20, 50, and 100 mg/kg/day.[55] All rats had no visible clinical signs of toxicity, had no evidence of

mortality, and did not show changes in weight gain, haematological parameters or markers, or biochemical markers across all treatment groups.[55]

The 90-day repeated dose toxicity study confirmed the safety profile of garcinol, as a full set of analyses did not show any dose-related toxicity from garcinol.[55] The reproductive and developmental toxicity studies demonstrated that garcinol does not have a deleterious impact on fertility, pregnancy or fetal development.[55] Collectively, it is clear that garcinol has an exceptional safety profile that presents no adverse effects under experimental conditions [55]

6.1.2 Cell Line Safety Studies

In vitro studies have shown that garcinol is selectively toxic to cancer cells, while having limited effects on normal cells. Garcinol inhibits



viability in hepatocellular carcinoma, lung carcinoma, and renal carcinoma cells while sparing viability in normal cells. [16] This selective cytotoxicity may suggest a favorable therapeutic index for garcinol in clinical settings.

6.2 Pharmacokinetic Studies

6.2.1 Oral Bioavailability and Absorption

Garcinol's absorption characteristics have been informatively studied in Sprague-Dawley rats through pharmacokinetic studies.[21] After oral dosing of 22.5 mg/kg and 45 mg/kg doses of garcinol, moderate bioavailability has been reported, at $26.64 \pm 0.23\%$ and $35.72 \pm 0.97\%$, respectively.[21] Dose-dependent bioavailability suggests that garcinol has an initial first-order pharmacokinetics with a non-linear component likely due to dose-dependent saturation of elimination routes at higher doses.[21]

The C_{max} concentration was 2317.69 ± 180.44 ng/mL and 3446.14 ± 190.12 ng/mL, respectively for each dosed batch.[21] Garcinol has been demonstrated to be absorbed relatively quickly as T_{max} at 1-2 hours post-dose.[21] However, the observed increase in T_{max} with dose level suggests that absorption may also saturate.[21]

6.2.2 Distribution and Elimination

The calculated volume of distribution values were quite high ($57,891.66 \pm 4,334.00$ mL/kg and $39,184.87 \pm 3,603.87$ mL/kg), which is likely a reflection of garcinol's moderate lipophilicity ($\log P = 6.85$), and the extensive distribution throughout tissues. The terminal elimination half-life differed widely based on route of administration [Oral Dose - 25.43 ± 1.24 hour and 24.46 ± 3.52 hour] versus [Intravenous Dose - 11.94 ± 5.29 hour] 7. The extended half-life observed after the oral route may be one of the

results of garcinol accumulation as a function of its high lipophilicity.[21]

6.2.3 Hepatic Metabolism

The metabolic stability studies using human liver microsomes showed that garcinol has medium clearance characteristics with an intrinsic hepatic clearance (CL_{int}) of $33.94 \mu\text{L}/\text{min}/\text{mg}$ of microsomal protein. However, it was calculated that 94% of garcinol would circumvent hepatic first-pass metabolism, leading to moderate oral bioavailability [21]

6.3 Delivery Challenges

6.3.1 Solubility and Formulation Issues

Garcinol has a number of physicochemical characteristics that affect drug delivery and bioavailability. First, garcinol is poorly soluble in aqueous solvent and has rapid metabolic clearance, which contributes to its inability to reach its full therapeutic potential. When serum protein was assessed in various in vitro studies, garcinol proved effective, however 10% of fetal serum reduced the half-maximum inhibitory concentration value up to 10 times as a result of serum protein interactions.[16]

6.3.2 Nanotechnology-Based Solutions

Understanding these delivery limitations has prompted efforts to study nanotechnology-based approaches. For instance, researchers have developed numerous nanoparticle-based formulations to increase garcinol's aqueous solubility and increase bioavailability and pharmacologic efficacy. Researchers have developed garcinol-loaded PLGA nanoparticles that alleviated the solubility issues with relatively promising in vitro release properties and stability. These formulations have been characterized in



terms of encapsulation efficiency, mean particle size, and in vitro release properties [16], [21].

