



**INTERNATIONAL JOURNAL OF
PHARMACEUTICAL SCIENCES**
[ISSN: 0975-4725; CODEN(USA): IJPS00]
Journal Homepage: <https://www.ijpsjournal.com>



Review Article

Nanotechnology –Based Drug Delivery System

Onkar Kalokhe*, Sameer Bhilare, Sangram Bandal, Sourav Hagavane, Shital Rokde, Dr. Kishorotari

Navsahyadri Institute of Pharmacy, Pune-412213, Maharashtra India.

ARTICLE INFO

Published: 01 June 2025

Keywords:

Nanoparticles, Design of Nanotechnology, Nanoparticles Medicated delivery of siRNA, Nano system in inflammation

DOI:

10.5281/zenodo.15569146

ABSTRACT

Nanoparticles having a huge potential for use as an effective medicine delivery system. In this post, we discussed recent developments in nanotechnology-based drug delivery. Nanotechnology has gained interest recently as a potential remedy for medical and gene delivery problems. Nanosystems with different compositions and biological properties have been thoroughly studied for the transportation of genes and medicines. Achieving effective drug delivery requires an understanding of how nanomaterials interact with the biological environment, including how they target cell-surface receptors, release drugs, administer multiple drugs, stabilize therapeutic agents, and use molecular mechanisms of cell signaling that are involved in the pathobiology of the disease in question. Numerous anti-cancer drugs, including doxorubicin, 5-fluorouracil, dexamethasone, and paclitaxel, have been efficiently formulated using nanomaterials. Furthermore, chitosan, quantum dots, polylactic/glycolic acid (PLGA), and PLGA-based nanoparticles have all been employed for the delivery of RNAi in vitro. Brain cancer is one of the most difficult tumors to detect and treat since imaging and treatment tools cannot penetrate the blood-brain barrier and reach the brain. Nanomaterial-attached anti-cancer drugs, such as doxorubicin and loperamide, have been shown to be able to penetrate the unaltered blood-brain barrier and reach the brain at therapeutic dosages.

INTRODUCTION

Because cells absorb nanoparticles more effectively than larger macromolecules, these particles could serve as efficient systems for delivering and transporting drugs. Medications can be attached to the surface of the particles or

embedded within their matrix for therapeutic use. The trajectory of a drug entering a biological system should be guided by a drug targeting mechanism. Various nanosystems with distinct compositions and biological properties have been extensively researched for drug and gene delivery purposes. It is essential to design nanosystems

***Corresponding Author:** Onkar Kalokhe

Address: Navsahyadri Institute of Pharmacy, Pune-412213, Maharashtra India.

Email ✉: onkarkalokhe201@gmail.com

Relevant conflicts of interest/financial disclosures: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.



logically, taking into account their interactions with the biological environment, targeted cell populations, cell-surface receptors on the target cells, changes in these receptors associated with disease progression, the mechanism and site of drug action, and the retention of the drug. Administration Understanding the molecular mechanisms and pathobiology of the disease in question would be an effective approach to facilitate efficient drug delivery. The destiny of a drug entering the biological environment should be managed by a targeted drug delivery system. Nanosystems with diverse compositions and biological properties have been extensively researched for drug and gene delivery applications. The development of nanosystems should be methodically informed by their interactions with the biological environment, the specific cell populations they aim to target, the receptors on target cells, the alterations in cell receptors associated with disease progression, the mechanisms and sites of drug action, drug retention, the administration of multiple drugs, as well as the molecular mechanisms and pathobiology of the relevant disease. Understanding the barriers to medication, such as the stability of therapeutic agents in living cells, is essential. Reduced drug effectiveness may occur due to the drug's inability to penetrate the cell, its lack of availability due to various delivery molecule characteristics, changes in the genetic makeup of cell-surface receptors, overexpression of efflux pumps, alterations in signaling pathways as the disease advances, or degradation of the drug. For instance, doxorubicin and cisplatin, which are two anti-cancer medications, are ineffective because of heightened DNA methylation that

happens as cancer develops. This review encompasses the drug delivery aspects of nanomedicine, including how nanoparticles engage with cell-surface receptors, the biological responses and cellular signaling involved, as well as the research necessary for the widespread implementation of nanodelivery systems in healthcare. This review encompasses the drug delivery aspects of nanomedicine, including how nanoparticles engage with cell-surface receptors, the biological responses and cellular signaling involved, as well as the research necessary for the widespread implementation of nanodelivery systems in healthcare

Design Of Nanotechnology-Based Drug Delivery System

The utilization of nanoparticles in targeted drug delivery can improve drug bioavailability, assist in drug targeting, and facilitate the absorption of poorly soluble drugs at the disease site [8,9]. Figure 1 illustrates a schematic comparison between targeted and untargeted drug delivery systems. Numerous anti-cancer drugs, including doxorubicin [12], 5-fluorouracil [13], paclitaxel [10,11], and dexamethasone [14], have been effectively formulated using nanomaterials.

