

# INTERNATIONAL JOURNAL OF PHARMACEUTICAL SCIENCES

[ISSN: 0975-4725; CODEN(USA): IJPS00] Journal Homepage: https://www.ijpsjournal.com



#### **Review Article**

## In-Vitro Evaluation of Diclofenac Sodium Gel

# Nand Lal Thakur\*, Sana Nusrat Praween

A and E College of Pharmacy, Samastipur, Bihar

#### ARTICLE INFO

Published: 30 Sept 2025

Keywords:

Diclofenac sodium, Antiinflammatory,

Hydroxypropyl

methylcellulose, Carbopol, Drug release, Glycerin

DOI:

10.5281/zenodo.17235413

#### **ABSTRACT**

To develop diclofenac sodium gel using high molecular weight hydroxypropyl methylcellulose (HPMC) and Carbopol 934P for topical and systemic delivery. Diclofenac sodium gel was prepared with HPMC K100M and Carbopol 934P as gelling agents. The formulations were examined for pH, spreadability, consistency, viscosity, homogeneity, drug content and stability. In vitro drug release was evaluated using Franz diffusion cell. Carrageenan induced rat paw oedema model was used for the evaluation of the anti-inflammatory activity of the gels. A commercial diclofenac sodium gel product was used as the reference drug. Formulations containing glycerin as permeation enhancer gave drug release patterns comparable to that of the reference product. The drug content of F2, F5 and F9 was 99.81, 99.75 and 99.96 %, respectively. Accelerated stability results showed no significant variation in the appearance and drug release after storage for 3 months. Diclofenac sodium gel containing HPMC K100M and Carbopol 934P exhibited pronounced anti-inflammatory activity and could be further developed for topical and systemic delivery.

#### INTRODUCTION

Drug delivery through the skin has been a promising concept for a long time because the skin is easy to access, has a large surface area with vast exposure to the circulatory and lymphatic networks and the route is non-invasive[1]. Transdermal delivery is of great importance for drugs that may cause systemic side effects, e.g., non-steroidal anti-inflammatory drugs (NSAIDs)

Diclofenac sodium is an effective NSAID often used in the treatment of acute and chronic arthritic conditions. It is prescribed for long-term treatment of rheumatoid arthritis, osteoarthritis and ankylosing spondylitis [3].

There is great interest to develop non-oral dosage forms of diclofenac sodium to minimize its gastric side effects and to provide relatively consistent drug levels at the application site for prolonged periods [4].

\*Corresponding Author: Nand Lal Thakur

Address: A and E College of Pharmacy, Samastipur, Bihar

**Email** : nandlalt47@gmail.com

**Relevant conflicts of interest/financial disclosures**: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.



Topical delivery of diclofenac sodium using various formulations has been described in the literature. However, effective permeation of the drug through skin is difficult to achieve due to its intrinsically poor permeability, though this is relatively good compared to other commonly used NSAIDs [5]. Skin permeation enhancers can improve drug skin penetration [6].

HPMC K100M and Carbopol 934P have been used as hydrophilic polymers in the formulation of gels for transdermal drug delivery. One of their most important characteristics is high swellability, which has a significant effect on the release kinetics of the incorporated drug [7]. Cellulose ethers, especially HPMC K100M, are frequently used as the basis for sustained release hydrophilic matrix tablets [8].

The overall drug release mechanism of HPMC K100M-based pharmaceutical devices strongly depend on the design (composition and geometry) of the particular delivery system. On imbibing water, HPMC K100M swells, resulting in dramatic changes in the viscous nature of the polymer, drug concentration and increased dimension of the dosage system. Upon contact with water, the incorporated drug dissolves and diffuses out of the device. Depending on the chain length and degree of substitution of the HPMC type used, the polymer itself dissolves more or less rapidly.

Diffusion, water uptake and erosion are the most important rate-controlling mechanisms of commercially available controlled release products [9].

A series of grades based on the molecular fraction of these polymers are used at concentrations between 1 and 5 % in topical gel formulation. Carrageenan-induced paw oedema in mice has been widely used for the evaluation of antiinflammatory activity of drugs [10].

The aim of this study was to develop suitable topical formulations of diclofenac sodium using HPMC K100M and Carbopol 934P as gelling agents and glycerin as permeation enhancers.

#### 1. In-vitro Diffusion Studies:

An "in vitro diffusion study" is a laboratory experiment where the movement of molecules (diffusion) through a membrane, like a biological tissue or a synthetic barrier, is observed outside of a living organism, allowing researchers to analyze how a substance might pass through a specific barrier under controlled conditions, typically using a diffusion cell apparatus to measure the rate of diffusion across the membrane.

# 2.1 Key points about in vitro diffusion studies:

### 2.1.1 "In vitro" means "in glass":

This signifies that the experiment is conducted outside a living organism, usually in a controlled laboratory setting.

## 2.1.2 Diffusion process:

Molecules naturally move from an area of high concentration to an area of low concentration, which is the principle behind diffusion.

#### 2.1.3 Diffusion cell:

A specialized device used to study diffusion, typically consisting of a donor compartment where the substance is placed, a membrane separating the compartments, and a receptor compartment where the diffused substance is collected.

#### 2.2 Applications of in vitro diffusion studies:

• **Drug delivery research:** Assessing how well a drug can penetrate through the skin (transdermal delivery) or other biological membranes to reach its target site.



- Cosmetics development: Evaluating the penetration of cosmetic ingredients into the skin.
- **Food science:** Studying the release of flavors or additives from food matrices.
- **Membrane research:** Characterizing the permeability properties of different membranes.

## Important factors in an in vitro diffusion study:

# Membrane type:

The choice of membrane (e.g., animal skin, synthetic membrane) significantly impacts the results.