Garcinol has an excellent toxicology profile in preclinical studies with little toxicity and low oral bioavailability (26 - 36%) and good properties such as, the liver being minimally involved in first-pass metabolism. When all is said and done, there are some major concerns about delivery, mainly low aqueous solubility issues and protein binding interactions limiting the therapeutic efficacy of garcinol. Nanotechnology-based delivery approaches appear promising for addressing some of these challenges, but clinical translation of the technology to eventual therapeutic translation of garcinol is still necessary.

6.4 Nanoparticle and Formulation Advancements in Preclinical Delivery of Garcinol

6.4.1 Gold Nanoparticle Conjugation Systems

6.4.2 Bioconjugated Gold Nanoparticles (G-AuNPs)

Recent advancements in the delivery of garcinol through bioconjugation to gold nanoparticles via glycation reactions represent one of the most exciting innovations. We achieved extraordinary bioconjugation efficiencies of 78.6% of garcinol across the AuNP surface. The synthesis employed an easy glycation reaction-mediated strategy to improve the sustainability of our nanoconjugates with better therapeutic characteristics.[56]

6.4.3 Enhanced Therapeutic Efficacy

The G-AuNPs exhibited much more favourable outcomes relative to pure garcinol in numerous therapeutic endpoint results. In antiglycation studies, G-AuNPs were better inhibitors of glycation reactions as they were able to block formation of early glycation adducts as well as

advanced glycation end products. The nanoconjugates were able to successfully shield glycation-prone free arginine and lysine residues from taking part in harmful glycation reactions rather than obtain glycation from free reactants.[56]

6.4.4 Improved Anticancer Activity

The anticancer potential of G-AuNPs was significantly enhanced compared to free garcinol.[56] The nanoconjugates disrupted mitochondrial membrane potential ($\Delta\Psi_m$) of A549 lung cancer cells at much lower concentrations (13.3 μM) compared to pure garcinol (28.7 μM).[56] This enhanced potency was attributed to improved cellular uptake, targeted delivery, and sustained release of garcinol from the gold nanoparticle surface.

6.4.5 Antidiabetic Enhancement

G-AuNPs also demonstrated superior α -amylase inhibition activity compared to standard inhibitors.[56] While pure garcinol showed significant α -amylase inhibition ($\text{IC}_{50} = 8.9 \mu\text{M}$) compared to acarbose ($\text{IC}_{50} = 0.118 \text{ mM}$), the nanoconjugated form exhibited even greater potency.[56] This enhanced antidiabetic activity positions G-AuNPs as promising candidates for diabetes management applications.

6.5 Polymer-Based Nanoparticle Systems

6.5.1 PLGA Nanoparticle Formulations

Garcinol-loaded vitamin E TPGS (D- α -tocopheryl polyethylene glycol 1000 succinate) emulsified poly (lactic-co-glycolic acid) (PLGA) nanoparticles represent a sophisticated approach to garcinol delivery. These biodegradable nanoparticles have been extensively characterized for physicochemical properties, including particle



size distribution, zeta potential, encapsulation efficiency, and in vitro release profiles.[16]

The PLGA system has multiple advantages, such as controlled drug release, better stability, and a better biocompatibility profile. The utilization of vitamin E TPGS as emulsifier improves stability but may also confer additional antioxidant and free radical scavenging benefits. PLGA formulations incorporated with garcinol have shown promising results in both in vitro and in vivo characterizing designs showing improved therapeutic effects over the free drug (garcinol).[16]

6.5.2 Characterization and Performance

The PLGA nanoparticles have been thoroughly characterized using advanced analytical techniques including differential scanning calorimetry, Fourier transform infrared spectroscopy, and X-ray diffraction. These analyses confirm successful garcinol encapsulation and provide insights into drug-polymer interactions that influence release kinetics and stability.[16]

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spectroscopy, and x-ray diffraction. These analyses confirm the successful encapsulation of garcinol and suggest associations with drug-polymer interactions that enhance release and stability profiles during characterization and testing on the formulations.[16]