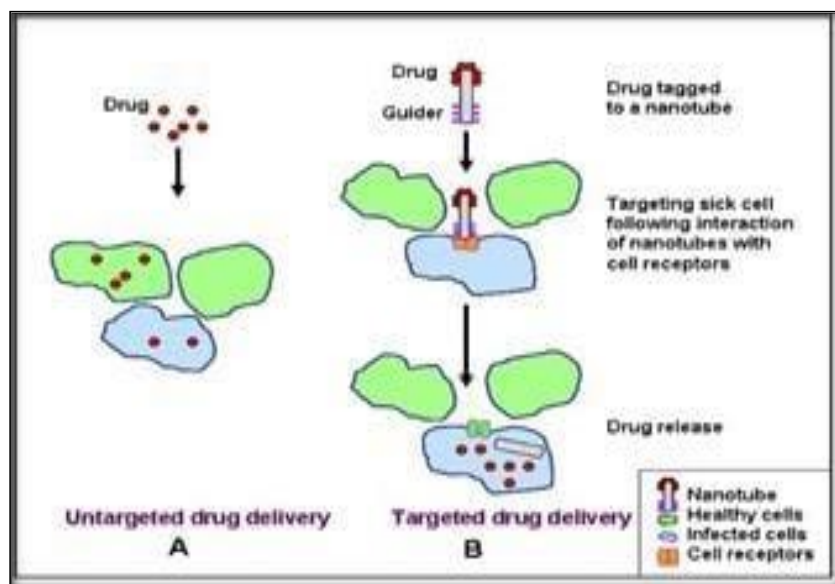


Figure No. 1

Dexamethasone, a glucocorticoid with an intracellular site of action, has been encapsulated in polylactic/glycolic acid (PLGA) and polylactic acid (PLA) based nanoparticles. One chemotherapy drug with anti-inflammatory and anti-proliferative properties is dexamethasone. Following the drug's binding to the cytoplasmic receptors, the drug-receptor complex is carried to the nucleus, where it causes the production of specific genes that regulate cell division. The application of colloidal drug delivery methods, such as liposomes, micelles, or nanoparticles, in cancer treatment has been thoroughly studied. Drug delivery systems are effective because of their compact size, decreased toxicity, regulated drug release time, and adjustments to the drug's pharmacokinetics and biological distribution. Chemotherapy fails to cure cancer far too frequently because certain tumour cells become resistant to several anticancer medications.

Nanoparticles Medicated Delivery of Sirna

Short interfering RNA (siRNA) has several applications and is emerging as a potent technique

for controlling gene expression. Clinical studies of nucleic acid-based therapy will not be possible until significant advancements in the delivery mechanism have been made. Quantum dots (QD) have been demonstrated to deliver RNAi. [16] PLA and PLGA-based nanoparticles have also been used to deliver RNAi in vitro. [17] Even though siRNA delivery using various nanomaterials has shown some success, it is difficult to monitor their distribution and gauge the efficacy of transfection in the absence of a suitable monitoring agent or marker. Creating a transfection agent that is both efficient and self-tracking for RNA interference is a challenging task. Tan and others. [18] siRNA to the human epidermal growth factor receptor-2 (HER2/neu) using recently developed quantum dot-encapsulated chitosan nanoparticles. Because the chitosan nanoparticles contain fluorescent QDs, this novel nano carrier aids in siRNA monitoring. Chitosan/quantum dot nanoparticles surface-labeled with HER2 antibody, which targets the SKBR3 cells' HER2 receptors, have been used to deliver HER2 siRNA specifically to HER2-overexpressing SKBR3 breast cancer cells.