# • Donor solution composition:

Concentration of the substance being tested and the vehicle used to dissolve it.

### • Receptor fluid:

The medium used to collect the diffused substance, often a buffer solution.

# • Temperature control:

Maintaining a consistent temperature throughout the experiment.

#### 2. Diclofenac Gel

If you've dealt with osteoarthritis pain in the past, you may be familiar with diclofenac. Diclofenac is a non-steroidal anti-inflammatory drug (NSAID) and it falls in the same class as drugs such as ibuprofen, naproxen and aspirin. Diclofenac eases pain and reduces inflammation like other NSAID medications. It works by blocking the production of certain enzymes in our body called COX1 and COX2. These enzymes are responsible for the production of chemicals called prostaglandins which cause pain and inflammation. So, when you block the production of COX 1 and 2, you reduce the production of prostaglandins, resulting in less pain and inflammation.

# 3.1 What is over-the-counter diclofenac gel used for?

Diclofenac sodium is the active ingredient in Voltaren Arthritis Pain, an over-the-counter arthritis pain relief gel. OTC diclofenac gel is used to relieve pain from arthritis in the knees, ankles, feet, elbows, wrists, and hands.<sup>2</sup>

Over-the-counter diclofenac sodium topical gel can be good option for people who don't want to take oral NSAIDs or they can't swallow pills.

# 3.2 How should I use diclofenac gel?

Over-the-counter topical diclofenac comes as a 1% gel to apply on up to two joint areas, four times daily.<sup>2</sup> These areas can include the foot, ankle, knee, hand, wrist or elbow. It's recommended to apply your OTC diclofenac gel around the same time every day. Some basic tips for application for Voltaren Arthritis Pain diclofenac gel:

- Apply diclofenac gel on clean, dry skin.
- Apply the gel to two joints only. For example, a hand and an elbow, a knee and an ankle or two knees. Use your hands to gently rub the gel into your skin.
- For each upper body area (hand, wrist, or elbow), squeeze out 2.25 inches (2 grams) and apply. For each lower body area (foot, ankle, or knee), squeeze out4.5 inches (4 grams) and apply.
- Wash your hands thoroughly after applying the gel unless you are using the medication for your hands. If so, do not wash your hands for an hour after application.
- Do not apply gel to skin that is broken, infected, cut or covered in a rash
- Diclofenac topical gel is only for external use on the skin. Be careful not to get diclofenac gel in your eyes, nose or mouth.



Diclofenac sodium gel may cause a severe allergic reaction, especially in people allergic to aspirin. Symptoms include:<sup>3</sup>

- Hives
- Asthma (wheezing)
- Skin reddening
- Blisters
- Facial swelling
- Shock
- Rash

Make sure to keep your diclofenac sodium topical gel in the container it came in and out of reach of children or pets. It's important to keep it out of sight and high enough that children can't reach it or see it.

## 3.3 Who should use diclofenac gel?

Before using topical diclofenac gel, you should read the product label to determine if it is right for you. People with certain underlying medical conditions are advised to talk to their doctor before use so it is important to understand if this includes you. It may not be right for you if you already take oral NSAIDs, you have sensitive skin, or you have several affected joints.<sup>4</sup>

There are also several drugs' interactions you should consider if you're considering using OTC diclofenac gel. Ask your doctor or pharmacist before using diclofenac gel if you are taking any other pain reliever or are planning on taking any other pain reliever, especially prescription or nonprescription NSAIDs (aspirin, ibuprofen, naproxen, or others).

Now that you know more about diclofenac gel, the active ingredient in Voltaren, make sure to visit our arthritis resource center to learn more about managing your arthritis symptoms.

Content contained in this article is not meant to be used as a direct indication of Voltaren product use or results.



Fig.1: Voltaren Gel (Diclofenac Sodium Topical Gel)



Fig.2: Diclofenac Sodium Topical Gel



Fig.3: Diclofenac Sodium Topical Gel

# 4. All the key aspects of in vitro evaluation of Diclofenac Sodium Gel:

# **4.1. Formulation Preparation** [11]

- Composition: Diclofenac Sodium Gel is typically formulated with the active ingredient diclofenac sodium, a gel base (such as carbopol or hydroxypropyl methylcellulose), preservatives, and other excipients.
- **Gel Preparation:** The gel is mixed in a way that ensures a consistent texture, uniform



distribution of the active ingredient, and appropriate viscosity for easy application.

## 4.2. Drug Release Studies

- Release Testing: The in vitro drug release is measured using apparatus like the Franz diffusion cell or a modified USP dissolution apparatus. The gel is applied on a membrane, and the drug release is monitored over time by sampling the receptor solution.
- Cumulative Release: The amount of diclofenac sodium released from the gel at specific time intervals is measured using UV spectrophotometry or HPLC (High-Performance Liquid Chromatography).
- Release Kinetics: Data is analyzed to determine the release mechanism, typically using models such as zero-order, first-order, or Higuchi to predict how the drug releases over time.

#### 4.3. Permeation Studies

- **Skin Permeation**: This study assesses how well diclofenac sodium passes through the skin. A rat or human skin membrane is typically used as a model. The gel is applied, and the amount of drug permeated into the skin is measured.
- Cumulative Permeation: Samples of the permeated drug are collected at regular intervals and analyzed to determine the drug's penetration rate. The rate of permeation is important for ensuring therapeutic effectiveness.

#### 4.4. Viscosity and Texture Analysis

• **Rheological Properties**: Viscosity tests are conducted to determine the gel's consistency. The gel must have the right viscosity to ensure

- it is easy to apply, stays on the skin, and releases the drug efficiently.
- **Spreadability**: The spreadability of the gel is tested to ensure it can be evenly spread across the skin, a factor that affects both user convenience and the drug's effectiveness.