6.6 Protein-Based Nanocarrier Systems

6.6.1 Papain Nanoparticles as Drug Carriers

Although primarily developed for imaging purposes, papain nanoparticles (P-NPs) made via radiation-induced processes represent a new class of protein-based drug delivery systems. These nanoparticles, with an average diameter of 9.3 ± 1.9 nm, are almost spherical and are extremely biocompatible with low cytotoxicity. The radiation-induced crosslinking of proteins forms stable nanoparticles via the free radical-mediated formation of covalent bonds, predominantly bityrosine linkages. Advantages of this green nanotechnology approach over other approaches include eco-friendliness, scalability and stability for functional protein nanoparticles - as they retain protein functionality yet provide space for manipulation to produce stable nanocarriers.[57]

6.6.2 Tumor-Targeting Capabilities

In terms of tumor targeting capability, P-NPs labelled with technetium-99m (^{99m}Tc -P-NPs) provided a radiochemical yield of $94.2 \pm 3.1\%$ and excellent stability ($\geq 90\%$) for 6 hours. These nanoparticles displayed significant tumor uptake by breast cancer models, with the tumor-to-muscle ratio increasing from approximately 5.3 at 2 hours post injection to 7.1 at 6 hours post injection. This increase in tumor targeting capability suggests an opportunity for applications related to drug delivery of garcinol to sites of cancer amid normal tissue. [57]



6.7 Titanium Dioxide Nanocomposites

6.7.1 Antimicrobial Applications

Garcinol has been embedded into titanium dioxide (TiO₂) nanoparticles in order to formulate multifunctional nanocomposites with greater antimicrobial activity.[16] In such systems, garcinol's bioactivities- antimicrobial, antioxidant, anti-inflammatory, are combined with the photocatalytic properties of TiO₂ which can provide different synergistic effects in the control of infectious and inflammatory diseases.

6.8 Advanced Delivery Strategies

6.8.1 Surface Modification and Targeting

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6.8.2 Enhanced Permeability and Retention (EPR) Effect

Since nanoparticles are lower than 200 nm, the garcinol-loaded systems are ideally modelled to escape from the reticuloendothelial system and can have extended circulation in the blood. This mechanism becomes a feature of increased retention and permeation and enhanced retention followed by preferential accumulation at tumor sites through enhanced permeability and retention effect (EPR) effect.[56]

7 Therapeutic Uses of Garcinol Beyond Cancer

7.1 Anti-Inflammatory Activity

Garcinol has significant anti-inflammatory effects by reducing the production of essential pro-inflammatory cytokines such as TNF- α , IL-6, and IL-1 β . Garcinol is also able to reduce the inflammatory mediator's nitric oxide (NO) and prostaglandin E₂ (PGE₂). Garcinol inhibits primary inflammatory signaling pathways (NF- κ B and C/EBP) and preclinical trials have shown orally and topically administered garcinol reduced inflammation in models for skin and ear edema, an indication that garcinol may have practical use in treating inflammatory disease processes.[41]

7.2 Antioxidant Effects

Garcinol is able to act as an antioxidant by scavenging free radicals (e.g., hydroxyl radicals) that contribute to oxidative stress and many condition-associated morbidity. Garcinol's antioxidant properties have been attributed to protection of tissue in experimental models of gastric mucosal injury and neurotoxicity. This implies the possibility of targeted therapy in conditions of oxidative stress. [41]

7.3 Neuroprotective and Cognitive Support

Evidence from preclinical studies shows that garcinol has neuroprotective effects against oxidative stress and inflammation, two critical drivers of neurodegenerative conditions. Garcinol has anticholinesterase activity at levels comparable to therapeutic agents used to treat Alzheimer's disease and has been shown to promote neuronal growth, memory, and cognition. Garcinol also appears to ameliorate neuropathic pain in animal models by reducing neuroinflammation and inhibiting microglial activation, indicating its potential as a candidate for managing both neurodegenerative conditions and neuropathic pain prevalence.[58]