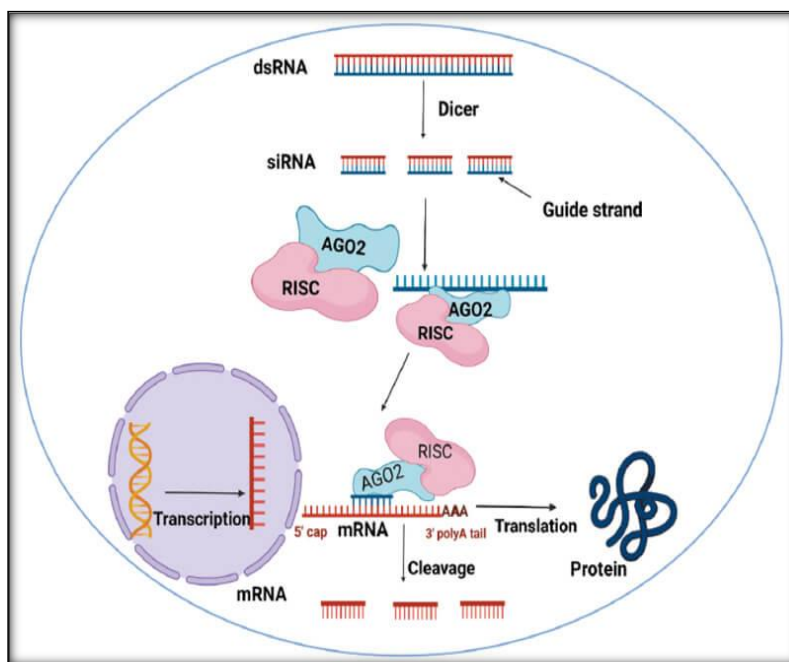


Figure No.2

Labelling nanoparticles with a fluorescent marker, like Cy-5, makes it easier to see the uptake and accumulation of nanotubes under a fluorescent microscope. Howard et al. recently found that K562 (Ph(+)) cells expressed 90% less BCR/ABL-1 leukaemia fusion protein when these nanoparticles were combined with siRNA specific to the BCR/ABL-1 junction sequence. The bronchiolar epithelial cells of transgenic EGFP mice also demonstrated efficient *in vivo* RNA interference after nasal administration of chitosan/siRNA formulations. These findings show the potential applications of this novel chitosan-based strategy in RNA-mediated therapy of systemic and mucosal disorders.

Cancer

Targeting cancer cells with nanoparticles

Cancer is now one of the most challenging diseases to diagnose and treat, and brain cancer is especially challenging since it is challenging to get imaging and therapy agents into the brain through the blood-brain barrier. Several researchers have

found that these chemicals could be delivered into the brain by nanoparticles. [20–22] Apolipoprotein E has been suggested as the mediator of drug transport across the blood-brain barrier. [23] Loperamide, which does not cross the blood-brain barrier but has antinociceptive effects when injected directly into the brain, was loaded onto human serum albumin nanoparticles. After then, it was linked to apolipoprotein E. Mice administered this substance intravenously demonstrated antinociceptive effects in the tail-flick test. The efficiency of this drug delivery method. While one agent may help visualise the target through magnetic resonance imaging, a third agent attached to the PEBBLE may deliver a deadly dose of medication or toxin to nearby cancer cells. All three roles can be combined into a single, small polymer sphere to generate a potent cancer-fighting tool. Another anti-cancer drug, doxorubicin, can penetrate the intact blood-brain barrier and reach the brain at therapeutic concentrations when it is affixed to polysorbate-coated nanoparticles [24, 25]. Compared to current methods, smart superparamagnetic iron oxide particle conjugates can target and locate

brain tumours earlier and more precisely. Folic acid and polyethylene glycol are known to further enhance intracellular absorption.