## 4.5. Stability Testing

- Storage Conditions: Stability studies are conducted by storing the gel under different conditions (e.g., at various temperatures and humidity levels) and assessing the physical and chemical stability of the formulation over time.
- **pH and Appearance**: The pH of the gel and its appearance are monitored during stability studies to ensure the gel does not undergo significant changes that could affect its performance.

## 4.6. Release Mechanism and Model Fitting

- Mathematical Modeling: Data from the release and permeation studies are fitted into different mathematical models (e.g., Higuchi, Peppas model, zero-order, and first-order kinetics) to understand the mechanism of drug release (e.g., diffusion, swelling, or erosion of the gel matrix).
- **Diffusion Coefficient (D)**: This is calculated to determine how effectively the drug diffuses across the skin.

#### 4.7. Drug Content and Uniformity

• Content Uniformity: The gel is tested to ensure that the active ingredient is uniformly distributed. This is done by measuring the diclofenac sodium content in different portions of the gel to ensure consistency.



 Assay: HPLC or UV spectrophotometry can be used to measure the exact concentration of diclofenac sodium in the gel.

#### 4.8. Microbial Testing

 Microbial Contamination: Tests are performed to ensure that the gel is free from harmful microbial contamination, especially since it is for topical application.

## 4.9. Irritation Testing

• **Skin Irritation**: Even though this is usually performed in vivo, in vitro skin irritation tests can be performed to ensure that the gel does not cause adverse reactions or irritation when applied to the skin [11].

#### 5. EXPERIMENTAL

#### 5.1. MATERIALS AND METHODS

### **5.1.1.** Materials<sub>[12]</sub>

Diclofenac sodium, was procured from Research fine lab Hyderabad. Mumbai. Methyl paraben, propyl paraben, isopropyl alcohol, menthol were obtained from Samar chemicals, Nagpur. Triethanolamine from Merck chemical Mumbai, Propylene glycol Carbopol 940P from Research Fine Lab, Mumbai. Hpmc k4m from Merck Chemical Mumbai and marketed gel from Intas Pharmaceuticles Ltd, Ahmedabad

#### **5.1.2.** Methods:

Formulation of Topical gel containing Diclofenac Sodium was prepared by direct compression technique using varying concentrations of different grades of polymers with carbopol 940 and Hpmc k4m. All the ingredients were accurately weighed and passed. Then, most of the ingredients were buy from summer chemicles, Nagpur.

## **5.1.3.** Extraction of Ginger oleoresin: [13]

Ginger oleoresin is extracted from dried crushed ginger in soxhlet extractor using ethyl ether as a solvent and linseed oil are two key ingredients added to propose formulation for antiinflamatiory effect.

# 5.1.4. Two different procedures are applied to formulate gel

- 1) formulation of hpmc gel Diclofenac was dissolved in propylene glycol. Menthol was dissolved in Isopropyl alcohol. The whole amount of HPMC was sprinkled on drug solution with slow stirring then methyl paraben and propyl paraben were added. The mixture of drug solution and polymer was kept aside for six hours to seven hours, for adequate swelling of polymer. The oil phase consisting of linseed oil, ginger oleoresin and methyl salicylate was added slowly in above aqueous gel with continuous stirring with overhead stirrer. The gel was packed in aluminium collapsible tube. Ur
- 2) formulation of carbopol gel Carbopol was added in given quantity of water and kept it for 8 hours for adequate swelling of polymer. Triethanolamine was added for neutralize the above Carbopol base for the proper pH adjustment. Diclofenac was then added in propylene glycol at 65 0C. After that methyl and propyl paraben was added in the solution of propylene glycol, cooled to room temperature and the resultant was added in carbopol base with the help of mechanical stirrer by rotating at 600 rpm.

#### 5.1.5. Solubility analysis

The solubility of diclofenac sodium was determined using various solvents of different



polarities including methanol, 95 % ethanol, water, glacial acetic acid, ether, chloroform and phosphate buffer (pH 6.8). The vials containing the solvent and excess diclofenac sodium were kept in a shaker for 24 h. It was filtered through Whatman filter paper. The filtrate was analyzed by UV spectrophotometry to determine the amount of diclofenac sodium.

# **5.1.6.** Fourier transform-infrared spectroscopy (FTIR) study

FT-IR study was used to check compatibility and interaction between the drug and the polymers. The spectra of pure diclofenac sodium and the physical mixture of diclofenac sodium and polymer, embedded in KBr discs, were recorded in the range of 4000 cm-1 and 400 cm-1 using IR spectroscopy (Shimadzu, IR Affinity-1).

## **5.1.7.** Preparation of diclofenac sodium gel [14]

For formulations F1, F2 and F3, 1 g of diclofenac sodium was dissolved in 15 ml of glycerin with the aid of mild heat (solution A). Weighed quantity of HPMC K100M was added to 75 ml of distilled water and stirred until dissolved (solution B). Solutions A and B were mixed thoroughly and the final weight made up to 100 g. For formulations F4, F5 and F6, 1 g of diclofenac sodium was dissolved in 15 ml of glycerin with the aid of mild heat (solution A). Weighed quantity of Carbapol 934P was added to 75 ml of distilled water, stirred

until dissolved and then neutralized with 10 % NaOH (solution B). Solutions A and B were mixed thoroughly and the final weight made up to 100 g. For formulations F7, F8 and F9, 1 g of diclofenac sodium was dissolved in 15 ml of glycerin with the aid of mild heat, and methyl paraben and propyl paraben added (solution A). Weighed quantity of sodium alginate was added to 75 ml of distilled water and stirred until dissolved (solution B). Solutions A and B were mixed thoroughly and the final weight made up to 100 g. The composition of the formulations is outlined in Table 1.