7.4 Cardioprotective Potential



Garcinol may exhibit cardioprotective action through its ability to decrease inflammatory status and oxydative stress, which are major contributors to cardiac injury. There is evidence that garcinol effectively scavenges reactive oxygen species, and garcinol appears to decrease pro-inflammatory cytokines, which may limit myocardial injury. There has been limited performance of garcinol in cardiovascular models, and studies are needed to specifically examine cardiac function in these contexts.[9]

7.5 Anti-Ulcer Activity

In animal models of stress-induced and nonsteroidal anti-inflammatory drug (NSAID)-induced gastric ulcers, garcinol reversed both types of ulcers by, in part, through its antioxidant properties and by stabilizing gastric mucosa. [9]

7.6 Antimicrobial Effects

Garcinol has broad-spectrum antimicrobial activity against pathogenic microorganisms including *Staphylococcus aureus*, methicillin resistant *Staphylococcus aureus* (MRSA), *Pseudomonas aeruginosa*, *Escherichia coli*, and *Candida albicans*. The antimicrobial properties of garcinol are enhanced in nanoparticle formulations, especially in silver or titanium dioxide nanoparticles that improve stability and delivery, and potency provide important evidence that support using garcinol as a therapeutic agent against fungal and bacterial infections, particularly with the global increase in antimicrobial resistance. [59]

7.7 Antiviral and Epigenetic Activity

Inhibitors, resulting from garcinol ability to block histone acetyltransferases (potentially contributing to viral replication), were observed. It is possible that these inhibitors serve as a basis for future drug

therapies against viral infections, including HIV and others. [9]

8. CONCLUSION

Garcinol, a polyisoprenylated benzophenone isolated from *Garcinia indica* and related species, has emerged as a promising multitargeted phytochemical with diverse pharmacological activities. Preclinical studies consistently demonstrate its potent antioxidant, anti-inflammatory, epigenetic modulatory, antimicrobial, and anticancer effects across a broad spectrum of in vitro and in vivo models. Mechanistically, garcinol exerts pleiotropic activity through inhibition of NF- κ B, STAT3, COX-2, and histone acetyltransferases, while simultaneously modulating apoptotic regulators, angiogenic factors, and pro-inflammatory cytokines. These multitarget actions highlight its potential role as a lead natural compound in drug discovery and integrative therapeutic strategies.

Toxicological evaluations in rodents indicate a favorable safety profile, with high tolerated doses and no significant adverse effects across acute, sub-chronic, and reproductive studies. Pharmacokinetic analyses further reveal moderate oral bioavailability with minimal hepatic first-pass metabolism, though poor aqueous solubility and rapid clearance remain key translational challenges. Advances in nanotechnology-based delivery platforms—including PLGA nanoparticles, gold nanoconjugates, and protein-based carriers—offer promising solutions to improve garcinol's stability, bioavailability, and therapeutic efficacy.

Despite these encouraging preclinical findings, significant research gaps persist. The absence of clinical pharmacokinetic, pharmacodynamic, and safety data limits definitive conclusions on its translational potential. Moreover, scalable



manufacturing processes, regulatory considerations for nano pharmaceutical formulations, and optimization of dosage strategies require systematic investigation.

In summary, garcinol represents a versatile and multitargeted phytochemical with considerable promise for oncology and beyond, including anti-inflammatory, antimicrobial, neuroprotective, and metabolic applications. To realize its therapeutic potential, future work must prioritize well-designed clinical trials, advanced formulation development, and rigorous regulatory evaluation. Bridging these gaps will be essential for translating garcinol from a promising laboratory compound into a clinically viable therapeutic agent.

DECLARATIONS: -

Ethics approval

This article does not include any studies involving human participants or animals conducted by the authors. Therefore, ethics approval and consent to participate are not applicable.

Consent for publication

All authors have reviewed and approved the manuscript for submission and have given their consent for its publication.

Competing interests

The authors declare that they have no competing interests.