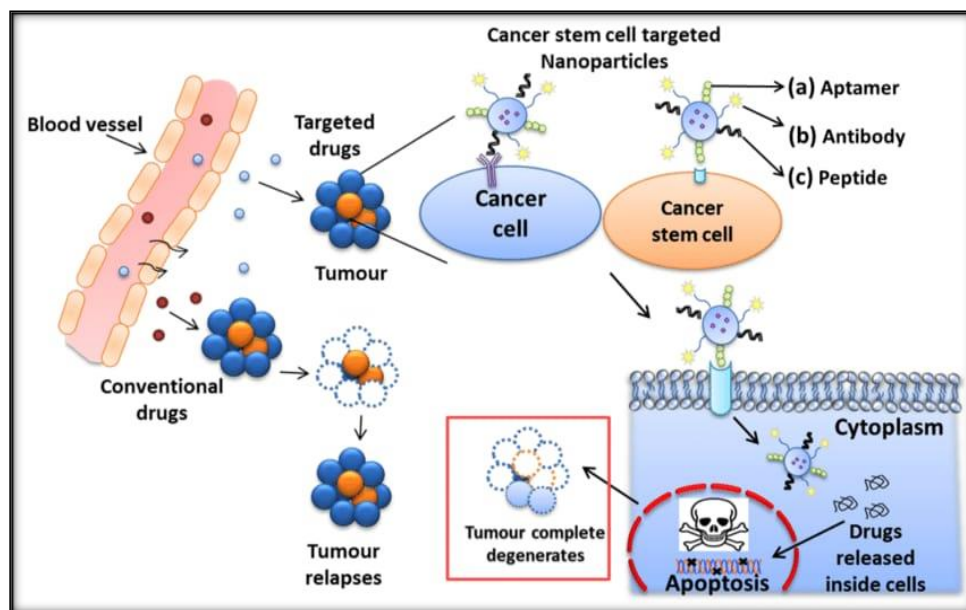


Figure No.3

Targeting angiogenesis with nanoparticles

Strong angiogenesis causes tumours to grow rapidly. Thus, one way to stop angiogenesis is to start tumor cells. A complex network of mediators controls angiogenesis, and new research indicates that integrin $\alpha\beta3$ and vascular endothelial growth factors (VEGFs) are key regulators. Thus, a unique anti-angiogenesis approach for treating a broad range of solid tumours is the selective targeting of VEGFs and $\alpha\beta3$ integrin. Coating nanoparticles with peptides that selectively bind to the $\alpha\beta3$ integrin and the VEGF receptor is one method. [26] The Arg-Gly-Asp (RGD) sequence of the synthetic peptide is known to bind especially to the $\alpha\beta3$ integrin expressed on endothelial cells in angiogenesis blood arteries, potentially limiting the growth and proliferation of tumours. The blood

vessels and angiogenic tissue that surround the tumor tissue may be traced or removed with the aid of these FITC-GRGDS-[27] loaded nanotubes. Our research team has been investigating the biological effects of RGDSK self-assembling rosette nanotubes (RGDSK-RNT). These rosette nanotubes are a novel kind of nanotubes with biological inspiration and inherent water solubility during manufacturing [28, 29]. These nanotubes are constructed from the guanine-cytosine motif. However, the ability of the RNT to adopt various functional groups at the G/C motif is one of its distinctive features, providing the nanotubes with functional versatility for certain biological or therapeutic uses. Therefore, in order to treat cancer, it might be conceivable to modify RNTs to target various therapeutic molecules in vivo.