The three different hydrophilic polymers were used for study and in each formulation code the combination of two polymers were used by keeping one of the polymers constant. The different nine formulation of diclofenac sodium gel (F1-F9) with their code are listed in table 1. The gel were kept in plastic well closed container and stored at room temperature until the time of analysis.

## 5.1.8. In vitro evaluation of diclofenac gels [15]

The diclofenac sodium gels were subjected to evaluation for the following parameters - pH, spreadability, consistency, viscosity, homogeneisity [14], drug content, in vitro drug release studies, in vivo anti-inflammatory activity by carrageenan induced paw oedema and accelerated stability studies.

Table 1: Composition of diclofenac sodium gel

Formulation	Drug	HPMC	Carbopol	10%	Glycerin	Propyl	Distilled
Code	(g)	K100M (g)	934P (g)	NaOH (ml)	(ml)	Paraben (g)	water to (g)
$F_1$	1	2.0	0.25	q.s.	15	-	100
$F_2$	1	2.0	0.50	q.s.	15	-	100
$F_3$	1	2.0	0.75	q.s.	15	1	100
F <sub>4</sub>	1	1	0.50	q.s.	15	0.1	100
$F_5$	1	1	0.50	q.s.	15	0.1	100
$F_6$	1	1	0.50	q.s.	15	0.1	100
$F_7$	1	1	-	q.s.	15	0.1	100
F <sub>8</sub>	1	1.5	_	q.s.	15	0.1	100
F <sub>9</sub>	1	2.0	-	q.s.	15	0.1	100



# 5.1.9. Determination of pH of gel formulations

The pH of the gel formulations was measured with a pH meter (Eutech, Cyberscan) using 1 % aqueous solutions of the gels at room temperature.

This parameter was determined with a wooden block and glass slide apparatus. The gel (approx. 20 g) was added to the pan and the time for the upper slide (movable) to separate completely from the fixed was noted. Spreadability was calculated formulas in Eq 1.

$$S = W \times L/T \dots (1)$$

where S = spreadability, W = weight tide to upper slide, L = length of glass slide, and T = time taken to separate the slide completely from each other

#### **5.1.10.** Consistency [16]

Measurement of consistency of the gels was carried out by dropping a cone attached to a holding rod from a fix distance of 10 cm in such way that it falls in the centre of a glass cup filled with the gel. The penetration by the cone was measured from the surface of the gel to the tip of the cone inside the gel. The distance traveled by the cone after 10 s was noted.

#### **5.1.11.** Viscosity

The viscosity of the formulations was determined using a Brookfield digital viscometer (model DVII, USA) equipped with spindle S27.. The gel sample (5 g) was placed in the sample holder of the viscometer and allowed to settle for 5 min and the viscosity measured a rotating speed of 50 rpm at room temperature (25 - 27 oC).

#### 5.1.12. Spreadability

Spreadability was performed with extensiometer apparatus. The apparatus consists of two square

glass plates, 11 cm on each side. On the outside part of the inferior plate a coordinate paper is attached, on which five concentric circles with perpendicular diameter in millimeters are drawn. The spreadability was determined as follows: 1 g gel was placed between the plates and the upper plate was increasingly loaded with weights at equal time intervals. Based on the results of 3 measurements the mean calculated surfaces were plotted in the form of extensiometric curves. On Ycoordinate the ointment surfaces, in cm2 were marked and on Xcoordinate the loadings value, in grams (g) were marked. [17]

## 5.1.13. pH

The pH was measured in water solutions of each gel, using a digital pH meter, which was calibrated before each use with standard buffer solutions at pH 4.6 and 8.6. The solutions are prepared by dissolving 2.5 g of each gel in 25 g water [18]

# 5.1.14. Homogeneity

All the gels formed were tested for homogeneity by visual inspection after the gels have been allowed to set in a container. They were tested for their appearance and presence of any aggregates.

# 5.1.15. Drug content

A quantity (100 mg) of the gel was dissolved in 100 ml of phosphate buffer of pH 6.8. The volumetric flask containing gel solution was shaken for 2 h on a mechanical shaker to allow the drug to dissolve completely. The solution was filtered and drug content determined spectrophotometrically at 276 nm using phosphate buffer (pH 6.8) as blank.

# 5.1.16. In-vitro evaluation of diclofenac sodium release



Pretreated skin of albino mice was used in the Franz diffusion cell experiment. The receptor compartment contained 100 ml of phosphate buffer pH 6.8. One gram of the test formulation or reference was applied to the skin over an area of 1.131 cm2 and placed across the donor compartment. The donor cell was exposed to ambient temperature and covered with parafilm to prevent evaporation. The temperature of the diffusion medium was maintained at  $37 \pm 1$  oC while the buffer solution was stirred continuously with a Teflon-coated magnetic bar at 500 rpm. Samples (1 ml each) were withdrawn from the release medium at 30, 60, 90 and 120 min and replaced with an equal volume of fresh buffer solution to maintain sink conditions. The samples were analyzed spectrophotometrically at 276 nm against their respective blank. [20]

#### 5.1.17. Accelerated stability studies

All the selected formulations were subjected to accelerated stability test over a period of three months as per ICH guidelines at a temperature of  $40 \pm 2$  oC/ and 75 % relative humidity (RH). All the formulations were analyzed for changes in appearance, pH and drug content as described above.