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Conflict of interest

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Authors' contributions

NIKHIL H R conceived the idea for the review and written original manuscript. Manjunatha PM. conceived the idea for the review and reviewed the manuscript. And all other authors reviewed the manuscript

REFERENCES

1. Wu SB, Long C, Kennelly EJ. Structural diversity and bioactivities of natural benzophenones. *Nat Prod Rep.* 2014;31(9):1158–74.
2. Bailly C, Vergoten G. Anticancer Properties and Mechanism of Action of Oblongifolin C, Guttiferone K and Related Polyprenylated Acylphloroglucinols. *Natural Products and Bioprospecting.* 2021 Dec 29;11(6):629–41.



3. Dzoyem JP, Lannang AM, Fouotsa H, Mbazoa CD, Nkengfack AE, Sewald N, et al. Anti-inflammatory activity of benzophenone and xanthone derivatives isolated from *Garcinia* (Clusiaceae) species. *Phytochemistry Letters*. 2015 Dec;14:153–8.
4. Zhang Q, Sun J, Fu Y, He W, Li Y, Tan H, et al. Guttiferone K Exerts the Anti-inflammatory Effect on *Mycobacterium Tuberculosis*-(H37Ra-) Infected Macrophages by Targeting the TLR/IRAK-1 Mediated Akt and NF- κ B Pathway. *Mediators of Inflammation*. 2020 Oct 10;2020:1–16.
5. Marinov T, Kokanova-Nedialkova Z, Nedialkov PT. Naturally Occurring Simple Oxygenated Benzophenones: Structural Diversity, Distribution, and Biological Properties. *Diversity*. 2023 Sep 22;15(10):1030.
6. Newman DJ, Cragg GM. Natural Products as Sources of New Drugs over the Last 25 Years. *Journal of Natural Products*. 2007 Mar 1;70(3):461–77.
7. Gulick RM, McAuliffe V, Holden-Wiltse J, Crumpacker C, Liebes L, Stein DS, et al. Phase I Studies of Hypericin, the Active Compound in St. John's Wort, as an Antiretroviral Agent in HIV-Infected Adults: AIDS Clinical Trials Group Protocols 150 and 258. *Annals of Internal Medicine*. 1999 Mar 16;130(6):510–4.
8. Acuna U, Jancovski N, Kennelly E. Polyisoprenylated Benzophenones from Clusiaceae: Potential Drugs and Lead Compounds. *Current Topics in Medicinal Chemistry*. 2009 Nov 1;9(16):1560–80.
9. Padhye S, Ahmad A, Oswal N, Sarkar FH. Emerging role of Garcinol, the antioxidant chalcone from *Garcinia indica* Choisy and its synthetic analogs. *Journal of Hematology & Oncology*. 2009 Dec 2;2(1):38.
10. Ahmad DrR. *garcinia indica choisy: a review on the biological activities of the underutilized medicinal plant of ayurveda*. *Era's Journal of Medical Research*. 2023 Dec;10(2):60–4.
11. Desai S, Sharma P, Kashyap P, Choudhary B, Kaur J. Bioactive compounds, bio - functional properties, and food applications of *Garcinia indica*: A review. *Journal of Food Biochemistry*. 2022 Oct 7;46(10).
12. Hartati S, Artanti N, Sari L, Ernawati T. Antioxidant and Anticancer Activity of Garcinol Isolated from *Garcinia maingayi* Hook and Molecular Modeling Studies. *Research Journal of Pharmacy and Technology*. 2024 Aug 26;3546–52.
13. Schobert R, Biersack B. Chemical and Biological Aspects of Garcinol and Isogarcinol: Recent Developments. *Chemistry & Biodiversity*. 2019 Sep 30;16(9).
14. Wang L, Wang M, Guo H, Zhao H. Emerging Role of Garcinol in Targeting Cancer Stem Cells of Non-small Cell Lung Cancer. *Current Pharmacology Reports*. 2019 Feb 21;5(1):14–9.
15. Biersack B. Effects of Garcinol from Kokum (*Garcinia indica*) on the Prevention and Treatment of Cancer. In: *Critical Dietary Factors in Cancer Chemoprevention*. Cham: Springer International Publishing; 2016. p. 253–71.
16. Aggarwal V, Tuli HS, Kaur J, Aggarwal D, Parashar G, Chaturvedi Parashar N, et al. Garcinol Exhibits Anti-Neoplastic Effects by Targeting Diverse Oncogenic Factors in Tumor Cells. *Biomedicines*. 2020 Apr 30;8(5):103.
17. Tomasiak P, Janisiak J, Rogińska D, Perużyńska M, Machaliński B, Tarnowski M. Garcinol and Anacardic Acid, Natural Inhibitors of Histone Acetyltransferases, Inhibit Rhabdomyosarcoma Growth and Proliferation. *Molecules*. 2023 Jul 8;28(14):5292.