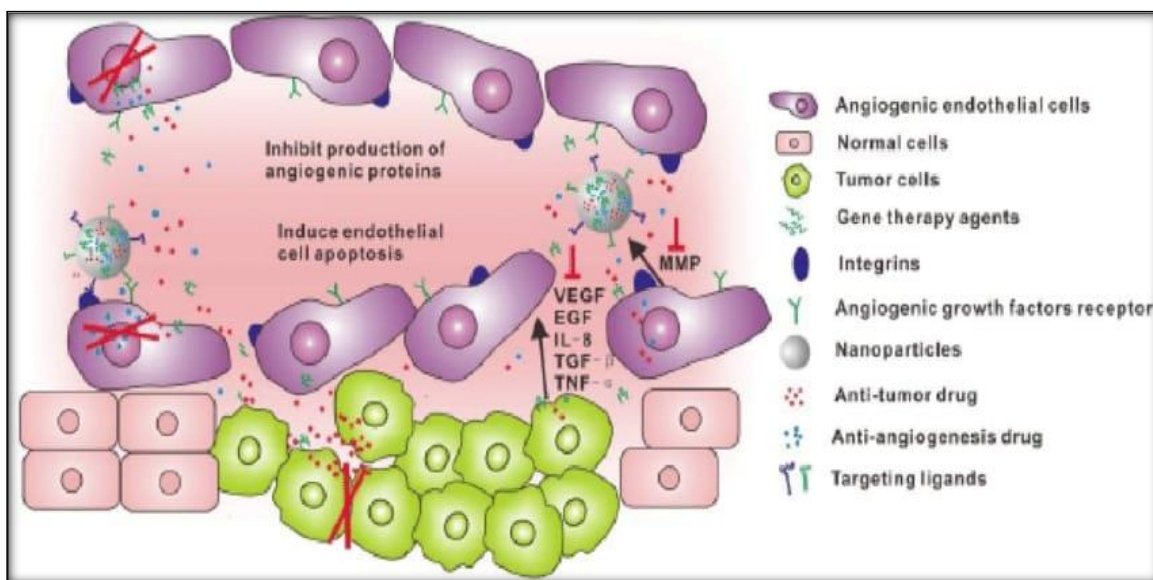


Figure No.4

Nanosystem In Inflammation

Targeting macrophages to control inflammation

It makes sense to use nanoparticles for macrophage-specific targeting because of the rapidity with which macrophages can recognise and destroy foreign particles. Because macrophages release a diverse array of inflammatory mediators, they can regulate inflammation in a number of illnesses. Consequently, macrophages could be therapeutic targets for a range of human and animal diseases. Although most infections can be eliminated by macrophages, several germs, such as *Toxoplasma gondii*, *Leishmania* sp., *Mycobacterium tuberculosis*, and *Listeria monocytogenes*, have developed the potential ability to withstand the phagocytosis action of macrophages. These infections infiltrate altered lysosomes and interfere with the macrophage's ability to eliminate them by disrupting its molecular machinery. Therefore, it may be helpful to remove cellular reservoirs by delivering antimicrobial agent(s) via nanoparticles into pathogen-containing intracellular vacuoles in macrophages. [30,31] This technique can be

utilised to reduce pro-inflammatory cytokine production, decrease adverse medication administration side effects, and increase therapeutic drug concentrations in the vacuoles of infected macrophages. Antileishmanial medications have been delivered to macrophages using polyalkylcyanoacrylates (PACA) nanoparticles. Macrophages did not secrete interleukin-1 in response to this nanomaterial. [32] Therefore, targeting macrophage infections in chronic disorders may benefit greatly from similarly constructed nanosystems. Amphotericin B, an antifungal and antileishmanial medication, has been complexed with lipid-based nanotubes to produce a less toxic form of AmB. Gupta and Vyas developed AmB in trilaurin-based nanosize lipid particles (emulsomes) stabilised by soy phosphatidylcholine as a unique intravenous drug delivery technique for macrophage targeting. Nanocarrier-mediated delivery of macrophage toxins has been shown to be an effective way to eradicate unwanted macrophages in gene therapy and other clinically relevant conditions such as autoimmune blood disorders, T cell-mediated autoimmune diabetes, rheumatoid arthritis, spinal cord injury, sciatic nerve injury, and restenosis.

after angioplasty. Another option is to use nanoparticles that have the capacity to destroy macrophages. Using a variety of macrophage cell receptors as therapeutic targets may be more successful.

Targeting Inflammatory Molecules

Over the past 20 years, a large number of cell adhesion molecules have been discovered. Cell adhesion molecules are glycoproteins found on the cell surface that act as receptors for attachment between cells and between cells and the extracellular matrix. [34–35] These cell adhesion molecules fall into four classes: integrins, cadherins, selectins, and the immunoglobulin superfamily. Both the effective migration of inflammatory cells, such as neutrophils and monocytes, into inflamed tissues and the generation of the host response to infections depend on these substances. However, there is compelling evidence that excessive neutrophil migration results in disproportionate tissue death and damage in inflammatory lungs. Consequently, there is a great deal of work being done to precisely regulate neutrophil migration into inflammatory tissues. Recent advances in our knowledge of cell adhesion molecules have influenced the design and development of drugs (such as peptides and proteins) for the potential treatment of autoimmune illnesses, cancer, and heart disease. [36–40] These compounds are important in thrombosis, cancer and autoimmune diseases such type-1 diabetes [41–45]. RGD peptides have been shown to target integrins $\alpha\beta3$ and $\alpha\beta5$, whereas peptides generated from the intercellular adhesion molecule-1 (ICAM-1) have been shown to target the $\alpha\beta2$ integrin. Peptides derived from $\alpha\beta2$ can target cells that express ICAM-1. Cyclic RGD peptides have been added to doxorubicin (Dox-RGD4C) and paclitaxel (PTX-RGD) to enhance their targeted distribution. Mice