#### 5.1.18. Statistical analysis

The results were expressed as mean  $\pm$  standard deviation. The data were analyzed by one way

analysis of variance (ANOVA) followed by Bonferroni's multiple comparison. A level of significance of p < 0.05 was set to determine any significance. [21]

#### 6. RESULTS

## 6.1. Characterization of Formulations

The prepared formulations shared a smooth and homogeneous appearance. The HEC diclofenac sodium gels were transparent while Vurdon gel was white viscous, opalescent. All preparations spreadable, with acceptable were easily bioadhesion and fair mechanical properties. At table 2 are shown the values of pH, viscosity and drug content for each gel. The pH values ranged from  $7.33 \pm 0.016$  to  $8.35 \pm 0.136$ , which are considered acceptable to avoid the risk of irritation after skin application. Viscosity is an important physical property of topical formulations, which affects the rate of drug release; in general, an increase of the viscosity vehicles would cause a more rigid structure with a consequent decrease of the rate of drug release.

#### 6.2. Spreadability

Mean results of three measurements expressed in the form of spreadability curve are shown at figure 4.

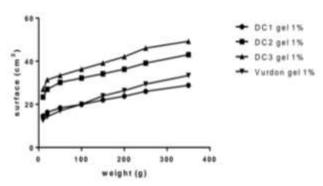


Fig. 4: It shows curves of the four gels In vitro release



The three DC formulations were found to express good spreadabilty compared with that of commercial gel. Considering the stability studies and physiochemical parameters, batch DC1 and DC3 were selected for in vitro permeability release studies as well as compared with the marketed gel. The results are shown at the figures 5 and 6.

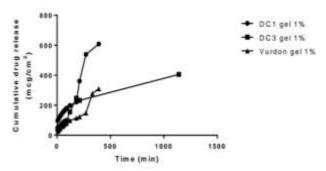


Fig. 5. It shows release of DS from the two formulations and Vurdon Gel

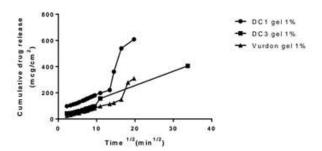


Fig. 6. It shows release rate of DS from the two formulations and Vurdon Gel

When the amounts of drug released per unit area ( $\mu$ g/cm2) were plotted against the square root of time, a linear relationship was obtained for each gel, showing that the release of drug from the gels could be well described by the Higuchi model, where the ratecontrolling step is the process of diffusion through the gel matrix. It is possible to calculate the steady state flux (J) from the slope of the linear portion (5-300 min) of the graph of the release rate of drug.

## 6.3. Drug-excipient compatibility

There were no significant changes in the peak pattern of the IR spectra of pure diclofenac sodium and of the combined drug/polymer mixture (fig 4), which implies that there was no interaction between the drug and the polymers. [22]

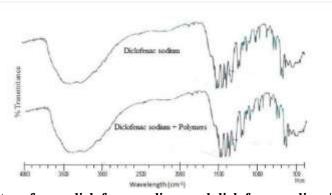


Fig 7: FT-IR spectra of pure diclofenac sodium and diclofenac sodium/polymer mixtures



# **6.4.** Physical properties of gel [23]

Precipitation occurred in some of the gel batches (F1, F3, F4, F6, F7 and F9) of polymer-based gel containing diclofenac sodium which was probably due to incompatibility. Hence, these batches were discarded and remaining batches (F2, F5 and F8) were used in further studies.

The results for pH, spreadability, viscosity, consistency, homogeneity and drug content are shown in Table 2. Spreadability data indicate that

the gel is easily spreadable by a small amount of shear. Consistency reflects the capacity of the gel to get ejected in uniform and desired quantity when the tube is squeezed. Consistency in terms of distance travelled by the cone was 6 mm. The gel formulations were homogeneous in texture and fell within a pH range of 6.8 to 7.4 which is within the normal skin pH in healthy people. The results show that there was no significant difference between the viscosities of the gel formulations and that of the reference.

Table 2: Properties of some diclofenac sodium gel formulations

Batch	pН	Spreadability (g.cm/s)	Viscosity (dyn·s/cm )	Consistency (mm)	Homogeneity	Drug Content (%)
F <sub>2</sub>	7.4	5.6	0.94×10 <sup>-3</sup>	6	Very good	99.81
F <sub>5</sub>	6.8	3.8	1.60×10 <sup>-3</sup>	6	Good	99.75
F <sub>9</sub>	7.1	3.9	1.70×10 <sup>-3</sup>	6	Good	99.96

#### 6.5. In vitro diclofenac sodium release

Diclofenac sodium release from the F2 formulation was comparable with that of the reference standard (commercial formulation), as Table 3 shows. The pH of F2 and the commercial gel was was 6.8.

Table 3: In vitro release characteristics of diclofenac sodium gel

Time (min)	Drug rele	ase (%)
18400000 N	F <sub>2</sub>	Ref*
30	38.54	39.64
60	64.78	65.86
90	81.23	82.92
120	96.87	97.34

\*Reference standard (commercial formulation

# 6.6. In vivo anti-inflammatory activity of diclofenac gel

The results of the anti-inflammatory test (Fig 2) indicate there was no significant difference

between the anti-inflammatory activities of the test formulations and the reference [24]

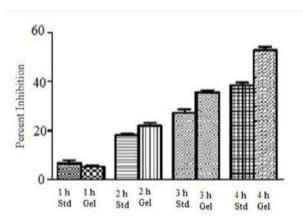


Fig 8: Inhibition of rat paw edema by reference standard (Std) and test (Gel) formulations

#### 6.7. Stability of the gels

The results of the accelerated stability studies are shown in Table 4. The appearance of the gels remained clear and no significant variation in pH was observed after subjecting the formulations to stability stress for 3 months. Drug content was 96.91, 95.33 and 95.65 % formulations F2, F5 and F9, respectively, after the 3-month period.