18. Banerjee S, Parasramka MA, Paruthy SB. Garcinol: Preclinical Perspective Underpinning Chemo- and Radiosensitization of Cancer. In: *Role of Nutraceuticals in Chemoresistance to Cancer*. Elsevier; 2018. p. 297–324.
19. Abu Bakar MF. A multi-targeted approach of garcinol for obesity intervention: Mechanistic insights and possible clinical applications. *PharmaNutrition*. 2024 Jun;28:100388.
20. Nutritional Benefits, Phytoconstituents, and Pharmacological Properties of Garcinia Fruits: A Review. *Biointerface Research in Applied Chemistry*. 2024 Oct 15;14(5):102.
21. Rao Gajula SN, Talari S, Chilvery S, Godugu C, Sonti R. A unique in vivo pharmacokinetic profile, in vitro metabolic stability and hepatic first-pass metabolism of garcinol, a promising novel anticancer phytoconstituent, by liquid chromatography–mass spectrometry. *RPS Pharmacy and Pharmacology Reports*. 2023 Apr 1;2(2).
22. Ganguly SC, Mahanti B, Ganguly S, Majumdar S. Bovine serum albumin as a nanocarrier for efficient encapsulation of hydrophobic garcinol-A strategy for modifying the in vitro drug release kinetics. *International Journal of Biological Macromolecules*. 2024 Oct;278:134651.
23. Gupta NS. Therapeutic Efficacy of the Plant Bioactive Phytochemicals with Special Reference to Alkaloids, Terpenoids, Phenolics and Cardiac Glycosides. *International journal of plant and environment*. 2024 Mar 30;10(01):22–30.
24. Zeeshan Bhatti M, Ismail H, Khan Kayani W. Plant Secondary Metabolites: Therapeutic Potential and Pharmacological Properties. In: *Secondary Metabolites - Trends and Reviews*. IntechOpen; 2022.
25. Chihomvu P, Ganesan A, Gibbons S, Woollard K, Hayes MA. Phytochemicals in Drug Discovery—A Confluence of Tradition and Innovation. *International Journal of Molecular Sciences*. 2024 Aug 13;25(16):8792.
26. Satpathy R. Application of Bioinformatics Techniques to Screen and Characterize the Plant-Based Anti-Cancer Compounds. In 2022. p. 466–84.
27. Olugbemi OT, Charles AO. Roles of metagenomics and metabolomics in computational drug discovery. In: *Phytochemistry, Computational Tools and Databases in Drug Discovery*. Elsevier; 2023. p. 181–93.
28. Salem MA, Perez de Souza L, Serag A, Fernie AR, Farag MA, Ezzat SM, et al. Metabolomics in the Context of Plant Natural Products Research: From Sample Preparation to Metabolite Analysis. *Metabolites*. 2020 Jan 15;10(1):37.
29. Nasim N, Sandeep IS, Mohanty S. Plant-derived natural products for drug discovery: current approaches and prospects. *The Nucleus*. 2022 Dec 18;65(3):399–411.
30. Akram J, Ahmed W, Azmat R. Plants as a natural resource of bioactive compounds for drug formulation to control infectious diseases. *Journal Advances of Nutrition Science and Technology*. 2021 May 13;1(1):11.
31. Najmi A, Javed SA, al Bratty M, Alhazmi HA. Modern Approaches in the Discovery and Development of Plant-Based Natural Products and Their Analogues as Potential Therapeutic Agents. *Molecules*. 2022 Jan 6;27(2):349.
32. Kaloudas D, Penchovsky R. Plant-Derived Compounds and Their Potential Role in Drug Development. In: *Research Anthology on Recent Advancements in Ethnopharmacology and Nutraceuticals*. IGI Global; 2022. p. 502–17.
33. Hsia CW, Huang WC, Jayakumar T, Hsia CH, Hou SM, Chang CC, et al. Garcinol acts as a novel integrin α IIb β 3 inhibitor in human