with human breast cancer cells (i.e., MDA-MB-435) survived the disease when treated with Dox-RGD4C, while all untreated control mice died [46]. This substance targets the tumour vasculature's integrins $\alpha\beta3$ and $\alpha\beta5$ during angiogenesis. Extracellular regulated kinases (ERK) may govern apoptosis and cell survival through increased $\text{TNF-}\alpha$ [47] expression, decreased Akt activity, increased caspase-3 and caspase-8 activities, and -1 is upregulated during tissue inflammation and several different types of cancer, this combination may be useful for directing drugs to tumour and inflammatory cells. MTX has an anti-inflammatory effect because it inhibits the production of anti-inflammatory cytokines such as interleukin-6 (IL-6) and interleukin-8 (IL-8). Thus, the MTX-cLABLE conjugate's capacity enhanced p53 and BAX activity. Our group is investigating the migration of neutrophils in cows and horses, the role of P38 kinases in cell signalling in human lung epithelial cells, and the connection between RGD-RNT and $\alpha\beta3$ integrins. Cyclo(1,12)PenITDGEATDSGC peptide (cLABLE peptide), found in the I-domain of the α subunit of Leukocyte Function-Associated Factor-1 (LFA-1), is known to bind ICAM-1. The cLABLE peptide and methotrexate (MTX) have been joined to create the MTX-cLABLE conjugate [48]. Given that ICAM-1 suppresses these cytokine production in $\text{TNF-}\alpha$ -activated human coronary artery endothelial cells was contrasted with that of MTX alone. More specifically than IL-8, MTX-cLABLE inhibits the synthesis of IL-6. Further understanding of the process or mechanisms of internalisation and intracellular trafficking of cell adhesion molecules is required in order to use them to deliver medication molecules to a specific cell type or for the diagnosis of cancer and other disorders (such as cardiac and autoimmune diseases).

CONCLUSION



In cancer and inflammation, nanomedicine delivery technologies appear to have great potential for overcoming some of the barriers to efficient cell and molecular targeting. Furthermore, there is a significant opportunity to address drug resistance in target cells and assist drugs in overcoming obstacles such as those found in the brain. The chemicals are solely expressed in the targeted organs to prevent harm to healthy tissues. Second, it's important to understand how the drugs behave once they enter the nucleus and other sensitive cell organelles. Additionally, because nanosystems increase the effectiveness of drug delivery, doses might need to be adjusted.

REFERENCES

1. Pison U, Welte T, Giersing M, Groneberg DA. Nanomedicine for respiratory diseases. *Eur J Pharmacol.* 2006;533:341–50.
2. Brannon-Peppas L, Blanchette JQ. Nanoparticle and targeted systems for cancer therapy. *Adv Drug Deliv Rev.* 2004;56(11):1649–59.
3. Stylios GK, Giannoudis PV, Wan T. Applications of nanotechnologies in medical practice. *Injury.* 2005;36(Suppl 4):S6–13.
4. Yokoyama M. Drug targeting with nano-sized carrier systems. *J Artif Organs.* 2005;8(2):77–84.
5. Schatzlein AG. Delivering cancer stem cell therapies – a role for nanomedicines? *Eur J Cancer.* 2006;42(9):1309–15.
6. Groneberg DA, Rabe KF, Fischer A. Novel concepts of neuropeptide-based therapy: Vasoactive intestinal polypeptide and its receptors. *Eur J Pharmacol.* 2006;533:182–94.
7. Grady WM. Epigenetic events in the colorectum and in colon cancer. *Biochem Soc Trans.* 2005;33:684–8.
8. Ould-Ouali L, Noppe M, Langlois X, Willems B, TeRiele P, Timmerman P, et al. Self-assembling PEG-p(CL-co-TMC) copolymers for oral delivery of poorly water-soluble drugs: a case study with risperidone. *J Control Release.* 2005;102(3):657–68.
9. Kipp JE. The role of solid nanoparticle technology in the parenteral delivery of poorly water-soluble drugs. *Int J Pharm.* 2004;284(1–2):109–22.
10. Fonseca C, Simoes S, Gaspar R. Paclitaxel-loaded PLGA nanoparticles: preparation, physicochemical characterization and in vitro anti-tumoral activity. *J Control Release.* 2002;83(2):273–86.
11. Koziara JM, Whisman TR, Tseng MT, Mumper RJ. In-vivo efficacy of novel paclitaxel nanoparticles in paclitaxel-resistant human tumors.
12. Yoo HS, Lee KH, Oh JE, Park TG. In vitro and in vivo anti-tumor activities of nanoparticles based on doxorubicin-PLGA conjugates. *J Control Release.* 2000;68(3):419–31.
13. Bhadra D, Bhadra S, Jain S, Jain NK. A PEGylated dendritic nanoparticulate carrier of fluorouracil. *Int J Pharm.* 2003;257(1–2):111–24.
14. Panyam J, Labhasetwar V. Sustained cytoplasmic delivery of drugs with intracellular receptors using biodegradable nanoparticles. *Mol Pharm.* 2004;1(1):77–84.
15. Koziara JM, Lockman PR, Allen DD, Mumper RJ. Paclitaxel nanoparticles for the potential treatment of brain tumors. *J Control Release.* 2004;99(2):259–69.
16. Chen AA, Derfus AM, Khetani SR, Bhatia SN. Quantum dots to monitor RNAi delivery and improve gene silencing. *Nucleic Acids Res.* 2005;33(22):e190.
17. Shinde RR, Bachmann MH, Wang Q, Kasper R, Contag CH. PEG-PLA/PLGA nanoparticles for in-vivo RNAi delivery. *NSTI Nanotech.* 2007.
18. Tan WB, Jiang S, Zhang Y. Quantum-dot based nanoparticles for targeted silencing of