Table 4: Accelerated stability data for diclofenac sodium gel

Formulation	Month	Appearance	pH	Drug content (%)
F <sub>2</sub>	0	Clear	7.4	99.81
	1	Clear	7.4	98.27
	2	Clear	7.3	97.54
	2 3 0	Clear	7.3	96.91
F <sub>5</sub>	0	Clear	6.8	99.75
	1	Clear	6.8	98.87
	2	Clear	6.7	96.05
	2 3 0	Clear	6.6	95.33
F <sub>9</sub>	0	Clear	7.1	99.96
	1	Clear	7.1	97.12
	2	Clear	7.1	96.43
	3	Clear	7.0	95.65
Commercial	0	Clear	6.8	99.90
gel	1 2	Clear	6.7	98.70
	2	Clear	6.6	97.20
	3	Clear	6.5	95.90

#### 7. DISCUSSION

The aim of this study was to develop suitable topical gel formulations of diclofenac sodium gel using HPMC K100M and Carbopol 934P as a gelling agents and glycerin as permeation enhancer. The apparent viscosity of the test formulations was comparable to that of the reference standard. Due to the viscous and hydrophilic nature of HPMC K100M and Carbopol 934P, the complex might have expanded more in water than the individual polymers would in water and hence the increase in solution

viscosity. The use of glycerin as permeation enhancers significantly increased drug release rate. It is generally agreed that in vitro drug release data for topical formulation cannot be used to accurately predict permeation across the skin, due to the barrier properties of stratum corneum which can be altered by the presence of various permeation enhancers in the formulation. The anti-inflammatory difference between the activities of the reference standard and test gels after 1 and 2 h was not significant, but after 3 and 4 h, the test gel exhibited significantly higher activity than that of standard formulation [25]

Table No. 5: Formulation batches from F1 toF3 Gel with Hydroxy propyl Methyl Cellulose

Name of Ingredients	Formula	tion	
	F1	F2	F3
Diclofenac	1	1	1
Linseed oil	3	3	3
Ginger oleoresin	0.5	1	1.5
Methyl salicylate	10	10	10
Propylene glycol	5	5	5
HPMC	1	1.5	2
Methyl paraben	0.1	0.1	0.1
Propyl paraben	0.03	0.03	0.03
Menthol	5	5	5
Isopropyl alcohol	6.5	6.5	6.5
Water q.s. to make	100	100	100

\*All the quantities are in mg.

Table No. 6 Formulation batches from F4 to F7 Gel with Carbopol

	Components	F <sub>4</sub>	F <sub>5</sub>	F <sub>6</sub>	F <sub>7</sub>
Drug	Diclofenac (%)	2	2	2	2
Excipient	Propylene Glycol (%)	10	10	10	10
	Methyl Paraben(%)	0.15	0.15	0.15	0.15
	Propyl Paraben (%)	0.05	0.05	0.05	0.05
	Carbopol 940 (%)	1.5	1.5	1.5	1.5
	Triethanolamine (%)	2	2	2	2
Vehicle	Ethanol (%)	0	16.86	33.72	67.4
	Purified water (%)	74.4	57.28	41.68	9
	oleoresin	1.4	1.4	1.4	1.4
	Linseed oil	8.5	8.5	8.5	8.5

Table No. 7 Organoleptic Characterization of formulation Batches gels

Formulation	Color	Clogging	Viscosity change	Feel
F <sub>1</sub>	Clear	)3 <del></del> 1	Not	Smooth
F <sub>2</sub>	Clear	3.70	Changed	Smooth
	Clear	-	Not	Smooth
$F_3$	Clear		Changed	Smooth
$F_4$	Clear	(C=)	Not	Smooth
F <sub>5</sub>	Clear	15=	Changed	Smooth
F <sub>6</sub>			Not	
1 6			Changed	
			Not	
			Changed	
			Not	
			Changed	

Table No. 8: Skin Irritation and Apperance test

Formulation	Appearance	Skin irritation
F1	Cream	0
F2	Cream	0
F3	Cream	0
F4	White	0
F5	White	0
F6	White	0
F7	White	0
M1	White	0

Table No. 9 Drug Content of formulation Batches gels

Sr. No.	Formulations	3.0
1,	F <sub>1</sub>	97.28
2.	F <sub>2</sub>	96.67
3.	F <sub>3</sub>	98.18
4.	$F_4$	98.18
5.	F <sub>5</sub>	99.06
5. 6.	F <sub>6</sub>	99.69
7	F <sub>7</sub>	99.80

Table No. 10 pH of gels

Sr. No.	Formulation	PH Mean ± S. D
1.	Fi	7.36±0.012
2.	$F_2$	$7.35 \pm 0.030$
3.	F <sub>3</sub>	$7.29 \pm 0.017$
4.	$F_4$	$7.41 \pm 0.020$
5.	F <sub>5</sub>	$7.30 \pm 0.046$
5. 6.	$F_6$	$7.22 \pm 0.023$
7.	F7	$6.84 \pm 0.021$
8.	Marketed	$5.86 \pm 0.03$

Table No. 11 Viscosity of various formulations at fixed time

S. No.	Formulations	Viscosity	
1.	F1	30600	
2.	F2	31200	
3.	F3	32000	
4.	F4	40400	
5.	F5	38800	
6.	F6	32000	
7.	F7	27000	

Table No. 12 Viscosity observation of various formulations

Formulations		RPM					
13		0.3 0.		1.5	3	6	12
Ala	Ascending	16.75	21.20	27.70	36.0	49.70	66.80
	Descending	11.50	14.50	21.70	30.70	46.40	66.90
Alb	Ascending	17.20	21.50	30.10	37.70	50.0	67.0
	Descending	11.10	15.20	22.20	31.10	47.0	67.0
Alc	Ascending	18.10	22.70	31.10	38.0	50.60	70.0
	Descending	11.20	16.10	22.70	32.5	47.50	67.50
A2a	Ascending	20.2	28.20	34.40	45.50	62.30	74.40
	Descending	14.90.	18.40	26.80	34.40	52.20	73.20
A2b	Ascending	19.4	24.50	32,60	42.40	61.80	70.70
	Descending	11.20	18.20	25.80	32.40	52.00	69.80
A2c	Ascending	16.30	22.10	29.40	35.0	48.10	65.50
	Descending	11.20	16.10	21.40	30.40	45.50	64.70
A3a	Ascending	14.40	20.70	28.10	35.70	46.20	64.20
	Descending	10.0	15.10	20.70	29.80	41.10	63.10