- platelets. *Life Sciences*. 2023 Aug;326:121791.
34. Ye X, Yuan L, Zhang L, Zhao J, Zhang CM, Deng HY. Garcinol, an Acetyltransferase Inhibitor, Suppresses Proliferation of Breast Cancer Cell Line MCF-7 Promoted by 17 β -Estradiol. *Asian Pacific Journal of Cancer Prevention*. 2014 Jun 30;15(12):5001–7.
35. Huang MT, Liu Y, Badmaev V, Ho CT. Antiinflammatory and Anticancer Activities of Garcinol. In 2008. p. 293–303.
36. Gaonkar RH, Ganguly S, Dewanjee S, Sinha S, Gupta A, Ganguly S, et al. Garcinol loaded vitamin E TPGS emulsified PLGA nanoparticles: preparation, physicochemical characterization, in vitro and in vivo studies. *Scientific Reports*. 2017 Apr 3;7(1):530.
37. Kopytko P, Piotrowska K, Janisiak J, Tarnowski M. Garcinol—A Natural Histone Acetyltransferase Inhibitor and New Anti-Cancer Epigenetic Drug. *International Journal of Molecular Sciences*. 2021 Mar 11;22(6):2828.
38. Majeed M, Bani S, Bhat B, Pandey A, Mundkur L, Neupane P. Safety profile of 40% Garcinol from *Garcinia indica* in experimental rodents. *Toxicology Reports*. 2018;5:750–8.
39. Hulangamuwa AC, Ediriweera MK, Rajagopalan U, Karunaratne DN, Tennekoon KH, Samarakoon SR. Development of a New Nanocarrier for Dietary Garcinol: Characterization and In Vitro Efficacy Evaluation Using Breast Cancer Stem Cells Grown in Hypoxia. *Journal of Food Quality*. 2021 Feb 10;2021:1–10.
40. Paul B, Gaonkar RH, Dutta D, Dasi R, Mukherjee B, Ganguly S, et al. Inhibitory potential of iRGD peptide-conjugated garcinol-loaded biodegradable nanoparticles in rat colorectal carcinoma. *Biomaterials Advances*. 2022 Mar;134:112714.
41. Liu C, Ho PCL, Wong FC, Sethi G, Wang LZ, Goh BC. Garcinol: Current status of its anti-oxidative, anti-inflammatory and anti-cancer effects. *Cancer Letters*. 2015 Jun;362(1):8–14.
42. Saadat N, Gupta S v. Potential Role of Garcinol as an Anticancer Agent. *Journal of Oncology*. 2012;2012:1–8.
43. Ahmad A, Sarkar SH, Bitar B, Ali S, Aboukameel A, Sethi S, et al. Garcinol Regulates EMT and Wnt Signaling Pathways In Vitro and In Vivo , Leading to Anticancer Activity against Breast Cancer Cells. *Molecular Cancer Therapeutics*. 2012 Oct 1;11(10):2193–201.
44. Chen CS, Lee CH, Hsieh CD, Ho CT, Pan MH, Huang CS, et al. Nicotine-induced human breast cancer cell proliferation attenuated by garcinol through down-regulation of the nicotinic receptor and cyclin D3 proteins. *Breast Cancer Research and Treatment*. 2011 Jan 13;125(1):73–87.
45. Tu SH, Chiou YS, Kalyanam N, Ho CT, Chen LC, Pan MH. Garcinol sensitizes breast cancer cells to Taxol through the suppression of caspase-3/iPLA 2 and NF- κ B/Twist1 signaling pathways in a mouse 4T1 breast tumor model. *Food & Function*. 2017;8(3):1067–79.
46. DONG Haitao 1 CJ 2, HC 2, SY 3, ZX 3, CX 2. Role of 8-allyl Garcinol in the Chemoprevention of Oral Squamous Cell Carcinoma.
47. Ranjbarnejad T, Saidijam M, tafakh M sadat, Pourjafar M, Talebzadeh F, Najafi R. Garcinol exhibits anti-proliferative activities by targeting microsomal prostaglandin E synthase-1 in human colon cancer cells. *Human & Experimental Toxicology*. 2017 Jul 1;36(7):692–700.
48. Parasramka MA, Gupta SV. Synergistic Effect of Garcinol and Curcumin on Antiproliferative and Apoptotic Activity in Pancreatic Cancer Cells. *Journal of Oncology*. 2012;2012:1–8.