- HER2/neu gene via RNA interference. *Biomaterials*. 2007;28(8):1565–71.
19. Howard KA, Rahbek UL, Liu X, Damgaard CK, Glud SZ, Andersen MØ, et al. RNA interference in vitro and in vivo using a novel chitosan/siRNA nanoparticle system. *MolTher*. 2006;14(4):476–84.
20. Kreuter J, Shamenkov D, Petrov V, Ramge P, Cychutek K, Koch-Brandt C, et al. Apolipoprotein-mediated transport of nanoparticle-bound drugs across the blood-brain barrier. *J Drug Target*. 2002;10(4):317–25.
21. Costantino L, Gandolfi F, Tosi G, Rivasi F, Vandelli MA, Forni F. Peptide-derivatized biodegradable nanoparticles able to cross the blood-brain barrier. *J Control Release*. 2005;108(1):84–96.
22. Sumner JP, Kopelman R. Alexa Fluor 488 as an iron sensing molecule and its application in PEBBLE nanosensors. *Analyst*. 2005;130(4):528–33.
23. Michaelis K, Hoffmann MM, Dreis S, Herbert E, Alyautdin RN, Michaelis M, et al. Covalent linkage of apolipoprotein E to albumin nanoparticles strongly enhances drug transport into the brain. *J PharmacolExpTher*. 2006;317(3):1246–53.
24. Steiniger SC, Kreuter J, Khalansky AS, Skidan IN, Bobruskin AI, Smirnova ZS, et al. Chemotherapy of glioblastoma in rats using doxorubicin-loaded nanoparticles. *Int J Cancer*. 2004;109(5):759–67.
25. Zhang Y, Sun C, Kohler N, Zhang M. Self-assembled coatings on individual monodisperse magnetite nanoparticles for efficient intracellular uptake. *Biomed Microdevices*. 2004;6:33–40.
26. Li L, Wartchow CA, Danthi SN, Shen Z, Dechene N, Pease J, et al. A novel antiangiogenesis therapy using an integrin antagonist or anti-Flk-1 antibody coated 90Y-labeled nanoparticles. *Int J RadiatOncolBiol Phys*. 2004;58(4):1215–27.
27. Park JH, Kwon S, Nam JO, Park RW, Chung H, Seo SB, et al. Self-assembled nanoparticles based on glycol chitosan bearing 5beta-cholanic acid for RGD peptide delivery. *J Control Release*. 2004;95(3):579–88.
28. Fenniri H, Deng BL, Ribbe AE, Hallenga K, Jacob J, Thiyagarajan P. Entropically driven self-assembly of multichannel rosette nanotubes. *Proc Natl AcadSci U S A*. 2002;99:6487–92.
29. Fenniri H, Mathivanan P, Vidale KL, Sherman DM, Hallenga K, Wood KV, et al. Helical rosette nanotubes: design, self-assembly, and characterization. *J Am Chem Soc*. 2001;123:3854–5.
30. Zhang D, Tan T, Gao L, Zhao W, Wang P. Preparation of azithromycin nanosuspensions by high pressure homogenization and its physicochemical characteristics studies. *Drug Dev Ind Pharm*. 2007;33(5):569–75.
31. Gaspar R, Pr  at V, Oppendoes FR, Roland M. Macrophage activation by polymeric nanoparticles of polyalkylcyanoacrylates: activity against intracellular leishmaniadonovani associated with hydrogen peroxide production. *Pharm Res*. 1992;9(6):782–7.
32. Balland O, Pinto-Alphandary H, Viron A, Puvion E, Andreumont A, Couvreur P. Intracellular distribution of ampicillin in murine macrophages infected with *Salmonella typhimurium* and treated with (3H)ampicillin-loaded nanoparticles. *J AntimicrobChemother*. 1996;37:105–15.
33. Gupta S, Vyas SP. Development and characterization of amphotericin B bearing emulsomes for passive and active macrophage targeting. *J Drug Target*. 2007;15(3):206–17.