Table No. 13 Spreadability of Various formulations

Formulations	Time (sec)	Spreadability in time			
F1	10	40.68+0.698			
F2	10	44.74+0.964			
F3	11	47.30+0.964			
F4	12	48.46=1.735			
F5	13	51.50+1.114			
F6	14	52.84+1.206			
F7	12	54.99+1.304			
Marketed	08	65.71+0.894			



Table No. 14 Extrudability of various formulations

Formulations	Wt. Required (in gms) to extrude ribbon of 0.5 cm in 10 sec				
F1	580				
F2	565				
F3	560				
F4	553				
F5	500				
F6	475				
F7	490				
Marketed	502				

Table 15: Cumulative % Drug Release Profile of Diclofenac Gels.

Time	Formulation								
(Hr)	F1	F2	F3	F4	F5	F6	F7	Marketed	
1	14.36	18.5	8.78	18.37	18.52	15.72	31.40	20.83	
2	17.13	25.19	20.78	22.74	24.96	18.60	36.40	25.80	
3	21.69	31.47	29.07	31.43	31.78	25.33	44.27	36.41	
4	28.01	37.05	37.66	36.18	38.59	30.43	51.37	43.92	
5	32.99	43.86	43.47	40.68	42.03	36.71	59.09	49.58	
6	41.07	49.57	52.32	46.65	50.98	42.59	67.67	57.71	
7	47.04	56.86	60.41	59.09	57.87	50.35	71.34	65.58	
8	52.00	62.81	67.06	63.12	60.71	58.70	73.69	75.01	

#### **CONCLUSION**

- Formulation batch F7 (2% Diclofenac, 10%PG, 0.15%Methylparaben, 0.05%Propyl Paraben,1.5%Carbopol 940, 67.44%ethanol,16.86%water) had shown higher amount of percent cumulative release as compared to marketed gel(Intas pharmaceutical).
- Spreadiability of formulation F7 was good to cover the painful area.
- In formulation F7 the ethanol in 67.44% concentration had shown the higher penetration enhancer activity which result in higher drug release, flux and permeatability value.
- On the basis of organoleptic characteristic the improved patient acceptability was achieved through formulation F7.
- The improved patient convenience might thus be obtained by the administration of such a

dosage form with minimal blood level fluctuations.

The release penetration enhancer and other materials are cheap, readily available, safe, having wide regulatory acceptance and easy to handle for economic point of view. It may beneficial to adopt such simple technology for the commercial production of Diclofenac gel. The future scope of this study is that formulation should be subjected for long-term stability and in-vivo performance study and anti-inflammatory activity.

#### REFERENCES

- 1. Daniels R, Knie U. Galenics of dermal products vehicles, properties and drug release. JDDG 2007; 5: 367-381.
- 2. Babar A, Solanki UD, Cutie AJ, Plakogiannis F. Piroxicam release from dermatological bases: in vitro studies using cellulose



- membrane and hairless mouse skin. Drug Dev Ind Pharm 1990; 16: 523-540.
- 3. Busson MJ. Update on ibuprofen: review article. J Int Med Res 1986; 14: 53-62.
- 4. Babar A, Solanki UD, Cutie AJ, Plakogiannis F. Piroxicam release from dermatological bases: in vitro studies using cellulose membrane and hairless mouse skin. Drug Dev Ind Pharm 1990; 16: 523-540.
- 5. Brown MB, Hanpanitcharoen M, Martin GP. An in vitro investigation into the effect of glycosaminoglycans on the skin partitioning and deposition of NSAIDs. Int J Pharm 2001; 225: 113-121.
- 6. Chen H, Chang X, Du D, Li J, Xu H, Yang H. Microemulsion-based hydrogel formulation of ibuprofen for topical delivery. Int J Pharm 2006; 315: 52-58.
- 7. Brain KR, Green DM, Dykes PJ, Marks R, Bola TS. The role of menthol in skin penetration from topical formulations of ibuprofen 5% in vivo. Skin Pharmacology and Physiology 2005; 19: 17-21.
- 8. Park E, Chang S, Hahn M, Chi S. Enhancing effect of polyoxyethylene alkyl ethers on the skin permeation of ibuprofen. Int J Pharm 2000; 209: 109-119.
- 9. Siepmann J, Peppas NA. Modeling of drug release from delivery systems based on hydroxypropyl methylcellulose (HPMC). Adv Drug Deliv Rev, 2001; 48: 139-157.
- 10. Enayatifard R, Saeedi M, Akbari J, Haeri Tabatabaee Y. Effect of hydroxypropyl methylcellulose and ethyl cellulose content on release profile and kinetics of diltiazem HCl from matrices. Trop. J. Pharm. Res., 2009; 8: 425-432.
- 11. Awasthi S, Irshad M, Das MK, Ganti SS, Moshahid A. Rizvi. Anti-Inflammatory Activity of Calotropis gigantea and Tridax procumbens on CarrageeninInduced Paw