49. Aamir Ahmad I, Zwcwradkmymssbbbspfhs. Garcinol-induced apoptosis in prostate and pancreatic cancer cells is mediated by NF- κ B signaling. 2011;
50. Rizvi SMD, Almazni IA, Moawadh MS, Alharbi ZM, Helmi N, Alqahtani LS, et al. Targeting NF- κ B signaling cascades of glioblastoma by a natural benzophenone, garcinol, via in vitro and molecular docking approaches. *Frontiers in Chemistry*. 2024 Feb 16;12.
51. Ahmad A, Sarkar SH, Aboukameel A, Ali S, Biersack B, Seibt S, et al. Anticancer action of garcinol in vitro and in vivo is in part mediated through inhibition of STAT-3 signaling. *Carcinogenesis*. 2012 Dec 1;33(12):2450–6.
52. Huang WC, Kuo KT, Adebayo BO, Wang CH, Chen YJ, Jin K, et al. Garcinol inhibits cancer stem cell-like phenotype via suppression of the Wnt/ β -catenin/STAT3 axis signalling pathway in human non-small cell lung carcinomas. *The Journal of Nutritional Biochemistry*. 2018 Apr;54:140–50.
53. Yoshida K, Tanaka T, Hirose Y, Yamaguchi F, Kohno H, Toida M, et al. Dietary garcinol inhibits 4-nitroquinoline 1-oxide-induced tongue carcinogenesis in rats. *Cancer Letters*. 2005 Apr;221(1):29–39.
54. Wang Y, Tsai ML, Chiou LY, Ho CT, Pan MH. Antitumor Activity of Garcinol in Human Prostate Cancer Cells and Xenograft Mice. *Journal of Agricultural and Food Chemistry*. 2015 Oct 21;63(41):9047–52.
55. Patwa N, Chauhan R, Chauhan A, Kumar M, Ramniwas S, Mathkor DM, et al. Garcinol in gastrointestinal cancer prevention: recent advances and future prospects. *Journal of Cancer Research and Clinical Oncology*. 2024 Jul 27;150(7):370.
56. Rafi Z, Hassan Baig M, Mabood Husain F, Yousef Alomar S, Dong JJ, Sajid Khan M. Biological reaction mediated engineered AuNPs facilitated delivery enhance the anticancer, antiglycation, and antidiabetic potential of garcinol. *Journal of King Saud University - Science*. 2023 Apr;35(3):102524.
57. Ferreira A, Marques F, Real C, Thipe V, Freitas L, Lima C, et al. Green Nanotechnology Through Papain Nanoparticles: Preclinical in vitro and in vivo Evaluation of Imaging Triple-Negative Breast Tumors. *Nanotechnology, Science and Applications*. 2024 Sep;Volume 17:211–26.
58. Mishra A, Sankar Satapathy B, Gatadi S, Vasavi M, Kumar Sahu P, Nuthakki V. Garcinol as A Promising Therapeutic Candidate for Alzheimer's Disease: Integrated Computational and In Vitro Target Analysis. *ChemistrySelect*. 2025 Jul 4;10(25).
59. Fernando HN, Kumarasinghe UR, Gunasekara CP, Wijekoon SK, Ekanayaka AK, Rajapaksha SP, et al. Synthesis, Characterization and Antimicrobial Activity of Garcinol Capped Silver Nanoparticles. *Journal of Microbiology and Biotechnology*. 2019 Nov 28;29(11):1841–51

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