34. Hynes RO. A reevaluation of integrins as regulators of angiogenesis. *Nat Med.* 2002;8:918–21.
35. Hynes RO, Zhao Q. The evolution of cell adhesion. *J Cell Biol.* 2000;24:F89–95.
36. Chen X, Plasencia C, Hou Y, Neamati N. Synthesis and biological evaluation of dimeric RGD peptide-Paclitaxel conjugate as a model for integrin-targeted drug delivery. *J Med Chem.* 2005;48:1098–106.
37. Gupta AS, Huang G, Lestini BJ, Sagnella S, Kottke-Marchant K, Marchant RE. RGD modified liposomes targeted to activated platelets as a potential vascular drug delivery system. *ThrombHaemost.* 2005;93:106–14.
38. Schiffelers RM, Koning GA, ten Hagen TL, Fens MH, Schraa AJ, Janssen AP, et al. Anti-tumor efficacy of tumor vasculature-targeted liposomal doxorubicin. *J Control Release.* 2003;91:115–22.
39. Haass NK, Smalley KS, Li L, Herlyn M. Adhesion, migration and communication in melanocytes and melanoma. *Pigment Cell Res.* 2005;18:150–9.
40. Christofori G. Changing neighbours, changing behaviour: Cell adhesion molecule-mediated signalling during tumour progression. *EMBO J.* 2003;22:2318–30.
41. Pancioli AM, Brott TG. Therapeutic potential of platelet glycoprotein IIb/IIIa receptor antagonists in acute ischaemic stroke: Scientific rationale and available evidence. *CNS Drugs.* 2004;18:981–88.
42. Andrews RK, Berndt MC. Platelet physiology and thrombosis. *Thromb Res.* 2004;114:447–53.
43. Anderson ME, Siahaan TJ. Targeting ICAM-1/LFA-1 interaction for controlling autoimmune diseases: Designing peptide and small molecule inhibitors. *Peptides.* 2003;24:487–501.
44. Yusuf-Makagiansar H, Anderson ME, Yakovleva TV, Murray JS, Siahaan TJ. Inhibition of LFA-1/ICAM-1 and VLA-4/VCAM-1 as a therapeutic approach to inflammation and autoimmune diseases. *Med Res Rev.* 2002;22:146–67.
45. Shimaoka M, Springer TA. Therapeutic antagonists and the conformational regulation of the $\beta 2$ integrins. *Curr Top Med Chem.* 2004;4:1485–95.
46. Arap W, Pasqualini RR, Ruoslahti E. Cancer treatment by targeted drug delivery to tumor vasculature in a mouse model. *Science.* 1998;279:377–80.
47. Zhuang S, Schnellmann RG. A death-promoting role for extracellular signal-regulated kinase. *J PharmacolExpTher.* 2006;319:991–7.
48. Duneahoo AL, Anderson M, Majumdar S, Kobayashi N, Berkland C, Siahaan TJ. Cell adhesion molecules for targeted drug delivery. *J Pharm Sci.* 2006;95:1856–72.
49. Zhang N, Berkland C. Synthesis of PLGA nanoparticles with conjugated CLABL as targeted vascular delivery vehicles. *Science Talks*, Higuchi Biosciences Center, University of Kansas, Lawrence; 2006.

HOW TO CITE: Onkar Kalokhe*, Sameer Bhilare, Sangram Bandal, Sourav Hagavane, Shital Rokde, Dr. Kishorotari, Nanotechnology –Based Drug Delivery System, *Int. J. of Pharm. Sci.*, 2025, Vol 3, Issue 6, 46-55. <https://doi.org/10.5281/zenodo.15569146>