- Edema in Rats. Ethnobotanical Leaflets 2009; 13: 568-577.
- 12. Shivhare DU, Jain BK, Mathur BV, Bhusari PK, Roy AA. Formulation development and evaluation of diclofenac sodium gel using water soluble polyacrylamide polymer. Digest Journal of Nanomaterials and Biostructures 2009; 4(2): 285-290.
- 13. Kartin Moser, Katrin Kriwet, Aarti Naik, Yogeshwar N.Kalia, Richard H.Guy 2001, "Passive skin penetration enhancement and its qualification in vitro" European Journal of Pharmaceutics and Biopharmaceutics 52,103-112
- 14. Gupta A, Mishra KA, Singh KA. Formulation and evaluation of topical gel of diclofenac sodium using different polymers. Drug Invention Today 2010; 2(5): 250-253.
- 15. Bookya P, Jagadeesh I. Excipient screening and development of formulation design space for diclofenac sodium fast dissolving tablets. Int J Pharm Pharm Sci 2012; 4: 241-248.
- 16. Sera VV, Raman VN. In vitro skin absorption and drug release. The Indian Pharmacist 2006; 73: 356-360.
- 17. Induru J, Bookya P. Excipient screening and development of formulation design space for diclofenac sodium fast dissolving tablets. Int J Pharm Pharm Sci 2012; 4(1): 241-248.
- 18. Indian Pharmacopoeia. Ministry of Health and Family Welfare, New Delhi, India. 2007; 1: 148, 289, 1020.
- 19. Bazigha AK, Eman AF, Sahar FA, Heyam SS, Saeed KA. Development and evaluation of ibuprofen transdermal gel formulations. Trop J Pharm Res 2010; 9(4): 355-363.
- 20. Golinkin SH. Process for fracturing well formations using aqueous gels. US Patent No. US 4137182, 1979.
- 21. Shivhare DU, Jain BK, Mathur BV, Bhusari PK, Roy AA. Formulation development and evaluation of diclofenac sodium gel using



- water soluble polyacrylamide polymer. Digest Journal of Nanomaterials and Biostructures 2009; 4(2): 285-293.
- 22. Park E, Chang S, Hahn M, Chi S. Enhancing effect of polyoxyethylene alkyl ethers on the skin permeation of ibuprofen. Int J Pharm 2000; 209: 111-122.
- 23. Awasthi S, Irshad M, Das MK, Ganti SS, Moshahid A. Rizvi. Anti-Inflammatory Activity of Calotropis gigantea and Tridax procumbens on CarrageeninInduced Paw Edema in Rats. Ethnobotanical Leaflets 2009; 13: 585-597.
- 24. Sera VV, Raman VN. In vitro skin absorption and drug release. The Indian Pharmacist 2006; 73: 379-395.
- 25. Deshpande, S.G. and Shirolkar, S., J. Pharma. Pharmacol., 49, 1989
- 26. Bidkar sanjay,jain devendra, padsalg amol, patel kishna and mokale vinod, Apriljun2007, "Formulation development and fluconazole evaluation of in various polymeBased formulation development evaluation of fluconazole gel in various bases,"Asian journal of polymer pharmaceutics, volume 1, issue 1, 63-68
- 27. Table 1. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:3.
- 28. Table 2. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:4.
- 29. Table 3. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:5.

- 30. Table 4. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:5.
- 31. Table 5. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1 , Kapil R Bare1 , Venkatesh B Parik1 , Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:5.
- 32. Table 6. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1 , Kapil R Bare1 , Venkatesh B Parik1 , Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:5.
- 33. Table 7. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:5.
- 34. Table 8. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1 , Kapil R Bare1 , Venkatesh B Parik1 , Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:5.
- 35. Table 9. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:6.
- 36. Table 10. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:6.
- 37. Table 11. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1 , Kapil R Bare1 , Venkatesh B Parik1 , Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:6.
- 38. Table 12. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1,



- Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:7.
- 39. Table 13. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1 , Kapil R Bare1 , Venkatesh B Parik1 , Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:8.
- 40. Table 14. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:8.
- 41. Table 15. From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:10.
- 42. Fig 1: https://images.app.goo.gl/jMkMGDZghcBbt Kz17
- 43. Fig 2: https://images.app.goo.gl/epaRkAJ4eKP59r Qf8
- 44. Fig 3: https://images.app.goo.gl/mmHBjUmqXJnzS vJQ8
- 45. Fig 4: From ENKELEJDA GOCI\*, ENTELA HALOCI, SKERDILAID XHULAJ,

- LEDJAN MALAJ, International Journal of Pharmacy and Pharmaceutical Sciences ISSN-0975-1491 Vol 6, Issue 6, 2014; 3.
- 46. Fig 5: From ENKELEJDA GOCI\*, ENTELA HALOCI, SKERDILAID XHULAJ, LEDJAN MALAJ, International Journal of Pharmacy and Pharmaceutical Sciences ISSN-0975-1491 Vol 6, Issue 6, 2014; 3.
- 47. Fig 6: From ENKELEJDA GOCI\*, ENTELA HALOCI, SKERDILAID XHULAJ, LEDJAN MALAJ, International Journal of Pharmacy and Pharmaceutical Sciences ISSN-0975-1491 Vol 6, Issue 6, 2014; 4.
- 48. Fig 7: From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1 , Kapil R Bare1 , Venkatesh B Parik1 , Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:4.
- 49. Fig 8: From Dheeraj T Baviskar1\*, Yogeshkumar A Biranwar1, Kapil R Bare1, Venkatesh B Parik1, Mangesh K Sapate1 and Dinesh K Jain2, Tropical Journal of Pharmaceutical Research August 2013; 12:5.

HOW TO CITE: Nand Lal Thakur, Sana Nusrat Praween, In-Vitro Evaluation of Diclofenac Sodium Gel, Int. J. of Pharm. Sci., 2025, Vol 3, Issue 9, 3667-3684. https://doi.org/10.5281/zenodo.17235413